

## Specialty Area: Molecular Imaging

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### Highlights

- Overview of state-of-the-art hyperpolarized (HP) pulse sequences
- Discussion of RF excitation, data acquisition, acceleration methods, acquisition timing and image contrast
- Comparison of tradeoffs between acquisition and reconstruction strategies

### How to Detect HP Agents: Pulse Sequences

**TARGET AUDIENCE:** Scientists, engineers, and physicians interested in MRI with hyperpolarized agents.

**OUTCOME/OBJECTIVES:** The goal of this talk is to describe MR pulse sequence strategies that are specifically tailored to the unique physics, biochemical conversions, and in vivo behavior of hyperpolarized (HP) agents. Participants will be able to understand various pulse sequence strategies and tradeoffs between them in terms of SNR, acquisition time, and robustness.

**PURPOSE:** Hyperpolarized agents can provide unique information about tissue function by probing metabolism, perfusion, redox state, and more. The unique behavior of these agents, including unrecoverable T1 relaxation, conversion to other compounds with different chemical shifts, and bolus injection dynamics, necessitates specialized MR pulse sequences.

**METHODS, RESULTS, & DISCUSSION:** HP agent pulse sequences involve specialized strategies for **RF excitation**, **data acquisition**, and **acquisition timing**. These sequences benefit from **acceleration** and can be combined with specialized **image contrast**.

#### RF excitation strategies

1. *Variable flip angles:* To efficiently sample the limited, unrecoverable hyperpolarization available, variable flip angles are used to account for magnetization losses from previous RF pulses [1]. These can be designed considering T1 decay and metabolic conversion [2].
2. *Multiband excitation:* When attempting to observe a metabolic product, it is advantageous to minimally perturb the hyperpolarized substrate, particularly in dynamic imaging. Multiband excitation refers to applying different flip angles to different compounds using spectrally-selective RF pulses [3, 4, 5].

Acquisition strategies: (For agents that are not converted into other compounds, such as  $^{13}\text{C}$ -urea,  $^{13}\text{C}$ -HP001,  $^{13}\text{C}$ -tert-butanol, conventional MRI acquisition strategies combined with HP RF excitation strategies are highly efficient.)

1. *MR spectroscopic imaging (MRSI):* In MRSI, spectral and spatial dimensions are fully resolved. For HP agents, spectral-spatial readout gradients are used to accelerate the acquisition, including echo-planar spectroscopic imaging (EPSI) [6, 7], concentric rings [8, 9], radial spectroscopic imaging [10], and spiral CSI [11]. Of these three, EPSI is the slowest but most robust, rings are faster but still robust, while spiral CSI is the fastest but most sensitive to system imperfections. For the large spectral bandwidths found with some HP agents, an alternating band strategy may be necessary [12]. Overall MRSI is generally the slowest acquisition strategy but is very flexible and provides the most spectral information.
2. *Chemical shift separation methods:* Images for compounds with known chemical shifts can be created from acquisitions at a few echo times (generally N+1 TEs for N different compounds) [13, 14]. This can be approximated using oversampled spiral trajectories

[15]. Different compound images can also be created by exploiting the chemical shift frequency or EPI phase encoding [16, 17, 18]. These methods are faster than MRSI, but are constrained by the chemical shifts present

3. *Metabolite-specific imaging*: Rapid imaging of compounds with sufficiently large chemical shifts can be performed by using a spectrally-selective excitation of a single compound followed by a fast imaging readout, such as spirals or EPI [19, 20, 21, 22]. This can also be accomplished using the steady-state free precession (SSFP) frequency response [23]. These methods are extremely fast and efficient, and work very well for pyruvate and its metabolic products. They may not be readily applicable to all HP agents, depending on the chemical shifts present.

