

# Validation of adiabatic T<sub>1</sub>ρ and T<sub>2</sub>ρ mapping of articular cartilage at 3T

Victor Casula<sup>1,2</sup>, Joonas Autio<sup>3</sup>, Mikko J. Nissi<sup>3,4</sup>, Michaeli Shalom<sup>4</sup>, Silvia Mangia<sup>4</sup>, Edward Auerbach<sup>4</sup>, Jutta Ellermann<sup>4</sup>, Eveliina Lammintausta<sup>3</sup>, and Miika T. Nieminen<sup>1,3</sup>

<sup>1</sup>Department of Radiology, University of Oulu, Oulu, Oulu, Finland, <sup>2</sup>Medical Research Center Oulu, Oulu University Hospital and University of Oulu, Oulu, Finland, <sup>3</sup>Department of Diagnostic Radiology, Oulu University Hospital, Oulu, Finland, <sup>4</sup>Center for Magnetic Resonance Research, Department of Radiology, University of Minnesota, Minneapolis, United States

**TARGET AUDIENCE:** Researchers and clinicians aiming to apply quantitative MRI techniques in osteoarthritis research.

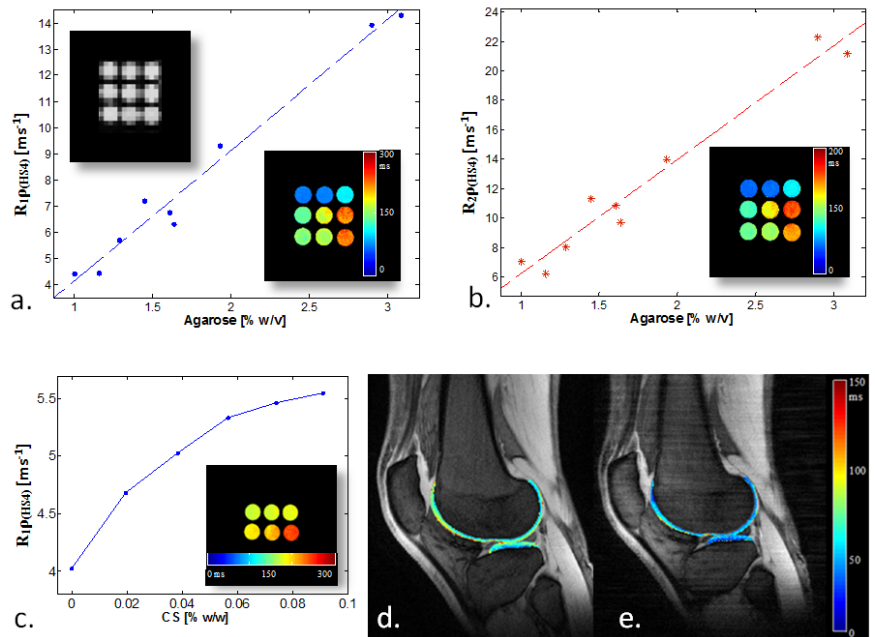
**PURPOSE:** To validate adiabatic T<sub>1</sub>ρ (AdT<sub>1</sub>ρ) and adiabatic T<sub>2</sub>ρ (AdT<sub>2</sub>ρ) sequences for quantitative measurements of cartilage with a 3 T clinical scanner.

