High Concentration Gadolinium-Based Contrast Reagent Transverse Relaxivities in *ex vivo* Physiologic Whole Blood and Plasma at 1.5T and 3.0T

Gregory J. Wilson¹, Charles S. Springer, Jr. ², Mark Woods^{2,3}, Sarah Bastawrous^{1,4}, Puneet Bhargava^{1,4}, and Jeffrey H. Maki^{1,4}

¹Radiology, University of Washington, Seattle, WA, United States, ²Advanced Imaging Research Center, Oregon Health and Science University, Portland, OR, United States, ³Chemistry, Portland State University, Portland, OR, United States, ⁴Radiology, Puget Sound VA HCS, Seattle, WA, United States

Introduction: Accurate characterization of contrast reagent (CR) relaxivity in arterial whole blood is necessary for optimization of contrast enhanced MR angiography (CE-MRA) [1]. To investigate relevant relaxivities, we have separately titrated approved CRs that do [gadobenate dimeglumine/MultiHance (MH) and gadofosveset trisodium/Ablavar (AB)] and do not [gadoteridol/ProHance (PH) and gadobutrol/Gadovist (GV)] interact with albumin in whole blood plasma. Limited 1.5 and 3.0T $^1\text{H}_2\text{O}$ T₂ and T₂* values [2] and some T₁ values [3] have been reported previously. This work explores the underlying mechanism of fast T₂* relaxation in oxygenated whole blood.

<u>Methods:</u> Measurements were performed on both 1.5T and 3.0T Achieva MRI scanners (Philips Healthcare, the Netherlands). The phantom consisted of two trays, each with 35 six mL (13×55 mm) HDPE tubes embedded in 2% agar gel. These were filled with fresh, whole blood at 99% O₂ saturation, physiologic pH, 3.3 g/dL albumin, 36% hematocrit, held at 37 °C, and periodically agitated to prevent RBC settling. CR was added to make up [CR_T] values of 1, 2, 3, 4, 5, 6, 8, 10, 14, and 18 mM {mmol(CR)/L(blood)}. 1 H₂O T₂ and T₂* values were measured in whole blood using a multi-echo TSE [TR/ Δ TE/#TE = 2000/6.7/32], and multi-echo FFE [TR/TE/ Δ TE/ α /NSA/#TE = 200/1.3/1.7/35°/4/32], respectively, allowing simultaneous measurement of all samples in the phantom. ROI signal intensity data was fitted with a mono-exponential decay curve using non-linear least squares (Matlab, Natick, MA). After 6 hours of settling, the measurements were repeated for the plasma supernatants.

Results: Relaxation rate constant $R_2 \equiv 1/T_2$) vs. [CR] is approximately linear in both whole blood and plasma (**Figure 1**). R_2 ', the rate constant for RF-refocusable transverse relaxation ($\equiv R_2^* - R_2$), is highly elevated in whole blood compared to plasma, and approximately twice as fast at 3.0T compared to 1.5T (**Figure 2**). R_2 ' in whole blood is approximately the same for all four contrast agents.

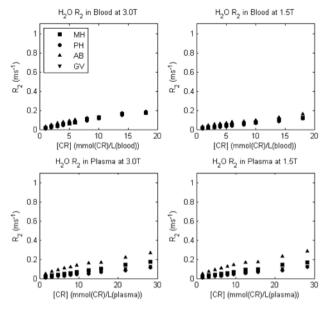


Figure 1. Measured R_2 in oxygenated whole blood (top) and plasma (bottom) at two field strengths. R_2 increases approximately linearly with [CR] in both whole blood and plasma, and exhibits CR-specific relaxivities in plasma.

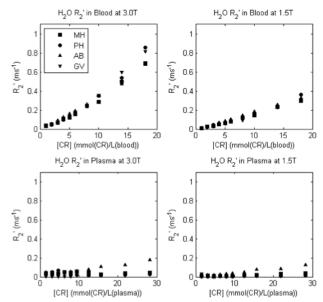


Figure 2. Measured R_2 ' (difference between R_2 * and R_2) in oxygenated whole blood (top) and plasma (bottom) at two field strengths. R_2 ' in whole blood is very high, appears to have super-linear dependence on [CR], and increases linearly with field strength.

Discussion: The fast RF-refocusable transverse relaxation suggests static dephasing is the dominant mechanism in whole blood. This is likely a result of the exclusion of CR from red blood cells (RBC) leading to microscopic magnetic field inhomogeneities; CR in the plasma space creates a bulk magnetic susceptibility (BMS) difference between RBCs and the surrounding plasma. While dipole-dipole ("hyperfine") interactions likely dominate the non-refocusable (R_2) relaxation and show plasma relaxivity variation between CRs, R_2 ' data show very little difference for the various CRs. BMS depends much more on the concentration of CR than on its chemical nature. BMS frequency shifts ($\Delta\omega$) are constant in dimensionless ppm units, but spin dephasing depends on the absolute $\Delta\omega$ value: hence the greater effect at 3T. Monte Carlo simulations have predicted dephasing due to the microscopic BMS inhomogeneities (including variation in blood oxygenation) [4]. They modeled RBCs as 3 μm radius spheres (of low BMS; when fully oxygenated) surrounded by (the high BMS) CR-containing plasma. The water molecules diffused through the resulting microscopic plasma inhomogeneities (accounting for $\Delta\omega$ "motional narrowing"), and the accumulated phase variations resulted in signal loss. The model predicted a quadratic [CR]-dependence of R_2 ' (with minimum corresponding to BMS "matching" of deoxygenated RBC with the surrounding CR in plasma). Our data follow a similar trend, including R_2 ' relaxivity values on-the-order-of 40 s⁻¹/mM at 3.0T and 20 s⁻¹/mM at 1.5T.

In contrast enhanced MR angiography (CE-MRA), first-pass blood [CR_T] may approach 15 - 20 mM. Large R_2^* may yield diminishing returns for CE-MRA performed at high [CR_T] (*i.e.*, rapid CR injection rates). These results may provide an explanation for diminishing return with increasing CR dose or injection rate, and lead to optimized dosing strategies for CE-MRA.

References: 1. Schneider G, et al., JMRI 26:1020-32 (2007); 2. Wilson GJ, et al., PISMRM 21:4459 (2013); 3. Wilson GJ, et al., PISMRM 21:3066 (2013); 4. Blockley NP, et al., MRM 60:1313-1320 (2008).