

Deoxyhemoglobin and hypercapnia based fMRI calibration methods

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Introduction: Stimulus-induced BOLD signal change is dependent on changes of the brain cerebral blood flow (CBF), venous cerebral blood volume (vCBV) and cerebral metabolic rate of oxygen (CMRO₂), as well as baseline physiological parameters (e.g. hematocrit level, baseline CBV and oxygen extraction fraction, OEF)¹. There are generally two methods of fMRI calibration for determining the maximal BOLD signal change from baseline: deoxyhemoglobin(dHb)-based R2' and CBF/CBV based hypercapnia or hyperoxia methods. The individual baseline physiological parameters needed for hypercapnia calibration involve baseline CMRO₂, reported by Jain et al to vary by ~8%, together with other unknown physiological parameters such as vascular geometries and OEF.² The advantage of using R2', in comparison to hypercapnia/hyperoxia, includes simple experimental setup, and no need for baseline physiological parameter estimation or assumptions about the metabolic response of the stimulus. However, R2' calibration might be complicated by non-dHb related field inhomogeneity sources such as brain iron. The purpose of this study was to compare the calibration parameter, M, using the two methods, and to establish a possible scaling factor between them.

Methods: For dHB based calibration, an asymmetric spin echo (ASE) pulse sequence³ using a segmented EPI with 33 lines of phase space were obtained for each excitation (i.e., EPI factor=33) at 3T field strength, with TE=40 ms, and 180° (sinc) echo offsets (t_e) equal to 0, -4ms, and -16 ms (negative signs indicating a reduction in the interval between the refocusing pulse and the initial 90° excitation pulse). Scan parameters include: FOV: 220x220 mm², matrix size: 128x128 mm², slice thickness: 1.7 mm, yielding an isotropic resolution of 1.7 mm, and TR: 3420 ms to obtain a total of 48 axial slices. The acquisition was repeated four times to facilitate error estimation. R2' maps were obtained with a quadratic exponential fitting⁴, given the short echo offset at 3T. The macroscopic (i.e. large scale, greater than voxel size) magnetic field inhomogeneity (yielding R2'_{macro}) was obtained from the phase difference between two successive echo offsets. Maps of R2' and corrected R2' (termed R2'_{meso} to express that the relaxation rate due to mesoscopic inhomogeneities caused by venous blood in the microvasculature) were scaled by TE to derive the calibration factor M_{R2'} based on the equation $M_{R2'} = TE \cdot k \cdot V_0 \cdot [dHb]_0^\beta = TE \cdot R2' [1]$. Hypercapnia based calibration involved the subject breathing both room air and 5% CO₂ in 95% room air. CBF and BOLD images were used for calibration assuming that mild hypercapnia does not change CMRO₂. BOLD data were acquired with a gradient-echo EPI sequence (TR/TE: 2 sec/25 msec, FA: 70°, FOV: 220 x 220 mm², matrix size: 74 x 74), and pseudo-continuous ASL (pCASL)⁵ based on 2D EPI (TR/TE: 3.9 sec/17 msec, FA: 90°, 1.5 sec labeling duration and 1.23 sec of post label delay) was implemented for CBF quantification. M_{hypercapnia} was calculated based on Davis' biophysical model⁶. As of this publication, sixteen healthy subjects have performed the hypercapnia study (mean age: 37.8 ± 14.3 years) and three subjects for the dHB study.

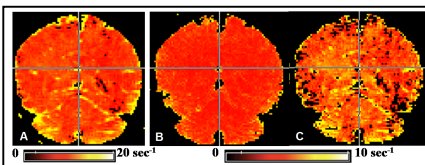


Figure 1. A: R2' map based on ASE sequence using quadratic exponential fitting, B: macroscopic magnetic field inhomogeneity map derived from phase images with larger variations around tissue boundaries and vessels. C: Corrected R2'_{meso} map obtained by subtraction of B from original R2' data.