Acquisition Timing: Data can be either as a dynamic time series or from a single time-point. Single time-point acquisition has an SNR advantage because all the available magnetization is devoted to one acquisition. Dynamic acquisitions allow for measurement of conversion and relaxation rates, monitoring perfusion, and are less sensitive to injection and acquisition timing, but have less SNR. [4, 5, 24, 25, 26, 27, 28] Another consideration in the acquisition timing is calibration. Rapid B1 mapping sequences have been applied to HP agents for calibration prior to imaging acquisition [29, 30].

Acceleration methods: HP pulse sequences benefit significantly from accelerated imaging methods. Unlike conventional MRI, there is no SNR penalty because the variable flip angles can be adjusted to redistribute the available magnetization. Furthermore, signal loss and blurring due to T1 relaxation, metabolic conversion, motion, and flow during the acquisition is reduced during a shorter acquisition.

1. *Parallel imaging* has been successfully applied for HP agents when appropriate coil arrays are available [31, 32, 33]. Autocalibrating methods such as GRAPPA are highly desirable because it is difficult to obtain non-proton sensitivity maps due to low natural abundance.
2. *Compressed sensing* has also been shown to be very effective taking advantage of sparsity in the HP agent spectrum as well as in the dynamics and spatial distribution [34, 35].

Contrast methods: Similar to conventional MRI, HP pulse sequences have been designed to provide novel contrast.

1. *T2-mapping* with multiple spin-echoes has shown promise for novel contrast [36, 37, 38].
2. *Diffusion-weighted* pulse sequences may provide additional information regarding HP agent compartmentalization and/or vascular suppression [38, 39, 40, 41, 42].
3. *2D NMR, saturation transfer, inversion recovery, and related methods* have promise for quantitative direction detection of HP agent exchange and metabolic conversion [5, 43, 44, 45, 46, 47]
4. *Selective spatial or spectral saturation* can be used to isolate the source of metabolites or reduce acquisition constraints [48].

**CONCLUSION**: Specialized pulse sequences must be designed for HP agents to account for unrecoverable signal decay, conversion between compounds, and in vivo dynamics. RF excitation should be done with variable flip angles for efficient magnetization usage. MRSI, chemical shift separation, and metabolite-specific imaging are all promising acquisition strategies, with MRSI the most flexible but slowest compared whereas metabolic-specific imaging is the fastest but most restrictive pulse sequence design. Dynamic imaging is less sensitive to injection timing but has generally less SNR than single time-point imaging. Parallel imaging and compressed sensing acceleration is very advantageous for HP agents. These can

all be combined with pulse sequence contrast strategies to be sensitive to agent compartmentalization and exchange/conversion.

