

**Course: Diabetes Hybrid Imaging****MR Imaging & Spectroscopy of the Influence of Insulin Resistance**

Chris Boesch, MD & PhD

[chris.boesch@insel.ch](mailto:chris.boesch@insel.ch)

University Bern / SWITZERLAND

**Highlights:**

- Magnetic resonance imaging (MRI) and spectroscopy (MRS) can be used in various organs to evaluate the effect of insulin resistance (IR). While the major target of this course is the musculoskeletal system, the use of MRI/MRS in studies of IR in the liver, heart, pancreas, and for the determination of whole body composition are briefly addressed.
- Skeletal muscle contains two types of lipids that can be distinguished by  $^1\text{H}$ -MRS: intra- (IMCL) and extramyocellular (EMCL) lipids. IMCL are metabolically active molecules in small droplets close to the mitochondria which are related to IR, however, not as a cause but rather as a consequence of an unbalanced lipid metabolism.
- IR affects glucose uptake and thus the replenishment of muscular glycogen which can be followed by  $^{13}\text{C}$ -MRS. Intermediate products like glucose-6-phosphate can be observed by  $^{31}\text{P}$ -MRS.
- $^{31}\text{P}$ -MRS saturation transfer experiments can determine flux through biochemical reactions and results have been interpreted as mitochondrial activity in IR; however, since glycolysis also contributes to the measured flux, the specificity of this method for mitochondrial activity is disputed.
- Recovery of  $^{31}\text{P}$ -phosphocreatine is related to oxidative phosphorylation and thus mitochondrial activity. Also specific for mitochondrial activity is the determination of the flux through the TCA-cycle by the observation of infused  $^{13}\text{C}$ -labeled substances.
- Hyperpolarized  $^{13}\text{C}$  substances are recently used to evaluate the effect of IR on metabolism, currently in particular in liver and heart.

Various reviews provide an extended list of citations dealing with the effect of IR and its observation by MRI/MRS [1-18].

Insulin resistance (IR), the (more epidemiologically motivated) “metabolic syndrome”, and overt diabetes are an interrelated complex of diseases which are affecting lipid- and carbohydrate metabolism. They represent major risk factors for cardiovascular diseases with serious consequences for the patients such as heart failure and cerebral stroke. Meanwhile, the number of affected subjects reaches endemic dimensions resulting also in an enormous threat to our health care systems. Several underlying mechanisms are currently discussed: (a) lipotoxicity, (b), mitochondrial activity (c) inflammation, and (d) oxidative stress [19-26].

MRI/MRS can determine the effect of IR on the metabolism and body composition in various organs. This is particular helpful in organs where biopsies are difficult such as in the liver or in the heart [27-40]. While  $^1\text{H}$ -MRS is accepted as the gold standard for the determination of intrahepatic lipids (IHCL), fat-water-imaging sequences - Dixon and in particular multi-echo-versions – provide also excellent and spatially resolved results. Techniques to measure intramyocardial lipids (ICCL) are demanding and not yet established in many places. Whole-body composition is an established MRI modality which is successfully used to follow the effect of interventions.

Muscular tissue contains two different types of lipid stores, intra- (IMCL) and extra- (EMCL) myocellular lipids which are rather different in many aspects ([1] and refs therein). Thanks to their physical characteristics (EMCL with plate-like structures vs. IMCL in droplets), it is possible to distinguish the two depots in  $^1\text{H}$ -MR spectra. Since IMCL are related to IR,  $^1\text{H}$ -MRS became a valuable alternative to muscle biopsies with subsequent histological or electron-microscopic determination. In studies on IR in skeletal muscle, the  $^1\text{H}$ -MRS based determination of IMCL is among the most frequently used in vivo methods. In particular dietary interventions or lifestyle changes with multiple determinations of IMCL are now done with help of  $^1\text{H}$ -MRS wherever available.

One remaining problem in the determination of IMCL is the separation of the large EMCL resonance from the much smaller IMCL signal, in particular in obese subjects who would be among the most interesting groups for studies of IR. Methods such as spatially highly resolved chemical shift methods and/or two-(spectral)-dimensional spectroscopy [1,15,41] are developed to overcome these limitations and to give insight into lipid composition.

Multinuclear MRS has been used to investigate the uptake of glucose and the synthesis of muscular glycogen ([2] and refs therein). These experiments are nicely showing the possibilities that are generated by the combination of various nuclei, in particular  $^{13}\text{C}$ - and  $^{31}\text{P}$ -MRS.

$^{31}\text{P}$ -MRS saturation transfer is an elegant method to estimate the flux through biochemical reactions [42-48], e.g. creatine kinase or ATP synthase. While these methods have been used to determine mitochondrial activity - based on the fact that the aerobic synthesis of ATP is located in these organelles – it has been argued that it is also influenced by the activity of ATP synthase in the glycolytic pathway, thus reducing the specificity but not necessarily a potential clinical significance.

