

DETECTION OF MATRIX METALLOPROTEINASES USING AN "ON/OFF" ^{19}F MR PROBE

Alex John Taylor¹, James Lee Krupa², Huw Williams³, Dorothee P Auer¹, Simon R Johnson⁴, Neil R Thomas², and Henryk Michael Faas¹

¹Sir Peter Mansfield Imaging Centre, School of Medicine, University of Nottingham, Nottingham, Nottinghamshire, United Kingdom, ²School of Chemistry, University of Nottingham, Nottingham, Nottinghamshire, United Kingdom, ³Centre for Biomolecular Sciences, University of Nottingham, Nottingham, Nottinghamshire, United Kingdom, ⁴School of Medicine, University of Nottingham, Nottingham, Nottinghamshire, United Kingdom

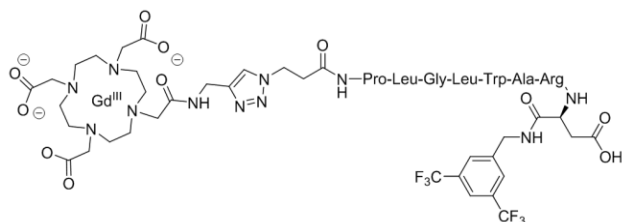


Figure 1: Chemical structure of MMP sensor: Chelated gadolinium is linked via a peptide sequence to a fluorinated amino acid; the linker is cleaved upon contact with specific MMP. Here, the peptide sequence is a generic substrate for MMPs.

have developed a generic MMP and an MMP 2 and 9 (2/9) substrate sensor. In the uncleaved state, the paramagnetic influence of the gadolinium will cause short relaxation times and a broad fluorine signal. If the linker is cleaved by the specific MMP, then the distance between the gadolinium is increased, causing a change in relaxation time; this "on/off" principle was first demonstrated for fluorine MR sensors by Mizukami et al.². Two samples were prepared with 600 μL of 100 μM of each MMP sensor and 5 μL of MMP 9 (10 mg / ml) was then added. Following MMP addition, a Bruker 600 MHz spectrometer was used to measure a 2-D time course in which a single scan (TR = 5 s, NS = 32, Sweep width = 50 kHz, Acq. time \sim 2 m 30 s) was repeated for a period of around 13 hours. A further inversion recovery experiment was performed for both sensors, before and after MMP addition, to investigate relaxation properties.

Results - A clear change is present in the fluorine signal intensity after MMP addition; a 3.5-fold increase is found, as shown in Fig. 3. The T_1 relaxation time (and T_2^* time) also increase after the enzyme is added (see Fig. 2); T_1 values change by around a factor of 60, with a ten-fold increase in T_2^* . Figure 3 B is a 3-D plot demonstrating the change in spectral shape over time; the effect upon the fluorine signal intensity following enzymatic cleavage was modelled using Michaelis-Menten type equations. Sensor specificity was also verified using mass spectroscopy for a sample combination of the MMP 2/9 with MMP 12; no cleavage products were found.

Discussion – Using a synthesised ^{19}F sensor, we have demonstrated an observable contrast mechanism following MMP detection. Since a signal is observed with spectroscopy, a higher sensor concentration (\sim mM) will be detectable with MRI on an imaging platform. Similar T_1/T_2^* relaxation times are found for both MMP sensors, implying that other enzyme substrates would exhibit a similar relaxation difference. Our findings are in agreement with a recent paper by Yue et al. which showed a similar order of magnitude change in T_1 relaxation³.

Conclusion – We have presented preliminary data for two sensors with different peptide specific sequences; both sensors show changes in the MR signal after contact with MMPs. Monitoring of MMP levels could give an important biomarker of the first stages of disease progression in neurodegenerative diseases and future work will aim to apply this "on/off" mechanism *in vivo* to an animal disease model which over expresses MMPs.

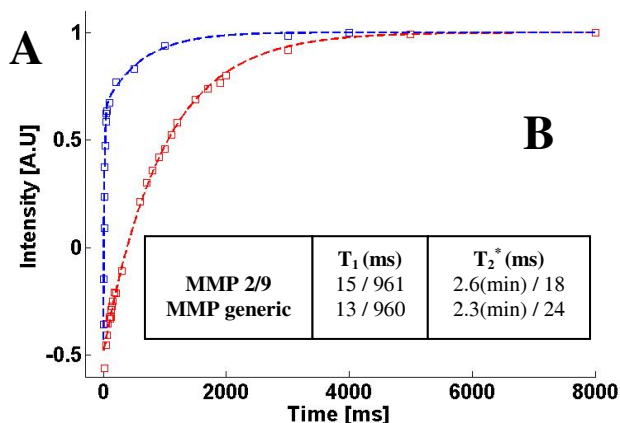


Figure 2: Relaxation differences between pre and post cleavage states for sensors: (A) T_1 inversion recovery experiment showing data points for pre (blue) and post (red) cleavage after MMP 9 addition for the MMP 2/9 sensor (fitted curve is used to find T_1 value.); (B) Table showing relaxation properties for the MMP 2/9 sensor; data is of the pre MMP addition/ post MMP addition form.

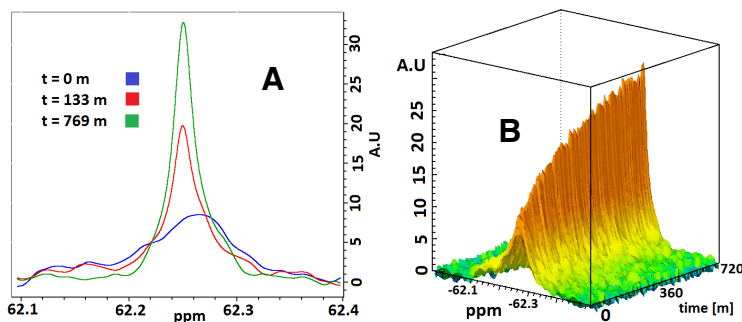


Figure 3: Signal intensity time course for generic MMP sensor after MMP 9 addition: (A) 1-D fluorine spectrum showing the narrowing and increase of the peak intensity over time; (B) 3-D intensity plot where the exponential nature of the intensity increase, due to the enzymatic cleavage, is visible.

References 1. McClain J, Phillips L, Fillmore H. *Neurosci Lett*, 2009;460(1):27-31 2. Mizukami S, Takikawa R, Fuminori S, et al. *J. Am. Chem. Soc.*, 2008;130 (3):794–795 3. Yue X, Wang Z, Zhu L, et al., *Mol. Pharmaceutics*, 2014;11 (11): 4208-4217