## REFERENCES

1. Zhao, L., Mulkern, R., Tseng, C.-H., Williamson, D., Patz, S., Kraft, R., Walsworth, R. L., Jolesz, F. A., & Albert, M. S. (1996) Gradient-Echo Imaging Considerations for Hyperpolarized  $^{129}\text{Xe}$  MR. *J Magn Reson B* 113, 179-183.
2. Xing, Y., Reed, G. D., Pauly, J. M., Kerr, A. B., & Larson, P. E. Z. (2013) Optimal variable flip angle schemes for dynamic acquisition of exchanging hyperpolarized substrates. *J Magn Reson* 234, 75-81.
3. Larson, P. E. Z., Kerr, A. B., Chen, A. P., Lustig, M. S., Zierhut, M. L., Hu, S., Cunningham, C. H., Pauly, J. M., Kurhanewicz, J., & Vigneron, D. B. (2008) Multiband excitation pulses for hyperpolarized  $^{13}\text{C}$  dynamic chemical-shift imaging. *J Magn Reson* 194, 121-7.
4. Lau, A. Z., Chen, A. P., Barry, J., Graham, J. J., Dominguez-Viqueira, W., Ghugre, N. R., Wright, G. A., & Cunningham, C. H. (2013) Reproducibility study for free-breathing measurements of pyruvate metabolism using hyperpolarized ( $^{13}\text{C}$ ) in the heart. *Magn Reson Med* 69, 1063-71.
5. Schulte, R. F., Sperl, J. I., Weidl, E., Menzel, M. I., Janich, M. A., Khagai, O., Durst, M., Ardenkjaer-Larsen, J. H., Glaser, S. J., Haase, A., Schwaiger, M., & Wiesinger, F. (2013) Saturation-recovery metabolic-exchange rate imaging with hyperpolarized [1-( $^{13}\text{C}$ )] pyruvate using spectral-spatial excitation. *Magn Reson Med* 69, 1209-16.
6. Yen, Y.-F., Kohler, S. J., Chen, A. P., Tropp, J., Bok, R., Wolber, J., Albers, M. J., Gram, K. A., Zierhut, M. L., Park, I., Zhang, V., Hu, S., Nelson, S. J., Vigneron, D. B., Kurhanewicz, J., Dirven, H. A. A. M., & Hurd, R. E. (2009) Imaging considerations for in vivo  $^{13}\text{C}$  metabolic mapping using hyperpolarized  $^{13}\text{C}$ -pyruvate. *Magn Reson Med* 62, 1-10.
7. Cunningham, C. H., Vigneron, D. B., Chen, A. P., Xu, D., Nelson, S. J., Hurd, R. E., Kelley, D. A., & Pauly, J. M. (2005) Design of flyback echo-planar readout gradients for magnetic resonance spectroscopic imaging. *Magn Reson Med* 54, 1286-9.
8. Furuyama, J. K., Wilson, N. E., & Thomas, M. A. (2012) Spectroscopic imaging using concentric circular echo-planar trajectories in vivo. *Magn Reson Med* 67, 1515-22.
9. Jiang, W., Lustig, M., & Larson, P. E. Z. (2013) Concentric Rings K-Space Trajectory for Hyperpolarized  $^{13}\text{C}$  MRSI. Proc. ISMRM, 1914.
10. Ramirez, M. S., Lee, J., Walker, C. M., Sandulache, V. C., Hennel, F., Lai, S. Y., & Bankson, J. A. (2013) Radial spectroscopic MRI of hyperpolarized [1-( $^{13}\text{C}$ )] pyruvate at 7 tesla. *Magn Reson Med*, Available Online.
11. Mayer, D., Levin, Y. S., Hurd, R. E., Glover, G. H., & Spielman, D. M. (2006) Fast metabolic imaging of systems with sparse spectra: application for hyperpolarized  $\{^{13}\text{C}\}$  imaging. *Magn Reson Med* 56, 932--7.
12. Josan, S., Hurd, R., Park, J. M., Yen, Y.-F., Watkins, R., Pfefferbaum, A., Spielman, D., & Mayer, D. (2013) Dynamic metabolic imaging of hyperpolarized [2-( $^{13}\text{C}$ )]pyruvate using spiral chemical shift imaging with alternating spectral band excitation. *Magn Reson Med*, Available Online.
13. Leupold, J., Månsson, S., Petersson, J. S., Hennig, J., & Wieben, O. (2009) Fast multiecho balanced SSFP metabolite mapping of ( $^1\text{H}$ ) and hyperpolarized ( $^{13}\text{C}$ ) compounds. *MAGMA* 22, 251-6.
14. Wiesinger, F., Weidl, E., Menzel, M. I., Janich, M. A., Khagai, O., Glaser, S. J., Haase, A., Schwaiger, M., & Schulte, R. F. (2012) IDEAL spiral CSI for dynamic metabolic MR imaging of hyperpolarized [1- $^{13}\text{C}$ ]pyruvate. *Magn Reson Med* 68, 8-16.
15. Gordon, J. W., Niles, D. J., Fain, S. B., & Johnson, K. M. (2013) Joint spatial-spectral reconstruction and k-t spirals for accelerated 2D spatial/1D spectral imaging of ( $^{13}\text{C}$ ) dynamics. *Magn Reson Med*, Available Online.