Two MRS methods are more specific for mitochondrial activity, the infusion of  $^{13}\text{C}$ -labeled substances with an observation of  $^{13}\text{C}$ -glutamate following the flux through the TCA-cycle and the recovery of  $^{31}\text{P}$ -phosphocreatine ( $^{31}\text{P}$ -PCr) after exercise. While PCr-recovery is a widely used MRS-method in other diseases and physiological conditions of the musculoskeletal system (see refs in [2]), it is much less applied in IR [49-51]. Higher magnetic fields with increased signal-to-noise allow nowadays at reduction of selected volumes and thus muscle specific observation [52,53]. The application of labeled substances is a very powerful technique, however, requires a lot of experience and generates considerable costs.

Infusion of hyperpolarized  $^{13}\text{C}$ -substances has been strongly promoted for imaging purposes, e.g. the visualization of the ischemic heart wall etc. Beside these applications, hyperpolarized substances are also metabolized and can be used to determine the effect of IR, so far mostly in the liver and the heart [54-57].

MRI and in particular MRS are well suited to study the effect of IR on various organs. In the musculoskeletal system, biopsy is an alternative with additional information content, e.g. molecular biology examinations of the tissue. Nonetheless, the non-invasiveness of MR is a strong argument in particular for repeated measurements. In other organs than skeletal muscle such as liver or heart, MR has even more valuable arguments in the competition with biopsy which is limited to severe clinical situations. Further development of non-invasive MR methods for studies of IR is crucial yet it is not sufficient. Another obstacle for the application of MRI and MRS in studies of IR is the fact that the involved clinicians in endocrinology, diabetology, hepatology, sports medicine, and many other clinical specialties usually have no MR system available. Therefore, it is mandatory that radiological and biomedical MR groups support these clinicians methodologically in the application of MRI/MRS in collaborative studies of IR.

**Reviews and selected recent papers:**

1. Boesch C, Machann J, Vermathen P, Schick F. Role of proton MR for the study of muscle lipid metabolism. *NMR Biomed* 2006;19:968-988.
2. Boesch C. Musculoskeletal Spectroscopy. *J Magn Reson Imaging* 2007;25:321-338.
3. Chang G, Wang L, Cardenas-Blanco A, Schweitzer ME, Recht MP, Regatte RR. Biochemical and physiological MR imaging of skeletal muscle at 7 tesla and above. *Semin Musculoskelet Radiol* 2010;14:269-278.
4. Conley KE, Amara CE, Jubrias SA, Marcinek DJ. Mitochondrial function, fibre types and ageing: new insights from human muscle in vivo. *Exp Physiol* 2007;92:333-339.
5. Costa AF, Di Primio GA, Schweitzer ME. Magnetic resonance imaging of muscle disease: a pattern-based approach. *Muscle Nerve* 2012;46:465-481.
6. Damon BM, Louie EA, Sanchez OA. Physiological basis of muscle functional MRI. *J Gravit Physiol* 2007;14:85-88.
7. Han W, Chuang KH, Chang YT, et al. Imaging metabolic syndrome. *EMBO Mol Med* 2010;2:196-210.
8. Johnson ML, Robinson MM, Nair KS. Skeletal muscle aging and the mitochondrion. *Trends Endocrinol Metab* 2013;24:247-256.
9. Lanza IR, Nair KS. Mitochondrial metabolic function assessed in vivo and in vitro. *Curr Opin Clin Nutr Metab Care* 2010;13:511-517.
10. Machann J, Stefan N, Schick F. <sup>1</sup>H MR spectroscopy of skeletal muscle, liver and bone marrow. *Eur J Radiol* 2008;67:275-284.
11. Noseworthy MD, Davis AD, Elzibak AH. Advanced MR imaging techniques for skeletal muscle evaluation. *Semin Musculoskelet Radiol* 2010;14:257-268.
12. Pedersen BL, Baekgaard N, Quistorff B. Muscle mitochondrial function in patients with type 2 diabetes mellitus and peripheral arterial disease: implications in vascular surgery. *Eur J Vasc Endovasc Surg* 2009;38:356-364.
13. Petersen KF, Shulman GI. New insights into the pathogenesis of insulin resistance in humans using magnetic resonance spectroscopy. *Obesity* 2006;14 Suppl 1:34S-40S.
14. Phielix E, Mensink M. Type 2 diabetes mellitus and skeletal muscle metabolic function. *Physiol Behav* 2008;94:252-258.
15. Pola A, Sadananthan SA, Yaligar J, et al. Skeletal muscle lipid metabolism studied by advanced magnetic resonance spectroscopy. *Prog NMR Spectroscopy* 2012;65:66-76.
16. Schrauwen-Hinderling VB, Roden M, Kooi ME, Hesselink MK, Schrauwen P. Muscular mitochondrial dysfunction and type 2 diabetes mellitus. *Curr Opin Clin Nutr Metab Care* 2007;10:698-703.
17. Subhawong TK, Wang X, Durand DJ, et al. Proton MR spectroscopy in metabolic assessment of musculoskeletal lesions. *AJR Am J Roentgenol* 2012;198:162-172.
18. Wells GD, Noseworthy MD, Hamilton J, Tarnopolski M, Tein I. Skeletal muscle metabolic dysfunction in obesity and metabolic syndrome. *Can J Neurol Sci* 2008;35:31-40.
19. Despres JP, Lemieux I. Abdominal obesity and metabolic syndrome. *Nature* 2006;444:881-887.
20. Van Gaal LF, Mertens IL, De Block CE. Mechanisms linking obesity with cardiovascular disease. *Nature* 2006;444:875-880.
21. Eckel RH, Grundy SM, Zimmet PZ. The metabolic syndrome. *Lancet* 2005;365:1415-1428.
22. Schrauwen P, Hesselink MK. Oxidative capacity, lipotoxicity, and mitochondrial damage in type 2 diabetes. *Diabetes* 2004;53:1412-1417.