Results: R2' and corrected R2' are shown in Figure 1A-C for a representative subject. The whole-brain calibration M-value based on R2' were 5.4%, 7.8%, and 8.6% respectively for three subjects. As expected, the M-values based on R2'_{meso} (M_{R2'}) after magnetic field inhomogeneity correction were lower (4.3±1.3%, 5.9±1.7% and 7.8±2.0% respectively).

BOLD and CBF based cerebral vascular reactivity (CVR) under hypercapnia as well as calibrated M of a representative subject is shown in Figure 2. The mean group

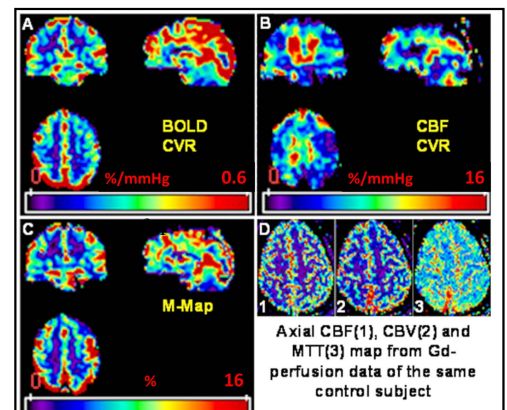


Figure 2. A) BOLD CVR map (in units of %/mmHg) of a representative subject; B) pCASL CVR map from CBF (%/mmHg) in the same subject. C) Relative deoxyhemoglobin concentration in % (i.e. calibration factor M)-map obtained by combining and fractional BOLD and CBF signal change based on Davis model with constant α-β = -1.12. D) Dynamic gadolinium (Gd)-contrast perfusion based CBF and CBV, mean transit time (MTT) maps with slightly greater MTT in white matter and similar CBF values to pCASL.

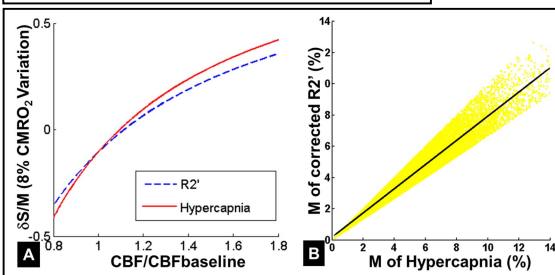


Figure 3. A: Simulation of CMRO₂ variation (normal ~8%) on the calibration methods (blue: R2', red: hypercapnia). B: Scaling factor derived by linear fitting of M_{hypercapnia} to M_{R2'} with a slope of 0.775, and intercept of 0.192 (black line). The slope was 0.805 for a constrained zero intercept.

M_{hypercapnia} calculated based on whole-brain CBF and BOLD signal change was 6.0±0.6%. A negative trend between the calibration parameter M_{hypercapnia} and subject age was observed (r=-0.45, P=0.08). Comparison of R2' and hypercapnia-based methods with simulation shows a possible underestimation of the M factor derived by the R2' method to achieve the same BOLD signal changes, given the normal variation of 8% in baseline CMRO₂ (Figure 3A). Voxel-wise scatter plot of two M-values are shown in Figure 3B, with an estimated up-scaling factor of 1.242 (=1/0.805) for the M_{R2'}.

Discussion and Conclusions: Our preliminary results show comparable calibration M values using R2' and hypercapnia methods, with slight underestimation of M_{R2'} relative to M_{hypercapnia}. One potential limitation of the R2' method is that it assumes deoxyhemoglobin as the dominant source of paramagnetism, which needs further inhomogeneity correction. Simulations suggest a quantitative scaling factor might be useful for applying the RF reversible faster R2' calibration method alone at the same magnetic field strength.

References: [1] Blockley et al NMR Biomed 2013; 26:987-1003. [2] Jain et al. JCBFM 2011;31(7):1504-12. [3] Jensen et al. Magn Reson Med 2009; 61:481-5. [4] Yablonskiy and Haacke. MRM 1994; 32:749-63. [5] Wang et al. MR Imag 2008;26(2):261-69. [6] Davis et al. PNAS 1998;95(4):1834-1839.

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