16. Mugler, 3rd, J. P., Altes, T. A., Ruset, I. C., Dregely, I. M., Mata, J. F., Miller, G. W., Ketel, S., Ketel, J., Hersman, F. W., & Ruppert, K. (2010) Simultaneous magnetic resonance imaging of ventilation distribution and gas uptake in the human lung using hyperpolarized xenon-129. *Proc Natl Acad Sci U S A* 107, 21707-12.
17. von Morze, C., Reed, G., Shin, P., Larson, P. E. Z., Hu, S., Bok, R., & Vigneron, D. B. (2011) Multi-band frequency encoding method for metabolic imaging with hyperpolarized [1-(13)C]pyruvate.. *J Magn Reson* 211, 109--113.
18. Reed, G. D., Larson, P. E. Z., von Morze, C., Bok, R. A., Lustig, M., Kerr, A. B., Pauly, J. M., Kurhanewicz, J., & Vigneron, D. B. (2012) A Method for Simultaneous Echo Planar Imaging of Hyperpolarized 13C Pyruvate and 13C Lactate. *J Magn Reson* 217, 41-7.
19. Cunningham, C. H., Chen, A. P., Lustig, M., Hargreaves, B. A., Lupo, J., Xu, D., Kurhanewicz, J., Hurd, R. E., Pauly, J. M., Nelson, S. J., & Vigneron, D. B. (2008) Pulse sequence for dynamic volumetric imaging of hyperpolarized metabolic products. *J Magn Reson* 193, 139-146.
20. Lau, A. Z., Chen, A. P., Ghugre, N. R., Ramanan, V., Lam, W. W., Connelly, K. A., Wright, G. A., & Cunningham, C. H. (2010) Rapid multislice imaging of hyperpolarized 13C pyruvate and bicarbonate in the heart. *Magn Reson Med* 64, 1323-31.
21. Lau, A. Z., Chen, A. P., Hurd, R. E., & Cunningham, C. H. (2011) Spectral--spatial excitation for rapid imaging of DNP compounds. *NMR in Biomedicine* , n/a--n/a.
22. Sukumar, S., Larson, P. E. Z., Keshari, K. R., Kurhanewicz, J., & Vigneron, D. B. (2010) Single Shot, Chemical Shift Specific Imaging Methods for Hyperpolarized Carbon-13 Studies at {14T}. *Proc ISMR*, 3268.
23. von Morze, C., Sukumar, S., Reed, G. D., Larson, P. E. Z., Bok, R. A., Kurhanewicz, J., & Vigneron, D. B. (2013) Frequency-specific SSFP for hyperpolarized (13)C metabolic imaging at 14.1 T. *Magn Reson Imaging* 31, 163-70.
24. Zierhut, M. L., Yen, Y.-F., Chen, A. P., Bok, R., Albers, M. J., Zhang, V., Tropp, J., Park, I., Vigneron, D. B., Kurhanewicz, J., Hurd, R. E., & Nelson, S. J. (2010) Kinetic modeling of hyperpolarized 13C1-pyruvate metabolism in normal rats and TRAMP mice. *J Magn Reson* 202, 85-92.
25. Larson, P. E. Z., Bok, R., Kerr, A. B., Lustig, M., Hu, S., Chen, A. P., Nelson, S. J., Pauly, J. M., Kurhanewicz, J., & Vigneron, D. B. (2010) Investigation of Tumor Hyperpolarized [1-13C]-Pyruvate Dynamics using Time-Resolved Multiband RF Excitation Echo-planar MRSI. *Magn Reson Med* 63, 582--591.
26. Xu, T., Mayer, D., Gu, M., Yen, Y.-F., Josan, S., Tropp, J., Pfefferbaum, A., Hurd, R., & Spielman, D. (2011) Quantification of in vivo metabolic kinetics of hyperpolarized pyruvate in rat kidneys using dynamic 13C MRSI. *NMR in Biomedicine* 24, 997--1005.
27. Kazan, S. M., Reynolds, S., Kennerley, A., Wholey, E., Bluff, J. E., Berwick, J., Cunningham, V. J., Paley, M. N., & Tozer, G. M. (2012) Kinetic modeling of hyperpolarized (13) C pyruvate metabolism in tumors using a measured arterial input function. *Magn Reson Med* , Available Online.
28. Smith, M. R., Peterson, E. T., Gordon, J. W., Niles, D. J., Rowland, I. J., Kurpad, K. N., & Fain, S. B. (2012) In vivo imaging and spectroscopy of dynamic metabolism using simultaneous 13C and 1H MRI. *IEEE Trans Biomed Eng* 59, 45-9.
29. Schulte, R. F., Sacolick, L., Deppe, M. H., Janich, M. A., Schwaiger, M., Wild, J. M., & Wiesinger, F. (2011) Transmit gain calibration for nonproton MR using the Bloch-Siegert shift. *NMR Biomed* 24, 1068-72.
30. Lau, A. Z., Chen, A. P., & Cunningham, C. H. (2012) Integrated Bloch-Siegert B<sub>1</sub> mapping and multislice imaging of hyperpolarized <sup>13</sup>C pyruvate and bicarbonate in the heart. *Magn Reson Med* 67, 62-71.
31. Arunachalam, A., Whitt, D., Fish, K., Giaquinto, R., Piel, J., Watkins, R., & Hancu, I. (2009) Accelerated spectroscopic imaging of hyperpolarized {C-13} pyruvate using {SENSE}