23. Bosma M, Kersten S, Hesselink MK, Schrauwen P. Re-evaluating lipotoxic triggers in skeletal muscle: relating intramyocellular lipid metabolism to insulin sensitivity. *Prog Lipid Res* 2012;51:36-49.
24. Samuel VT, Shulman GI. Mechanisms for insulin resistance: common threads and missing links. *Cell* 2012;148:852-871.
25. Schrauwen P, Schrauwen-Hinderling V, Hoeks J, Hesselink MK. Mitochondrial dysfunction and lipotoxicity. *Biochim Biophys Acta* 2010;1801:266-271.
26. Nagle CA, Klett EL, Coleman RA. Hepatic triacylglycerol accumulation and insulin resistance. *J Lipid Res* 2009;50 Suppl:S74-S79.
27. Bley TA, Wieben O, Francois CJ, Brittain JH, Reeder SB. Fat and water magnetic resonance imaging. *J Magn Reson Imaging* 2010;31:4-18.
28. Cobbold JF, Patel D, Taylor-Robinson SD. Assessment of inflammation and fibrosis in non-alcoholic fatty liver disease by imaging-based techniques. *J Gastroenterol Hepatol* 2012;27:1281-1292.
29. Holloway CJ, Suttie J, Dass S, Neubauer S. Clinical cardiac magnetic resonance spectroscopy. *Prog Cardiovasc Dis* 2011;54:320-327.
30. Hu HH, Nayak KS, Goran MI. Assessment of abdominal adipose tissue and organ fat content by magnetic resonance imaging. *Obes Rev* 2011;12:e504-e515.
31. Reeder SB, Cruite I, Hamilton G, Sirlin CB. Quantitative assessment of liver fat with magnetic resonance imaging and spectroscopy. *J Magn Reson Imaging* 2011;34:729-749.
32. Holloway C, Clarke K. Is MR spectroscopy of the heart ready for humans? *Heart Lung Circ* 2010;19:154-160.
33. Schwenzer NF, Springer F, Schraml C, Stefan N, Machann J, Schick F. Non-invasive assessment and quantification of liver steatosis by ultrasound, computed tomography and magnetic resonance. *J Hepatol* 2009;51:433-445.
34. Fischbach F, Bruhn H. Assessment of in vivo <sup>1</sup>H magnetic resonance spectroscopy in the liver: a review. *Liver Int* 2008;28:297-307.
35. Szczepaniak LS, Victor RG, Orci L, Unger RH. Forgotten but not gone: the rediscovery of fatty heart, the most common unrecognized disease in America. *Circ Res* 2007;101:759-767.
36. Weiss K, Martini N, Boesiger P, Kozerke S. Metabolic MR imaging of regional triglyceride and creatine content in the human heart. *Magn Reson Med* 2012;68:1696-1704.
37. van der Meer RW, Doornbos J, Kozerke S, et al. Metabolic imaging of myocardial triglyceride content: reproducibility of <sup>1</sup>H MR spectroscopy with respiratory navigator gating in volunteers. *Radiology* 2007;245:251-257.
38. Buehler T, Ramseier N, Machann J, Schwenzer NF, Boesch C. Magnetic resonance imaging based determination of body compartments with the versatile, interactive sparse sampling (VISS) method. *J Magn Reson Imaging* 2012;36:951-960.
39. Heni M, Machann J, Staiger H, et al. Pancreatic fat is negatively associated with insulin secretion in individuals with impaired fasting glucose and/or impaired glucose tolerance: a nuclear magnetic resonance study. *Diabetes Metab Res Rev* 2010;26:200-205.
40. Machann J, Thamer C, Stefan N, et al. Follow-up whole-body assessment of adipose tissue compartments during a lifestyle intervention in a large cohort at increased risk for type 2 diabetes. *Radiology* 2010;257:353-363.
41. Srikanthan P, Singhal A, Lee CC, et al. Characterization of Intra-myocellular Lipids using 2D Localized Correlated Spectroscopy and Abdominal Fat using MRI in Type 2 Diabetes. *Magn Reson Insights* 2012;5:29-36.

