

BEYOND EDEMA: MYOCARDIAL T₂ IN CHRONIC MYOCARDIAL INFARCTION SWINE

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Target Audience: Cardiologist, Cardiac MRI scientist, Clinicians

Introduction:

T₂ correlates with pathologic processes within myocardial tissue. Recently, quantitative T₂ mapping has been shown more robust than qualitative clinical T₂-weighted (T₂W) imaging in many diseases though most effort has been directed at acute injury and visualization of edema.^{2,3} Chronic myocardial infarction (MI), can also demonstrate altered T₂, as the fibrotic scar tissue, mainly consisting of collagen, increases the interstitial space per unit volume.¹ Yet there are limited reports of measurement of T₂ in chronic MI.^{4,5} In this work, we explore quantitative T₂ measurement as a way to characterize chronic MI without extraneous contrast agent.

Methods:

Animal Model: Reperfusion MI was induced by 120 min occlusion of the middle left anterior descending coronary artery in N=5 Yorkshire swine (37-45 kg). Imaging was performed 14-16 weeks post-MI. **MRI:** 3D whole-heart free-breathing T₂-mapping,⁶ consisting of interleaved volumes were acquired with T₂Prep TEs = 0, 25, 45 ms (TR/TE 4/1.2 ms, flip angle 18°, 1.25×1.25×5.0 mm³ voxels interpolated to 0.98×0.98×2.5 mm³) was achieved at 3T (Achieva TX, Philips Healthcare). Post-contrast late gadolinium enhancement (LGE) was acquired using a 2D phase sensitive inversion recovery (PSIR)⁷ sequence (TR/TE 5.3/2.6 ms, flip angle 18°, 1.25×1.25×5.0 mm³ voxels interpolated to 0.98×0.98×5 mm³). After *in vivo* MRI acquisitions one of the animal received second contrast injection and was excised. The heart was scanned with T₁-weighted (T₁W) spoiled gradient echo (SPGR) sequence about 1 hr post infusion for infarct identification (TR/TE 12/2.2 ms, flip angle 18°, voxel size 0.25×0.25×0.5 mm³). **Histology:** The heart was fixed and fully preserved in 10% formaldehyde. After preservation, the heart were sliced and photographed and histology with Masson's Trichrome staining was obtained. **Post processing:** 3D T₂ maps were calculated per voxel using linear regression of the log of the signal and poor fits (R²<0.9) were rejected. Representative apical/middle/basal slices were chosen from each case for quantitative comparison. 90 radial segments of each slice (total 1350 segments of each type of images) were made to extract the mean T₂ or signal intensity (SI) from T₂ maps and LGE images. Otsu's method⁸ was used to compute a global threshold which maximized the separability of classes in gray levels to differentiate MI from normal myocardium in LGE. Average T₂ value from infarct and normal area was analyzed by unpaired 2-tailed Student's *t*-test with significance defined as p<0.05.

Results:

Chronic infarct was detected in all animals by LGE. T₂ map showed excellent correlation with the myocardial distribution of infarct as evidenced by elevated T₂ and the correlation with hyper-enhanced infarct area from LGE (Fig 1). T₂ in infarct was significantly higher than that of normal myocardium (62.9±14.1 ms v.s. 46.3±4.7 ms, p = 0.0002) (Fig 2). High spatial resolution T₂ mapping enabled heterogeneity detection in and around the infarct area. Figure 3 showed that collagen penetrated into normal myocardium at the border zone of infarct in T₂ map (Fig 3A and B), which excellently matched with the *ex vivo* T₁W image and histology. Fine fibrosis structure can be appreciated in the zoomed in images (Fig 3 B and C) and the corresponding histology (Fig 3D).

Discussion:

The exact mechanism of myocardial T₂ enhancement in chronic myocardial scar is not completely understood. In scarred myocardium, both water and collagen content have an impact on relaxation times. The relative water content and its distribution impact T₂ measurements. Water in myocardial scar tissue might be also more mobile than those in healthy myocytes.⁹ In addition, there is a positive relationship between collagen content and T₂ measurements in cardiac muscles, in contrary to skeletal muscles and tendons.¹⁰ This might explain at least in part, the differences between the various tissues. It is not known yet, however, if T₂(W) is suitable to differentiate acute and chronic stage in MI. The substance in acute MI tissue is different from chronic fibrosis. Weighted images might be less reliable to differentiate the sophistic status because the accompanying changes of both T₁ and T₂ relaxation properties.¹¹ It was reported that T₂ in acute stage was extremely higher than the chronic stage, the corresponding reference values might be a approach for differentiating acute and chronic MI. Of note, in this study, we were limited to a window time of up to 4 months post MI, though the wound healing process was not completed yet. T₂ elevation mechanism in chronic MI needs to be further elucidated.