parallel imaging. *NMR Biomed* 22, 867--873.

32. Tropp, J., Lupo, J. M., Chen, A. P., Calderon, P., McCune, D., Grafendorfer, T., Ozturk-Isik, E., Larson, P. E., Hu, S., Yen, Y.-F., Robb, F., Bok, R., Schulte, R., Xu, D., Hurd, R., Vigneron, D., & Nelson, S. (2011) Multi-Channel Metabolic Imaging, with {SENSE} reconstruction, of Hyperpolarized {[1-13C]} Pyruvate in a Live Rat at 3.0 tesla on a Clinical {MR} Scanner. *J Magn Reson* 208, 171-177.

33. Ohliger, M. A., Larson, P. E. Z., Bok, R. A., Shin, P., Hu, S., Tropp, J., Robb, F., Carvajal, L., Nelson, S. J., Kurhanewicz, J., & Vigneron, D. B. (2013) Combined parallel and partial fourier MR reconstruction for accelerated 8-channel hyperpolarized carbon-13 in vivo magnetic resonance Spectroscopic imaging (MRSI). *J Magn Reson Imaging* 38, 701-13.

34. Hu, S., Lustig, M., Chen, A. P., Crane, J., Kerr, A., Kelley, D. A. C., Hurd, R., Kurhanewicz, J., Nelson, S. J., Pauly, J. M., & Vigneron, D. B. (2008) Compressed sensing for resolution enhancement of hyperpolarized 13C flyback 3D-MRSI. *J Magn Reson* 192, 258-64.

35. Larson, P. E. Z., Hu, S., Lustig, M., Kerr, A. B., Nelson, S. J., Kurhanewicz, J., Pauly, J. M., & Vigneron, D. B. (2011) Fast dynamic 3D MR spectroscopic imaging with compressed sensing and multiband excitation pulses for hyperpolarized 13C studies. *Magn Reson Med* 65, 610-9.

36. Yen, Y.-F., Le Roux, P., Mayer, D., King, R., Spielman, D., Tropp, J., Butts Pauly, K., Pfefferbaum, A., Vasanawala, S., & Hurd, R. (2010) T2 relaxation times of 13C metabolites in a rat hepatocellular carcinoma model measured in vivo using 13C-MRS of hyperpolarized [1-13C]pyruvate. *NMR Biomed* 23, 414--423.

