## Measurement of Local Cerebral Hematocrit with MRI

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**Purpose:** Propose and assess a method to measure local cerebral hematocrit using MRI.

Introduction: Red blood cells carry O<sub>2</sub> to tissue and transport CO<sub>2</sub> back to lungs. The percentage blood volume occupied by red blood cells is known as hematocrit (Hct). Whereas it is straightforward to measure Hct in large arteries (e.g. blood sample), it is very challenging to do it in brain microvasculature (i.e. cerebral Hct, Hct<sub>nissue</sub>). Currently, this can only be done using invasive methods, such as with PET, SPECT, and autoradiography<sup>1-3</sup>, but their use is very limited. Local variations in cerebral Hct have been reported in various brain abnormalities (e.g. stroke, vascular disease, and tumours). 4-6 Knowledge on cerebral Hct is also important for dynamicsusceptibility contrast (DSC) MRI and perfusion CT, since their contrast agents reside in blood plasma. These methods, therefore, commonly rely on assumed (uniform) literature values<sup>7</sup>, which can lead to errors. We propose here a MRI method to image cerebral Hct, which relies on combining data from two MRI measurements of hemodynamic parameters: one that provides Hct-weighted values, and another one that provides Hct-independent values of the same parameter. By combining these two measurements, it is possible to isolate local cerebral Hct, thus providing an easily obtainable measurement of this important physiological parameter.

Methods: There are various ways to combine MRI measurements to calculate cerebral Hct (see Discussion). We describe one approach in detail here, which relies on data from (i) a DSC-MRI study in absolute units (e.g. 8,9) and (ii) an arterial spin labelling (ASL) study. In this case, a DSC-MRI deconvolution analysis provides a Hct-weighted CBF (CBF<sub>Hct</sub>) in absolute units, i.e.  $CBF_{DSC-MR} = CBF_{Hct} = CBF_{Hct}$ , where  $k_{Hct}$  is the large-to-small vessel size plasma volume ratio:  $(1 - Hct_{art})/(1 - Hct_{insue})$ , and Hct<sub>art</sub> is the Hct in a large artery. Note that, for most DSC-MRI studies, this scaling factor is considered a nuisance (i.e. neglected or fixed to an assumed literature value), but its presence is central to the method proposed here. From a complementary ASL measurement, known to provide absolute CBF measurements (i.e., CBF<sub>ASL</sub>=CBF), <sup>10</sup> one can calculate  $k_{Hct}$  as the ratio:  $k_{Hct} = CBF_{ASL}/CBF_{Hct}$ , which therefore leads to cerebral Hct as:  $Hct_{tissue} = 1 - (CBF_{Hct}/CBF_{ASL}) \times (1 - Hct_{art})$ . Data acquisition: Data from 17 healthy subjects were included, each scanned twice on a 3T Philips MRI scanner. DSC-MRI was carried out using the pre-bolus method9 to obtain quantitative CBF-measurements: contrast agent pre-bolus (0.02 mmol/kg) and segmented EPI (single slice, temporal resolution=0.81s, 1.72×1.72×5 mm<sup>3</sup>). The pre-bolus scan was followed by the actual single-dose DSC-MRI study (0.1 mmol/kg), using single-shot gradient-echo EPI (1.72×1.72×5 mm<sup>3</sup>, 20 slices, TE/TR=29/1243ms). ASL was carried out using background-suppressed single-shot EPI pCASL (1.65s label duration, 1.6s post-labelling delay, 2.29×2.29×5 mm<sup>3</sup>, 16 slices, TE/TR=14/4000 ms, 30 repetitions); a reference M<sub>0</sub> scan was performed without labelling or background suppression, TR=10s, and 4 repetitions.

Data analysis: Hct-weighted CBF (from DSC-MRI) was obtained by scaling the AIF by the VOF area from the prebolus experiment,9 and using oSVD deconvolution.11 Hct-independent CBF (from ASL) was calculated using a model resembling the recent consensus recommendations. <sup>10</sup> CBF maps were normalized to MNI152 template using SPM8. Cerebral Hct was estimated using Eq.1. Since no Hct<sub>art</sub> measurements were available, this value was set to 0.45. However, to investigate the sensitivity to the assumed  $Hct_{art}$ , the analysis was repeated for  $Hct_{art}$ =0.3–0.6. For more detailed analysis, regions in cortical and subcortical grey matter structures were manually defined. Test-retest reproducibility was assessed using Bland-Altman analysis.