42. De Feyter HM, van den Broek NM, Praet SF, Nicolay K, van Loon LJ, Prompers JJ. Early or advanced stage type 2 diabetes is not accompanied by in vivo skeletal muscle mitochondrial dysfunction. *Eur J Endocrinol* 2008;158:643-653.
43. Befroy DE, Petersen KF, Rothman DL, Shulman GI. Assessment of in vivo mitochondrial metabolism by magnetic resonance spectroscopy. *Methods Enzymol* 2009;457:373-393.
44. From AH, Ugurbil K. Standard magnetic resonance-based measurements of the Pi-->ATP rate do not index the rate of oxidative phosphorylation in cardiac and skeletal muscles. *Am J Physiol* 2011;301:C1-11.
45. Kemp GJ, Brindle KM. What Do Magnetic Resonance-Based Measurements of Pi->ATP Flux Tell Us About Skeletal Muscle Metabolism? *Diabetes* 2012;61:1927-1934.
46. Parasoglou P, Xia D, Chang G, Convit A, Regatte RR. Three-dimensional mapping of the creatine kinase enzyme reaction rate in muscles of the lower leg. *NMR Biomed* 2013;26:1142-1151.
47. Szendroedi J, Schmid AI, Meyerspeer M, et al. Impaired mitochondrial function and insulin resistance of skeletal muscle in mitochondrial diabetes. *Diabetes Care* 2009;32:677-679.
48. Brehm A, Krssak M, Schmid AI, Nowotny P, Waldhausl W, Roden M. Acute elevation of plasma lipids does not affect ATP synthesis in human skeletal muscle. *Am J Physiol* 2010;299:E33-E38.
49. Sleigh A, Raymond-Barker P, Thackray K, et al. Mitochondrial dysfunction in patients with primary congenital insulin resistance. *J Clin Invest* 2011;121:2457-2461.
50. van Elderen SG, Doornbos J, van Essen EH, et al. Phosphorus-31 magnetic resonance spectroscopy of skeletal muscle in maternally inherited diabetes and deafness A3243G mitochondrial mutation carriers. *J Magn Reson Imaging* 2009;29:127-131.
51. Valkovic L, Ukropcova B, Chmelik M, et al. Interrelation of P-MRS metabolism measurements in resting and exercised quadriceps muscle of overweight-to-obese sedentary individuals. *NMR Biomed* 2013;[Epub ahead of print]:doi: 10.1002/nbm.3008.
52. Meyerspeer M, Robinson S, Nabuurs CI, et al. Comparing localized and nonlocalized dynamic 31P magnetic resonance spectroscopy in exercising muscle at 7 T. *Magn Reson Med* 2012;68:1713-1723.
53. Parasoglou P, Xia D, Chang G, Regatte RR. Dynamic three-dimensional imaging of phosphocreatine recovery kinetics in the human lower leg muscles at 3T and 7T: a preliminary study. *NMR Biomed* 2013;26:348-356.
54. Lee P, Leong W, Tan T, Lim M, Han W, Radda GK. In vivo hyperpolarized carbon-13 magnetic resonance spectroscopy reveals increased pyruvate carboxylase flux in an insulin-resistant mouse model. *Hepatology* 2013;57:515-524.
55. Kohler SJ, Yen Y, Wolber J, et al. In vivo 13 carbon metabolic imaging at 3T with hyperpolarized 13C-1-pyruvate. *Magn Reson Med* 2007;58:65-69.
56. Gallagher FA, Kettunen MI, Brindle KM. Biomedical applications of Hyperpolarized 13C Magnetic Resonance Imaging. *Prog NMR Spectroscopy* 2009;55:285-295.
57. Merritt ME, Harrison C, Sherry AD, Malloy CR, Burgess SC. Flux through hepatic pyruvate carboxylase and phosphoenolpyruvate carboxykinase detected by hyperpolarized 13C magnetic resonance. *Proc Natl Acad Sci USA* 2011;108:19084-19089.