37. Reed, G., von Morze, C., Bok, R., Koelsch, B., Van Criekinge, M., Smith, K., Shang, H., Larson, P., Kurhanewicz, J., & Vigneron, D. (2013) High Resolution C-13 MRI With Hyperpolarized Urea: Mapping and N-15 Labeling Effects. *IEEE Trans Med Imaging*, Available Online.

38. Kettunen, M. I., Kennedy, B. W. C., Hu, D.-E., & Brindle, K. M. (2013) Spin echo measurements of the extravasation and tumor cell uptake of hyperpolarized [1-(13) C]lactate and [1-(13) C]pyruvate. *Magn Reson Med* 70, 1200-9.

39. Larson, P. E. Z., Kerr, A. B., Reed, G. D., Hurd, R. E., Kurhanewicz, J., Pauly, J. M., & Vigneron, D. B. (2012) Generating Super Stimulated-Echoes in {MRI} and their Application to Hyperpolarized {C-13} Diffusion Metabolic Imaging. *IEEE Trans Med Imaging* 31, 265-275.

40. Schilling, F., Düwel, S., Köllisch, U., Durst, M., Schulte, R. F., Glaser, S. J., Haase, A., Otto, A. M., & Menzel, M. I. (2012) Diffusion of hyperpolarized (13) C-metabolites in tumor cell spheroids using real-time NMR spectroscopy. *NMR Biomed*, Available Online.

41. Koelsch, B. L., Keshari, K. R., Peeters, T. H., Larson, P. E. Z., Wilson, D. M., & Kurhanewicz, J. (2013) Diffusion MR of hyperpolarized (13)C molecules in solution. *Analyst* 138, 1011-4.

42. Larson, P. E. Z., Hurd, R. E., Kerr, A. B., Pauly, J. M., Bok, R. A., Kurhanewicz, J., & Vigneron, D. B. (2013) Perfusion and diffusion sensitive 13C stimulated-echo MRSI for metabolic imaging of cancer. *Magn Reson Imaging* 31, 635-42.

43. Frydman, L. & Blazina, D. (2007) Ultrafast two-dimensional nuclear magnetic resonance spectroscopy of hyperpolarized solutions. *Nat Phys* 3, 415--419.

44. Mishkovsky, M. & Frydman, L. (2008) Progress in Hyperpolarized Ultrafast 2D NMR Spectroscopy. *ChemPhysChem* 9, 2340--2348.

45. Kettunen, M. I., Hu, D., Witney, T. H., McLaughlin, R., Gallagher, F. A., Bohndiek, S. E., Day, S. E., & Brindle, K. M. (2010) Magnetization transfer measurements of exchange between hyperpolarized {[1-13C]}pyruvate and {[1-13C]}lactate in a murine lymphoma. *Magn Reson Med* 63, 872--880.

46. Larson, P. E. Z., Kerr, A. B., Leon Swisher, C., Pauly, J. M., & Vigneron, D. B. (2012) A rapid method for direct detection of metabolic conversion and magnetization exchange with application to hyperpolarized substrates. *J Magn Reson* 225C, 71-80.

47. Swisher, C. L., Larson, P. E. Z., Kruttwig, K., Kerr, A. B., Hu, S., Bok, R. A., Goga, A., Pauly, J. M., Nelson, S. J., Kurhanewicz, J., & Vigneron, D. B. (2013) Quantitative measurement of cancer metabolism using stimulated echo hyperpolarized carbon-13 MRS. *Magn Reson Med*, Available Online.
48. Chen, A. P., Leung, K., Lam, W., Hurd, R. E., Vigneron, D. B., & Cunningham, C. H. (2009) Design of spectral-spatial outer volume suppression RF pulses for tissue specific metabolic characterization with hyperpolarized  $^{13}\text{C}$  pyruvate. *J Magn Reson* 200, 344-8.