Conclusion:

Myocardial T₂ mapping has the potential to noninvasively characterize chronic MI size, location and transmuralty without exogenous contrast agents. The observed changes in T₂ may generally apply to any collagenous scar tissue, an important idea since collagen, though present in relatively small amounts even in normal tissue, strongly influences cardiac function.¹² Hence, native T₂ is a promising imaging probe to assist the further understanding on the myocardial tissue characterization.

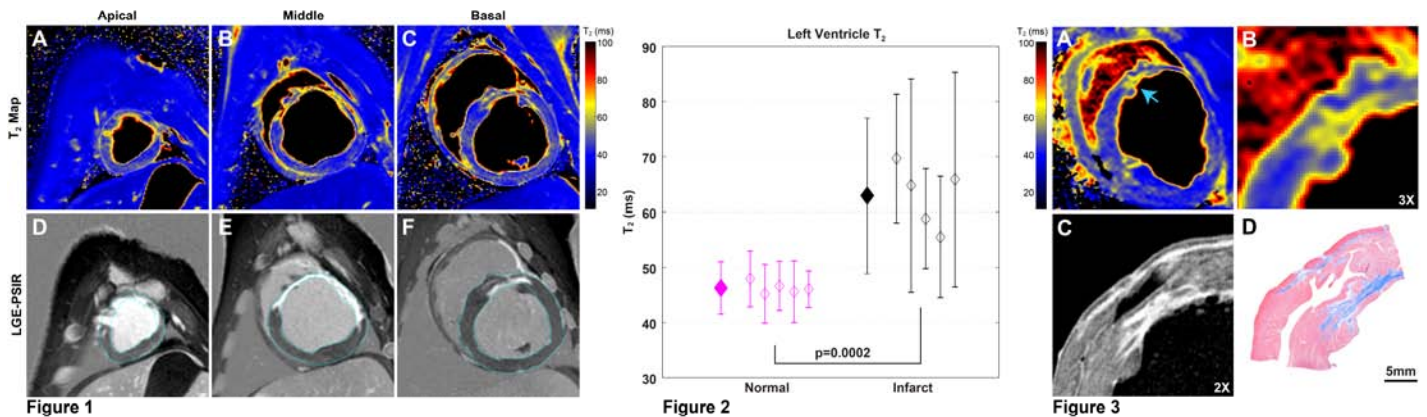


Figure 1: Representative T₂ map (A-C) and corresponding LGE-PSIR (D-F) for three short axis slices from apex and mid-cavity and base.

Figure 2: Comparison on the T₂ between normal myocardium and infarct identified from corresponding LGE. Unpaired 2-tailed Student's *t*-test result shows significant difference between the average T₂ from normal myocardium and infarct. Solid symbols denote group averages.

Figure 3: Comparison of T₂ map (A) with zoomed inside (blue arrow, B) acquired *in vivo* on an animal 4 months post MI with equivalent slices from *ex vivo* T₁W SPGR (C) and Masson's Trichrome stain (D). Note correspondence of collagen deposition as visible on D with the increased T₂ value observed with A and B.

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References: [1] Lima et al. *Circ* 1995 92:1117; [2] Verhaert et al. *JACC Card Img* 2011 4:269; [3] Zia et al. *Circ Card Img*, 2012 5:566; [4] Krauss et al. *Eur J Radiol*. 1990 11:110; [5] Pflugfelder et al. *JACC* 1986 7:843; [6] Ding et al. *ISMRM* 2011; [7] Kellman et al. *MRM* 2002 47:372; [8] Otsu et al. *Automatica*. 1975 11:285; [9] Witschey et al. *MRM* 2010 64:1453; [10] Scholz et al. *Inv. Radiol*. 1989 24:893; [11] Ugander et al. *JACC Card Img* 2012 5:596; [12] Janicki et al. *Adv Exp Med Biol*. 1993 346:291;