Results: Fig. 1 shows population average results, including average CBF and Hct<sub>tisssue</sub> maps. Fig. 2 shows the population average Hct<sub>tissue</sub> maps in 3 MNI space projections. Fig. 3 shows the cerebral-to-arterial Hct ratio in subcortical and cortical regions, as a function of the assumed Hct<sub>art</sub>: the ratio is, in general, consistent with literature

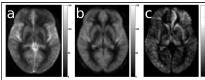
values. 1.2 The method had good reproducibility, with a Bland–Altman bias (and limits of agreement) for the subcortical structures of 0.007 (0.115), and for cortical structures 0.018 (0.102).

**Discussion:** An MRI method to measure cerebral Hct was proposed, which relies on combining measurements from 2 MRI techniques, one that provides Hct-dependent and another one with Hctindependent values. The method was illustrated using DSC-MRI and ASL data on a group of healthy subjects, and was shown to measure cerebral Hct in general agreement with literature values, 1.2 with good test-retest reproducibility. The proposed MRI method could have important applications in various neurological diseases, such as stroke and tumours.

Alternative implementations of this method could be done using CBV measurements instead of CBF (e.g. combining DSC-MRI with either steady-state T<sub>1</sub> data<sup>13</sup> or calibrated VASO data<sup>14</sup>), or

combining steady-state T<sub>1</sub> and T<sub>2</sub> CBV measurements (which are in fast and slow exchange, respectively)<sup>15</sup>. CBVbased implementations are likely to be more robust, given that these do not rely on deconvolution. Furthermore, these alternative implementations are likely to be more suited for white matter Hetzissue, given the known difficulties with quantifying white matter CBF using ASL. In particular, the steady-state implementation, when combined with blood pool agents, may open up the possibility of mapping cerebral Hct with high resolution and SNR. Finally, more robust measurements will be obtained when individual Hct<sub>art</sub> are available, to account for its inter-subject variability.

References: [1] Lammertsma AA et al, J Cereb Blood Flow Metab 1984; 4: 317-322. [2] Sakai F et al, J Cereb Blood Flow Metab 1985; 5: 207-213. [3] Bereczki D et al, J Appl Physiol 1985 1992; 73: 918-924. [4] Loutfi I et al, Am J Physiol Imag 1987; 2: 10-16. [5] Sakai F et al, Acta Neurol Scand Suppl 1989; 127: 9-13. [6] Brooks DJ et al, Microvasc Res 1986; 31: 267-276. [7] Calamante F et al, J Cereb Blood Flow Metab 1999; 19: 701-735. [8] van Osch MJP et al, Magn Reson Med 2003; 49: 1067-1076. [9] Knutsson L et al, Magn Reson Med 2014;72: 996-1006. [10] Alsop DC et al, Magn Reson Med 2014; doi: 10.1002/mrm.25197. [11] Wu O et al, Magn Reson Med 2003; 50: 164-174. [12] Purves WK et al, Life: The Science of Biology. W.H. Freeman & Co Ltd, 2004. [13] Shin W et al, Magn Reson Med 2007; 58: 1232-1241. [14] Lu H et al, Magn Reson Med 2005; 54: 1403-1411. [15] Donahue KM et al, J Magn Reson Imaging 1997; 7: 102-110.



**Figure 1**. Group results. (a) CBF<sub>Hct</sub> from DSC-MRI; (b) CBF from ASL; (c) Hct<sub>tissue</sub>. CBF colorbars in ml/100g/min (range 0-150); Hct colorbar is unitless (range 0-0.6).

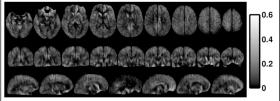


Figure 2. Population averaged Hct<sub>tissue</sub>, in axial (top), coronal (middle), and sagittal (bottom) projections.

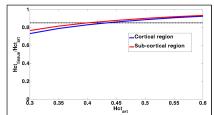


Figure 3. Cerebral-to-arterial Hct ratio as a function of assumed Hctart, for subcortical (red line) and cortical (blue line) regions. The classical literature value<sup>1,2</sup> of 0.85 is shown as dashed line.