

Simultaneous quantification of intravascular blood T₁ and T₂ with multiple-readout TRUST (mTRUST)

Zachary B Rodgers¹ and Felix W Wehrli¹

¹Radiology, University of Pennsylvania, Philadelphia, PA, United States

INTRODUCTION: Quantification of intravascular blood T₁ and T₂ has several important applications in quantitative MRI. Blood T₁ values are needed in order to accurately measure perfusion in arterial spin labeling (ASL)¹, to determine the blood inversion recovery null point in vascular-space occupancy (VASO)² MRI and black blood imaging techniques³, and can also be used for estimation of hematocrit (Hct)^{4,5,6}, the main protein constituent in blood. Quantification of blood T₂ can be used to estimate venous blood oxygen saturation (Y_v)⁷, for instance, using the T₂-relaxation-under-spin-tagging (TRUST) technique⁸. TRUST and other T₂-based Y_v quantification techniques require estimation of Hct in order to convert T₂ to Y_v. While Hct can be obtained from a venous blood draw, this is not always practical, and can vary substantially from day to day in certain diseases such as anemia, making a current value essential. Thus, simultaneous T₁ and T₂ measurement is an attractive approach for quantification of Y_v. This was recently demonstrated using the T₂-TRIR technique⁶, where intravascular venous blood signal is measured during inversion recovery following various degrees of CPMG T₂-preparation. Unlike TRUST, T₂-TRIR does not use a tag-control approach to eliminate partial voluming, relying instead of saturation of the repeatedly excited tissue signal. Partial voluming errors represent a major concern in blood T₁ and T₂ quantification techniques due to the small size of the vessels of interest relative to the limited spatial resolution required for fast EPI readouts. We propose a modification of the TRUST sequence using multiple EPI readouts (mTRUST), allowing rapid determination of whole-brain venous blood T₁, T₂, and, therefore, Y_v, with optimal control of partial volume errors.

METHODS: Pulse Sequence – Like TRUST, mTRUST (Figure 1) uses non-selective MLEV-16 CPMG T₂ preparation pulses of varying effective echo time (eTE) following either blood inversion with an adiabatic hyperbolic secant pulse (tag) or application of an equivalent off-resonance pulse without gradient (control). An inversion time allows complete replacement of venous blood in the SSS of the imaging slice with inverted spins, after which a series of 90° excitation pulses with EPI readouts are applied every 200 ms to allow complete replacement of blood spins in each readout. The TR (6 s) and number of EPI readouts (8) were chosen based on signal equation calculations suggesting this combination would produce maximum T₂ measurement precision, assuming perfect inversion and T₂-preparation and no noise correlation between EPI readouts. **T₁/T₂ Determination** - Control-tag subtraction of each eTE image pair isolates venous blood signal with various degrees of T₁ and T₂ relaxation: $S = S_0 e^{-eTE \cdot (1/T_2 - 1/T_1)} e^{-t/T_1}$ where t is time relative to the end of the CPMG. An ROI containing only the four brightest pixels is selected, and T₁ determined from the 0 eTE data only, as these signals will have the highest SNR for fitting. This T₁ value is plugged back into the signal equation to calculate T₂, although it is worth noting that because T₁ >> T₂, the fitted T₂ is negligibly dependent on T₁ values within the physiologic range. Y_v is then determined from T₂ using a published calibration curve⁷. **In-Vivo Study** – 4 young healthy subjects (ages 32 ± 7, 3 males) were scanned with 10 repeats of the mTRUST sequence (8 mins total acquisition time). Mean and standard deviation (s.d.) T₁, T₂, and Y_v values were determined across the 10 repeats for each subject.

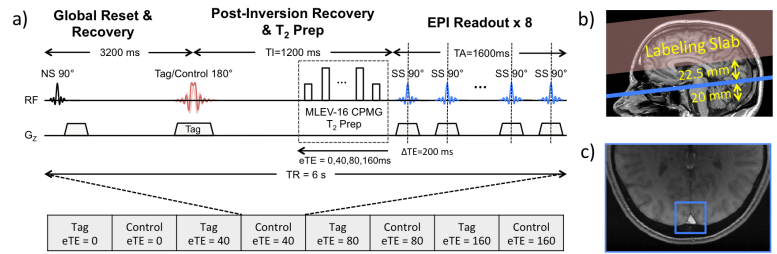


Figure 1: a) mTRUST pulse sequence diagram. Each 6 s module is repeated at 4 eTEs for both tag and control conditions, producing data for T₁ and T₂ fitting every 48 s. Additional sequence parameters: τ_{CPMG}=10ms, TE_{EPI}=7ms (5/8th partial Fourier), matrix=64x40, resolution=3.4x3.4x5.0mm. b) Sagittal image indicating locations of the labeling slab and imaging slice and c) axial image highlighting the SSS in the imaging slice.

