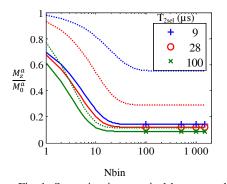
## Cross-relaxation parameter quantification in cortical bone from repeated binomial excitations

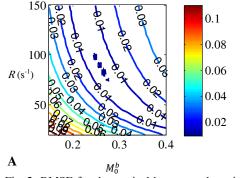
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<u>Purpose:</u> Evidence of magnetization transfer (MT) was recently shown in cortical bone between collagen-bound water protons and collagen methylene protons<sup>1</sup>. A quantitative assessment of cross-relaxation parameters is of interest considering the observation of Horch et al.<sup>2</sup> that the amount of collagen-bound water protons  $(M_0^b)$  was related to cortical bone mechanical properties. The cross-relaxation parameters were quantified using off-resonance saturation<sup>3</sup>. However, this approach is limited by the high Specific Absorption Rate and the long acquisition time. The purpose of this work was to quantify the cross-relaxation parameters in cortical bone using a scheme easily integrated in an imaging sequence.

Materials and Methods: Femoral bovine cortical bone was obtained from local butcher, 11 transverse sections were cut from the diaphysis over approximately 10 mm thickness. Experiments were run on a home-assembled 4.7 T scanner. Inversion-recovery (IR) was used to follow longitudinal relaxation with 31 inversion times using two pulse widths (pw):  $10 \,\mu s$  (TE =  $55 \,\mu s$ ) and  $100 \,\mu s$  (TE =  $100 \,\mu s$ ), and CPMG to follow transverse relaxation with TE =  $0.75 \,m$ s at both pw. A third order selective binomial excitation was implemented to saturate the longitudinal magnetization of protons with a specific T<sub>2</sub> called T<sub>2sel</sub> with a minor perturbation of the long-T<sub>2</sub> proton magnetization. To attain a steady state, the binomial excitation was repeated after a delay (11.2 ms) and for a determined number of excitations (Nbin = 100-1500). A hard  $90^{\circ}$  RF pulse followed by a  $100 \,\mu s$  dead time was applied to measure M<sub>z</sub><sup>2</sup>; M<sub>0</sub><sup>a</sup> was similarly measured after TR =  $15 \, s$ . Experiments were simulated with a matrix approach<sup>5</sup> for a two-pool model using the usual cross-relaxation parameters<sup>6</sup> with home-developed software written in Matlab (MathWorks, Natick, MA).

Results: At pw = 10 µs, IR data could be described as a monoexponential decay, whereas at pw = 100 µs, the IR data were compatible with a biexponential decay. This behavior was comparable for all examined samples. A biexponential decay for the long pw experiment is in agreement with two characteristic times<sup>7</sup> describing the return to equilibrium of  $M_z^a$  with the two-pool model. Data at both pw could be reproduced by simulation of the two-pool model. The model parameters were initialized as follows:  $T_1^a = T_1^b = \text{time of the monoexponential fit of IR data at pw = 10 µs, <math>T_2^a = T_2^*$ , then  $T_2^b$ , R and  $M_0^b$  were searched to minimize the Root Mean Square Error (RMSE) between simulation and experimental data. CPMG data were analyzed as a decreasing tri-exponential function and were not sensitive to MT. Indeed the mean fast  $T_2$  component was equal to 0.466 ms (±0.07 ms) at pw = 10 µs and to 0.488 ms (±0.1 ms) at pw = 100 µs, with similar relative fraction (≈ 90%). To simulate the repeated binomial experiment,  $T_1^a$  was initialized from IR monoexponential decay at pw = 10 µs and  $T_1^b$  was set =  $T_1^a$ . Four parameters ( $T_2^a$ ,  $T_2^b$ ,  $M_0^b$  and R) were set to minimize RMSE between experiments and simulations. The saturation data in cortical bone samples were systematically much lower than the one-pool simulation (Fig. 1), and the two-pool model with  $T_1^a = T_1^b = 0.4$  s,  $T_2^a = 1.9$  ms,  $T_2^b = 13$  µs,  $M_0^b = 0.24$ , R = 72 s<sup>-1</sup> was in a fair agreement with data. Increasing  $T_{2sel}$  caused a decrease of  $M_2^a/M_0^a$  mainly due to direct saturation. Therefore the largest difference between the one-pool simulation and the data could be found for the shortest  $T_{2sel}$  (9 µs) which was also the closest value to  $T_2^b$ . Fig. 2A shows as iso-contour lines RMSE between data of Fig. 1 and simulations for different  $M_0^b$  and R values, on a two-pool system. All iso-contour lines tended to be vertical for R > 100 s<sup>-1</sup> therefore R > 100 s<sup>-1</sup> cannot be accurately predicted.





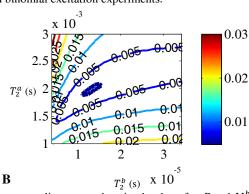


Fig. 1: Saturation in a cortical bone sample (symbols), one-pool simulation (dotted lines) and two-pool simulation (lines)

Fig. 2: RMSE for the cortical bone sample as iso-contour lines around optimal values for R and  $M_0^b$  (A) and  $T_2^a$  and  $T_2^b$  (B) minimizing RMSE (RMSE<sub>min</sub>= 0.002)

Table 1: Cross-relaxation parameters (mean±std.) of the 11 investigated cortical bone samples from IR and repeated binomial excitation experiments

	$T_1^a = T_1^b$ (s)	$T_2^a$ (ms)	$T_2^b$ (µs)	$M_0^b$ (%)	$\mathbf{R}(s^{-1})$
Inversion-Recovery	0.40±0.01	0.20±0.02	11±0.36	50±4	155±19
Repeated binomial excitation	$0.40\pm0.01$	$1.86 \pm 0.31$	14±3	25±5	89±39

<u>Discussion and Conclusion:</u>  $T_1^a$  from IR data was close to 0.4 s in agreement with literature<sup>1</sup> and  $T_2^b$  deduced from both experiments were comparable.  $M_0^b$  could not be precisely determined from the IR data. Indeed, as  $180^\circ$  excitation was not repeated,  $M_z^a$  was less perturbed by  $M_z^b$ .  $M_0^b$  from repeated binomial excitation simulation was in agreement with past findings<sup>1</sup>. The optimal  $T_2^a$  from repeated binomial excitation being longer than the fast  $T_2$  from CPMG is tentatively attributed to susceptibility inhomogeneities. IR can be used to detect MT (large  $M_0^b$  and short  $T_2^b$ ). However to quantify cross-relaxation parameters, a repeated binomial excitation can be easily integrated into a UTE sequence and should be used.

References: 1. Horch RA et al. Magn Reson Med 2010;64:680-687. 2. Horch RA et al. PLoS ONE 2011;6:e163592011. 3. Bouazizi-Verdier K and Guillot G. In: Proceedings of the ISMRM, 2014 (#3995). 4. Pachot-Clouard M and Darrasse L. Magn Reson Med 1995;34:462–469. 5. Müller DK et al. J Magn Reson 2013;230:88–97. 6. Henkelman RM et al. Magn Reson Med 1993;29:759–766. 7. Edzes HT and Samulski ET. J Magn Reson 1978;31:207–229.