

Assessing tumor microenvironment in rat glioma model using hyperpolarized ^{13}C MRSI with a sliding window

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Target Audience Researchers who are interested in characteristics of hyperpolarized ^{13}C -substrates in cancer.

Purpose *In vivo* characteristics of hyperpolarized ^{13}C -substrates are often critical in the design of optimized pulse sequence as well as in the proper data analysis. By far, T_1 has received most attention since the observation window of hyperpolarized ^{13}C MR signals are T_1 -limited. However, T_2 and T_2^* also provide useful to understand tissue characteristics and create unique image contrasts^[1-4], providing information for alternative MR acquisition strategies^[2,3,5]. In this study, we propose a method that measures *in vivo* T_2^* using a **sliding window spiral chemical shift imaging** and calculate (1) *in vivo* T_2^* maps of hyperpolarized ^{13}C -substrates and (2) B_0 maps from glioma-implanted rat.

Methods Approximately 10^6 glioma cells are implanted into the right striatum of male Wistar rats ~10 days prior to the ^{13}C imaging (N=5). A spiral chemical shift imaging (spCSI) pulse sequence^[6] is used to acquire single time-point metabolite maps of rat brain (four spatial interleaves, spectral width = 932.8 Hz, #echoes = 96, field of view = $43.5 \times 43.5 \text{ mm}^2$, nominal spatial resolution = $2.7 \times 2.7 \text{ mm}^2$, slice thickness = 6-8 mm, acquisition time = 0.5 s) following an *i.v.*

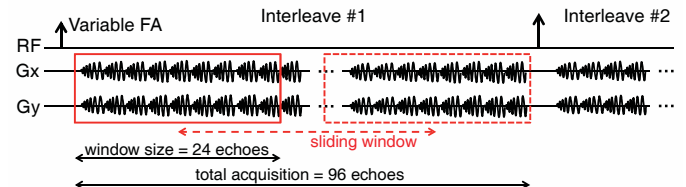


Fig.1. Schematics of sliding window spCSI for T_2^* measurement

injection of 125-mM hyperpolarized $[1-^{13}\text{C}]$ pyruvate (injection-to-scan = 25 s, polarized using HyperSense DNP). B_0 field inhomogeneity over the brain is minimized adjusting linear shim currents with ^1H PRESS sequence. Custom-made birdcage RF coil (^1H - ^{13}C dual-tuned) and high-performance insert gradient coil ($G_{\text{max}} = 500 \text{ mT/m}$, Slew rate_{max} = 1,865 mT/m/ms) are used in a 3T GE Signa clinical MR scanner. To assess T_2^* of individual ^{13}C -labeled metabolite, a sliding spectral window (window size = 24 echoes) with varying echo times ($\Delta\text{TE} = 1.07 \text{ ms}$) is applied to the acquired CSI data (**Fig.1**). T_2^* maps of $[1-^{13}\text{C}]$ pyruvate and $[1-^{13}\text{C}]$ lactate are calculated by fitting the temporal decay of each metabolite to an exponential function voxel-wise. For fitting, metabolite maps up to ~35 ms are used to maintain sufficient SNR.

Results and Discussion Longer $[1-^{13}\text{C}]$ lactate T_2^* s ($41.0 \pm 3.85 \text{ ms}$) were measured from glioma as compared to normal-appearing brain in the contralateral side ($31.6 \pm 5.3 \text{ ms}$, $P = 0.08$, **Figs.2-3**). T_2^* of $[1-^{13}\text{C}]$ pyruvate was also longer in glioma ($58.9 \pm 12.9 \text{ ms}$) than in normal-appearing brain ($37.2 \pm 6.8 \text{ ms}$, $P = 0.01$). The T_2^* map of ^{13}C -bicarbonate was not reliably measured due to the low SNR. Although T_2^* can be affected by other factors such as field inhomogeneity

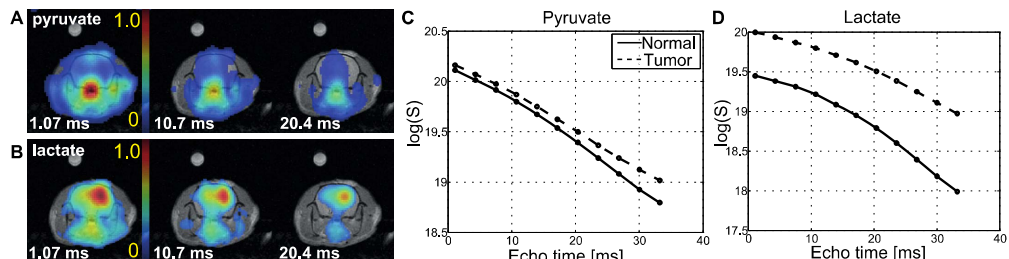


Fig.2. Metabolite maps of (A) $[1-^{13}\text{C}]$ pyruvate and (B) $[1-^{13}\text{C}]$ lactate reconstructed from a C6 glioma-bearing rat in right striatum using a sliding spectral window (24 echoes) at 1.07, 10.7, and 20.4 ms of echo times (TEs). Averaged signal decay of (C) pyruvate and (D) lactate in tumor (dashed line) and normal-appearing brain (solid line) ROIs.

and flow, the longer lactate T_2^* in tumor than in normal brain is consistent with longer lactate T_2 ^[1,2] in tumor. Off-resonance effects were detected in the tumor regions despite shimming over the entire brain ($|B_{0,\text{Tumor}} - B_{0,\text{NormalBrain}}| = 0.255 \pm 0.034 \text{ ppm}$, $P < 0.005$, **Fig.3D**). This observation is consistent with the magnetic susceptibility changes in tumor due to paramagnetic blood deposition and diamagnetic calcification around the tumor^[7-9]. Considering that the off-resonance effect shortens T_2^* , the contrasts between glioma and normal-appearing brain in T_2^* maps imply the significant contribution of T_2 differences. A non-exponential attenuation factor detected in lactate is probably due to the strong ^{13}C - ^1H spin-spin coupling ($J_{\text{CH}} \approx 25 \text{ ms}$)^[1,4] (**Fig.2C,D**).

Conclusion The proposed method provides the opportunity to assess complex transverse relaxation mechanism of hyperpolarized ^{13}C -substrates by observing multi-exponential decay patterns of the signal (as compared to the conventional spectral line-width measurement) as well as B_0 off-resonance. The study detected longer T_2^* in brain tumor than in normal appearing brain, reflecting T_2 rather than field inhomogeneity. Therefore, the measured T_2^* might serve as an indirect measure of relative T_2 of tumor and normal tissue.

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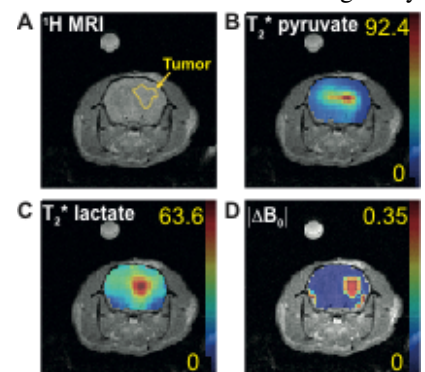


Fig.3. (A) ^1H MRI of tumor-bearing rat brain. Cerebral T_2^* maps of (B) $[1-^{13}\text{C}]$ pyruvate and (C) $[1-^{13}\text{C}]$ lactate. Numbers are in [ms]. (D) Field map indicates that B_0 field is -11 Hz (0.3 ppm) shifted in the tumor ROI.