RESULTS: Figure 2a shows mean ± s.d. raw difference signal intensities as well as corresponding Y_v values for Subject 2. In **Figure 2b**, 0 eTE log difference signal intensities are plotted to determine T₁. **Figure 2c** displays mean ± s.d. Y_v values across the four subjects vs. EPI readout number, demonstrating a consistent trend of Y_v over-estimation and reduced precision at later EPI readouts. Y_v values (derived from the first EPI readout only) and T₁ values for each subject are summarized in **Table 1**. Group Y_v values are similar to previous studies using TRUST⁸, whereas the quantified T₁ values are lower compared to recent reports using a similar fast T₁ approach⁹, especially for subjects 3 and 4.

Table 1: Subject Y_v and T₁ values.

Subject	Y _v	T ₁
1	61.3 ± 2.8	1643 ± 137
2	65.5 ± 1.6	1676 ± 105
3	67.7 ± 3.1	1466 ± 96
4	66.4 ± 2.3	1449 ± 57
Mean	65.2 ± 2.5	1559 ± 99

DISCUSSION: T₁ estimation – The main motivation for a combined blood T₁ and T₂ quantification approach is the ability to calibrate the T₂ estimation of Y_v based on T₁-derived Hct. Although the data suggest relatively precise T₁ fitting with a subject-averaged s.d. across repeats of 99 ms, the reported values are somewhat low. This would be expected were tissue partial voluming a concern, though this is unlikely given that the described sequence is optimally designed to reduce partial volume effects. A more likely explanation is that later arriving blood has a reduced degree of initial adiabatic inversion, causing a reduction in the difference signal measured at later EPI readouts unrelated to T₁ decay, and thus causing underestimation of T₁. This potential confound could be mitigated by using a larger inversion slab covering the entire head and measuring blood signal in the internal jugular vein, which would result in a longer bolus of fully inverted blood and also allow for a longer train of EPI readouts.

T₂ estimation – Another theoretical advantage of mTRUST as well as the previously described T₂-TRIR technique is the potential for improved T₂ estimation precision via the additional data acquired over multiple EPI readouts. In the T₂-TRIR study, all data was fitted to a single model, and it was not investigated whether the later EPI readouts improved T₂ estimation or not. However, it is clear from Figure 2a and 2c that Y_v is not only less precise at later EPI readouts, but also overestimated, suggesting that only the first EPI readout should be used for T₂/Y_v quantification. This reduced precision and bias is likely a function of the lower SNR of the later EPI readout signals, which would tend to result in T₂ overestimation.

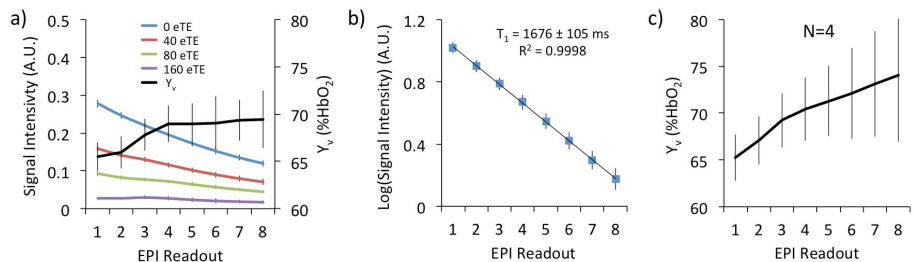


Figure 2: a) Difference signal values and corresponding Y_v values for Subject 2, with error bars indicating s.d. across repeats; b) T₁ fitting from data in Subject 2 demonstrates exponential behavior of the 0 eTE signal data; c) T₂-derived Y_v versus EPI readout across subjects with error bars indicating intersubject s.d.

CONCLUSIONS: mTRUST provides a fast approach to simultaneous T₁ and T₂ mapping of intravascular blood, but fitting should be preformed only with the 0 eTE data for T₁ and the first EPI readout data for T₂. Our results suggest that the T₂-TRIR approach of fitting all data to a single model may cause overestimation of T₂ and Y_v; however, determining the cause of this bias requires further investigation. While the proposed mTRUST approach is attractive in terms of control for partial volume effects, more work is needed to determine the validity of the approach for T₁ quantification, as the quantified values in the initial subjects studied were on the low end of normal. Future studies will directly compare mTRUST derived T₁ values to a similar sequence without T₂-preparation⁹, and also compare T₁ values to venipuncture-derived Hct to establish the feasibility of T₁-based Hct quantification for improved Y_v quantification.

REFERENCES: [1] Buxton et al., *MRM* (1998); [2] Lu et al., *MRM* (2003); [3] Chien et al., *JMRI* (1992); [4] Lu et al., *MRM* (2004); [5] Varela et al., *NMR Biomed* (2011); [6] De Vis et al., *Neuroimage* (2014); [7] Wright et al., *MRM* (1991); [8] Lu et al., *MRM* (2008); [9] Qin et al., *MRM* (2011). Grant Support: **NIH R21-HD069390 / T32-EB000814**.