## Quantitative Myocardial T1 and T2 mapping In a Swine Model of Ventricular Tachycardia

Sébastien Roujol<sup>1</sup>, Tamer A. Basha<sup>1</sup>, Cory Tschabrunn<sup>1</sup>, Kraig V. Kissinger<sup>1</sup>, Warren J. Manning<sup>1,2</sup>, Mark E. Josephson<sup>1</sup>, Elad Anter<sup>1</sup>, and Reza Nezafat<sup>1</sup>

Department of Medicine, Beth Israel Deaconess Medical Center and Harvard Medical School, Boston, Massachusetts, United States, <sup>2</sup>Department of Radiology, Beth Israel Deaconess Medical Center and Harvard Medical School, Boston, Massachusetts, United States

### **Target Audience**

Scientists/clinicians interested in myocardial tissue characterization.

#### Purpose/Introduction

Ventricular tachycardia ablation reduces the incidence of implantable cardioverter defibrillator (ICD) therapy in patients with history of myocardial infarction<sup>1</sup>. However, a high recurrence rate of VT is observed after ablation in these patients<sup>1</sup>. Therefore, the development of new techniques to identify the VT substrate may have important clinical impact. In this perspective, we have recently developed a swine model of re-entrant VT, where sustained monomorphic re-entrant VT can be induced in all animals. In this study, we sought to provide in-vivo characterization of this model using myocardial tissue characterization techniques of  $T_1$ ,  $T_2$  and high-resolution LGE.

#### **Materials and Methods**

All imaging was performed using a 1.5 T Philips scanner. A model of reentrant VT was induced in 11 Yorkshire swine using a 180 minutes balloon occlusion of the mid left anterior coronary artery. In-vivo CMR was performed at  $52\pm13$  days after infarction. Subsequently, each animal underwent an electrophysiology study with programmed stimulation to assess for VT inducibility.

 $\label{eq:cmr_objective} \underline{\text{In-vivo}} \ \ \underline{\text{CMR}} \ \ \underline{\text{Protocol:}} \ \ \underline{\text{In-vivo}} \ \ \underline{\text{CMR}} \ \ \text{was} \ \ \underline{\text{performed}} \ \ \text{with sedation,}$  intubation and mechanical ventilation of each animal. Pre-contrast imaging included native  $T_1$  mapping (using MOLLI²) and  $T_2$  mapping³. Post-contrast imaging was performed after bolus injection of 0.2 mmol/kg of gadobenate dimeglumine and included post-contrast  $T_1$  mapping (using MOLLI²) and high resolution late enhancement (LGE)⁴.

High resolution LGE (3) was performed using a free breathing navigator-gated inversion recovery gradient echo sequence with the following parameters (TR/TE/ $\alpha$ =6.5/3.0ms/25°, FOV=270×270×112 mm³, voxel size=1×1×1 mm³, compressed sensing factor=4).

## Results

Sustained re-entrant VT was induced in all animals. Areas of elevated native  $T_1$  times and  $T_2$  times corresponded well to areas of reduced post-contrast  $T_1$  times and LGE hyper-enhancement (Figure 1). These findings were found reproducible over all animals (Figure 2) where areas corresponding to LGE hyper-enhancement had higher native  $T_1$  times (1276±45 vs. 1047±29, p<0.001) and  $T_2$  times (85±6 vs. 52±4, p<0.001) than remote area.

# Conclusions

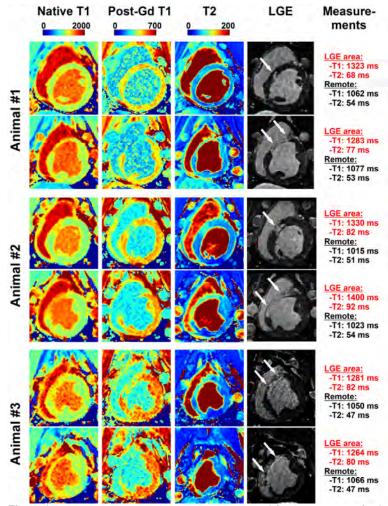
Areas corresponding to LGE hyper-enhancement have elevated native  $T_1$  times and  $T_2$  times in the developed swine model of re-entrant VT.

## Acknowledgements

Grant support from NIH R01EB008743-01A2 and Boston Scientific.

#### References

[1]Reddy,NEJM,2007; [2]Messroghli,MRM,2004; [3]Akçakaya,MRM, 2014; [4]Akçakaya,Rad.,2012



**Figure 1.** Native  $T_1$  maps, post-contrast  $T_1$  maps (post-Gd  $T_1$ ),  $T_2$  maps and LGE obtained in 3 swine. Elevated native  $T_1$  times and  $T_2$  times can be observed in the area of LGE hyper-enhancement.

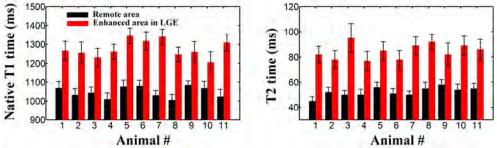


Figure 2. Native  $T_1$  times and  $T_2$  times measured in area corresponding to LGE hyper-enhancement (red) and in remote area (black). Higher  $T_1$  times and  $T_2$  times were obtained in area corresponding to LGE hyper-enhancement (p<0.001).