**METHODS:** AdT<sub>1</sub>ρ and AdT<sub>2</sub>ρ sequences<sup>1,2</sup> were tested in a 3 T clinical system (Siemens Skyra) in combination with a 15 channel knee coil for signal transmission/reception. Adiabatic T<sub>1</sub>ρ experiments were performed as described previously<sup>2,3</sup> using a preparation block which consisted of a train of 0, 4, 8, 12 and 16 adiabatic full passage (AFP) hyperbolic secant pulses of the HS<sub>n</sub> family, here HS<sub>4</sub>. For adiabatic T<sub>2</sub>ρ the AFP pulses were placed between two adiabatic half passage pulses (AHP). The preparation block was followed by a gradient recalled echo (FLASH) readout (TR / TE = 20 s / 3.75 ms, 15° flip angle, 256 x 256 matrix, 7 mm slice thickness, FOV = 120 x 120 mm<sup>2</sup>, 2 averages). For both AdT<sub>1</sub>ρ and AdT<sub>2</sub>ρ the RF peak amplitude was γB<sub>1max</sub> = 800 Hz; R=10 and pulse duration 6 ms was used, which resulted in the BW = 1.6 kHz. Longitudinal and transverse relaxation time constants in the rotating frame were calculated by mono-exponential fitting of the signal intensity decays on a pixel-by-pixel basis. Actual flip angle maps were calculated to evaluate B<sub>1</sub> homogeneity. Nine agarose gel nickel-doped phantoms were prepared with relaxation times within biological tissue range (T<sub>1</sub> = 400 - 2600 ms, T<sub>2</sub> = 40 - 170 ms). Reproducibility was assessed by calculating the coefficient of variation (CV %) of three measurements repeated within one month. Six chondroitin sulfate (CS), the main proteoglycan constituent (0.0, 2.0, 4.0, 6.0, 8.0, 9.0 % w/w, pH 7.1), tubes were prepared. R<sub>1</sub>ρ (=1/T<sub>1</sub>ρ) and R<sub>2</sub>ρ (=1/T<sub>2</sub>ρ) were studied as a function of CS concentration. Finally, the feasibility of AdT<sub>1</sub>ρ and AdT<sub>2</sub>ρ measurements in the human knee joint was demonstrated.

**RESULTS:** B<sub>1</sub> mean error in the region of interest was 4.8 % (Fig. 1a inset left), indicating good B<sub>1</sub> homogeneity. R<sub>1</sub>ρ and R<sub>2</sub>ρ measured with HS<sub>4</sub> pulses were linearly dependent on agarose concentration (Fig. 1a and 1b). Reproducibility as indicated by CV% was 0.77 % and 2.0 % for AdT<sub>1</sub>ρ and AdT<sub>2</sub>ρ, respectively. R<sub>1</sub>ρ and R<sub>2</sub>ρ were dependent on chondroitin sulfate concentration and on the inversion bandwidth (Fig. 1c). Figures 1d and 1e show T<sub>1</sub>ρ and T<sub>2</sub>ρ maps of human knee cartilage.

**DISCUSSION:** Previously, adiabatic T<sub>1</sub>ρ and adiabatic T<sub>2</sub>ρ have been applied in preclinical studies at different field strengths and their ability to probe slow molecular motion has been shown<sup>1,2</sup>; former *in vitro* studies have observed high sensitivity of AdT<sub>1</sub>ρ and AdT<sub>2</sub>ρ to animal and human cartilage degeneration at 9.4 T<sup>3-5</sup>. At 3 T the sequences showed very good reproducibility, higher for AdT<sub>1</sub>ρ compared to AdT<sub>2</sub>ρ. R<sub>1</sub>ρ and R<sub>2</sub>ρ were very sensitive to variations in both agarose and CS. Finally, the sequences were adapted for quantitative measurement of articular cartilage and then successfully applied *in vivo*.

**CONCLUSION:** Phantom experiments with hyperbolic secant pulses (HS<sub>4</sub>) revealed excellent accuracy of the sequences and strong dependencies of R<sub>1</sub>ρ and R<sub>2</sub>ρ on agarose and chondroitin sulfate concentration, which are relevant for cartilage. The findings demonstrate that adiabatic T<sub>1</sub>ρ and adiabatic T<sub>2</sub>ρ techniques are promising tools for *in vivo* cartilage imaging at 3T.



**Figure 1.** (a) R<sub>1</sub>ρ vs. agarose concentration (inset left; actual flip angle map; inset right: R<sub>1</sub>ρ map). (b) R<sub>2</sub>ρ vs. agarose concentration (inset: T<sub>2</sub>ρ map). (c) R<sub>1</sub>ρ vs. CS concentration (inset: T<sub>1</sub>ρ map). (d) T<sub>1</sub>ρ map of human cartilage. (e) T<sub>2</sub>ρ map of human cartilage.

**REFERENCES.** 1. Michaeli et al. Curr Anal Chem 2008, 4:8–25. 2. Mangia et al. MRI 2009, 27(8):1074-87. 3. Rautiainen et al., MRM 2014, doi: 10.1002/mrm.25401. 4. Rautiainen et al., OAC 2014, 22(10):1444-522. 5. Ellermann et al. MRI 2013, 31:1537-43. 6. Regatte et al. JMRI 2003, 17(1):114-21.