

CORTICAL PROTON MR SPECTROSCOPIC IMAGING ABNORMALITIES IN A MACAQUE MODEL OF NEUROAIDS

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TARGET AUDIENCE: HIV Clinicians. HIV/SIV Pathogenesis Researchers. MR Spectroscopy Researchers. The HIV-Infected Population.

PURPOSE: Even as highly-active antiretroviral therapy (HAART) reduces incidence of HIV-dementia, milder forms of HIV-associated neurocognitive disorders (HAND) continue to be a health concern due to complications from chronic infection¹. Previous quantitative MRI studies have shown HIV-related thinning of the cerebral cortex (even amongst those receiving HAART), which correlated with CD4⁺ T-lymphocyte depletion and cognitive deficits². Multivoxel proton-MR spectroscopic imaging (¹H-MRSI) studies, however, have yet to examine substantial portions of cortex. We previously reported unchanged N-acetylaspartate (NAA) in the global gray matter³ (GM) of simian immunodeficiency virus (SIV)-infected rhesus macaques, a well-established model system of HIV-infection. Prior histopathology, however, has revealed neuronal loss specifically in cortical regions⁴. (It is worth noting that our previous global analyses would have averaged out all regional changes.) Consequently, in this post hoc study we test the hypothesis that early cortical SIV-infection is characterized by: (i) neuronal damage, reflected by a decrease in their NAA concentration; and (ii) glial activation, marked by increased myo-inositol (mI), choline (Cho) and creatine (Cr)⁵.

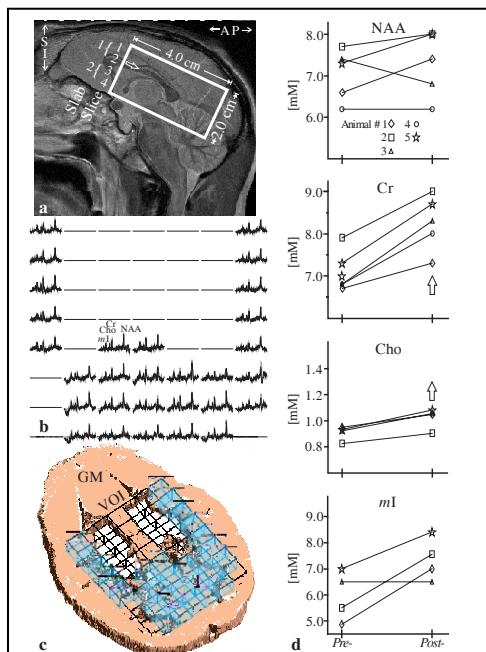


Fig 1. Top, left. **(a)**: Sagittal T₂-weighted MRI of a rhesus macaque head 4 weeks post-SIV infection showing the 3.5×4×2 cm³ VOI (solid lines) placement. Note the open arrow on **a** denotes the level of **b** and **c**. Middle, left. **(b)**: Real part of the 7×8 (LR×AP) ¹H spectra matrix on common chemical shift and intensity scales. Note the SNR and spectral resolution in those (0.5 cm)³ =0.125 cm³ voxels. Bottom, left. **(c)**: 3D view of the cortical partial volumes (beige) in a ¹H-MRSI slice with voxels meeting the ≥90% total volume fraction threshold highlighted (blue). Note the absence of deep GM structures whose voxels' contents were excluded. Right, **(d)**: Line-plots of the NAA, Cr, Cho and mI concentration changes from pre-to-post infection scans for each animal (denoted by symbols in the key). Note the statistically significant increases (open arrows) in mean Cr and Cho, as well as a near-significant elevation in mI.

METHODS: Five (two females; 5.0 to 8.6 kg weight) healthy 3-year-old rhesus macaques (*Macaca mulatta*) were scanned under constant veterinary supervision, as described elsewhere³. Each animal was then inoculated intravenously with SIV_{mac251} virus (10 ng SIVp27) and CD8⁺ T-lymphocyte depleted with CD8-targeted (cM-T807) antibody⁶. Two animals were rescanned 4 weeks and three 6 weeks later. Scans were performed in a whole-body 3 T imager (Magnetom TIM Trio, Siemens AG, Erlangen, Germany) using its transmit-receive knee-coil. Sagittal and axial T₂-weighted turbo spin echo (TSE) MRI: TE/TR = 16/7430 ms were acquired. A 4.0 cm anterior-posterior (AP) × 3.5 cm left-right (LR) × 2.0 cm inferior-superior (IS) =28 cm³ volume-of-interest (VOI) was then image-guided and excited with dual-slab PRESS (TE/TR =33/1440 ms; Fig. 1a). Relative NAA, Cr, Cho and mI levels in the VOI's 224 voxels – obtained with SIToolsFITT package⁷ (Fig. 1b) – were scaled into absolute concentrations against a 0.5 L phantom of known concentrations, as described previously³. Since metabolite concentrations may vary among deep GM structures, cerebellum and cortex, we produced GM and white matter (WM) masks from the axial TSE images, as described elsewhere³. We then carefully erased the striatum, thalamus and cerebellum from GM masks based on a rhesus macaque brain atlas⁸, keeping only voxels with ≥90% total (GM+WM) volume fraction, as shown in Fig. 1c. We then solved for the unknown “cortical” GM metabolites' concentrations in the remaining voxels with linear regression. Since five animals were insufficient for nonparametric tests of metabolic change, paired sample *t* tests were used to assess each metabolite's pre-to-post-infection change. Significance was tested at the *p*<0.05 level using SAS version 9.3 (SAS Institute, Cary, NC).

RESULTS: The ≥90% total voxel tissue constraint left approximately 450 “cortical” GM voxels (~90/animal×5 animals) for analysis. Their mean metabolite SNRs were: NAA=25±8, Cr=16±6, Cho=10±3 and mI=10±4, which led to reliable voxel fits (all Cramer-Rao lower bounds <15%). Mean pre-to-post infection Cr increased 15% (7.2±0.4 to 8.3±0.7 mM, *p*<0.05); Cho 10% (0.9±0.1 to 1.0±0.1 mM, *p*<0.01); and mI 28% (5.8±0.9 to 7.4±0.8 mM, *p*=0.06), as shown in Fig. 1d. NAA was unchanged.

DISCUSSION: Increases in mI, a glia-specific marker⁵, Cho and Cr reflect glial activation in cortex contained within the VOI. Elevated mI is consistent with previous neuropathology in this model⁴, showing widespread elevations of glial fibrillary acidic protein and ionized calcium binding adaptor molecule 1 – an immunohistochemical marker of microglial activation – at four and eight weeks post-infection. Unchanged NAA, however, suggests neuronal cell bodies may be spared early on in the disease. One implication is that astrocyte/microglial activation may precede neuropathogenesis, consistent with evidence previously reported³. It is also possible that WM injury at first or alone may explain cognitive deficits observed⁹. It is noteworthy that due to presence of lipids the VOI was limited to midline/parietal cortex, ~20% of its total.

CONCLUSION: Taken together with previous findings³, these results suggest treatment regimens to reduce gliosis may be helpful in preventing downstream neurodegeneration and cognitive impairment. Provided that further animal testing demonstrates safety, future HIV treatment regimens may benefit from anti-inflammatory drugs aimed at reducing gliosis as a possible therapeutic strategy against HAND.

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