

# Spin-lock MRI of glucose and deoxyglucose concentration changes in brain

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**Target Audience** Researchers and clinicians interested in imaging glucose and deoxyglucose transport and metabolism and/or the CEST technique.

**Purpose** With chemical exchange dependent saturation transfer (CEST) MRI, recent animal studies administering natural D-glucose demonstrated results comparable to PET [1-2], and showed wide potential applications in diseases such as cancer and Alzheimer's. However, this gluco-CEST approach has low sensitivity, and previous studies suggest tissue glucose concentration changes must be ~5-10 mM as a threshold for detection at 9.4 T [3]. Furthermore, the CEST signal is strongly affected by other relaxation effects such as  $T_1$ ,  $T_2$  and magnetization transfer [4-6], and lacks a reliable means to quantify glucose concentration. In this study we showed that the spin-lattice relaxation rate in the rotating frame ( $R_{1\rho}$ ) measured by spin-lock MRI, is highly sensitive to administration of D-glucose and 2-deoxy-D-glucose (2DG) with a much lower detection threshold, and is able to quantify concentration changes.

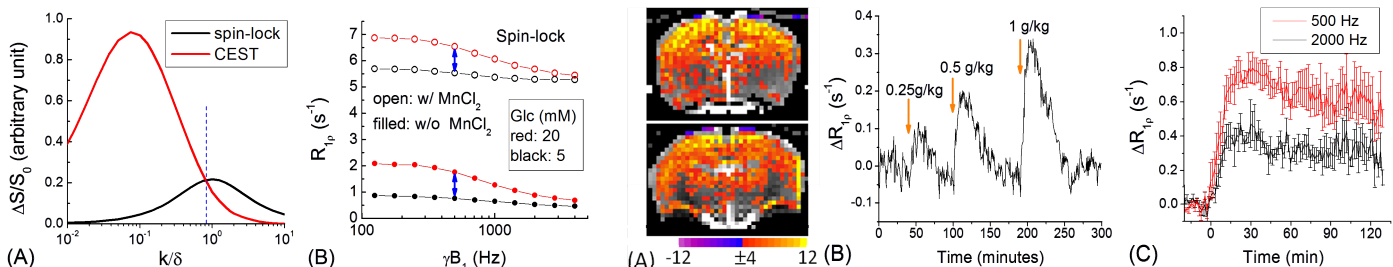
**Materials and methods** *Simulations:* The maximal chemical exchange (CE) contrast was compared by on-resonance spin-lock and CEST approaches for varied exchange rates ( $k$ ). The simulation of Bloch-McConnell equations assumed a chemical shift between water and labile proton of  $\delta = 2500 \text{ rad}\cdot\text{s}^{-1}$  (1 ppm at 9.4 T), a labile proton concentration of 10 mM, and water  $T_1$  and  $T_2$  of 2 s and 50 ms, respectively. Spin-lock contrast was calculated as the difference between  $T_{1\rho}$ -weighted signals with and without CE, normalized by signal without irradiation ( $S_0$ ). CEST contrast was calculated as the difference between  $\text{MTR}_{\text{asym}}(\delta)$  values with and without CE. At each  $k$  value, the maximum contrast of spin-lock and CEST was obtained by adjusting irradiation power and duration.

*MRI experiments:* All MR images were acquired at 9.4 T by single shot spin-echo or gradient-echo EPI with  $0.4 \times 0.4$  or  $0.5 \times 0.5 \text{ mm}^2$  in-plane resolution, 2-mm slice thickness, and a repetition time of 3 s. *Phantom:* 5 and 20 mM of D-glucose with and without addition of 0.1 mM  $\text{MnCl}_2$  were dissolved in phosphate buffered saline titrated to pH = 7.0. Spin-lock  $R_{1\rho}$  dispersion curves (i.e.,  $R_{1\rho}$  vs.  $B_1$ ) were measured at 37°C with spin-lock power  $\gamma B_1 = 125$  to 4000 Hz. *In vivo:* Spin-lock brain studies of Sprague Dawley rats were performed with i.v. injection of glucose and 2DG. *Paradigm 1:* Three consecutive doses of 0.25, 0.5, and 1 g/kg D-glucose ( $n=4$  rats) were given to determine the detection limit of spin lock MRI.  $T_{1\rho}$ -weighted images were measured with and without a spin-lock preparation of  $\gamma B_1 = 500$  Hz for 50 ms duration. *Paradigm 2:* Following 1 g/kg 2DG injection ( $n=4$  rats), three  $T_{1\rho}$ -weighted images, one without spin-lock preparation and two with spin-lock power of  $\gamma B_1 = 500$  and 2000 Hz for 50 ms, were obtained in an interleaved manner to determine  $R_{1\rho}$  dispersion. Time series of  $R_{1\rho}$  maps were calculated from these  $T_{1\rho}$ -weighted images [7], and statistical maps corresponding to glucose injection were determined.

**Results** Figure 1A compares the maximum sensitivity of CEST and spin-lock approaches for different chemical exchange rates. The CEST sensitivity is optimal when  $k/\delta \ll 1$ , but drops quickly when  $k/\delta > 1$ . In contrast, the sensitivity of spin-lock reaches a peak for intermediate chemical exchange rates ( $k/\delta = 1$ ), and is higher than CEST for  $k/\delta > \sim 0.9$  (dashed blue line). Figure 1B shows that  $R_{1\rho}$  changes due to glucose concentration changes (blue arrows) are independent of water  $T_1$  and  $T_2$  (modulations due to  $\text{MnCl}_2$ ). Fitting this  $R_{1\rho}$  dispersion data to a recent theoretical  $R_{1\rho}$  model [8], we found that the exchange rate between water and glucose hydroxyl protons is about  $5000 \text{ s}^{-1}$ , and the averaged chemical shift is  $\delta = 1.6$  ppm, which is  $4000 \text{ rad}\cdot\text{s}^{-1}$  at 9.4 T ( $k/\delta = 1.25$ ), suggesting spin-lock may be a good choice for glucose detection. In the t-map calculated from spin-lock  $R_{1\rho}$  measurement during injection of 0.5 g/kg D-glucose, a widespread increase of  $R_{1\rho}$  is robustly observed in the brain (Fig. 2A). Compared to phantom results (Fig. 1B), the cortical  $R_{1\rho}$  change corresponds to a 1-2 mM increase in brain glucose concentration, indicating high sensitivity of  $R_{1\rho}$  for glucose detection. Figure 2B shows the  $R_{1\rho}$  time course during subsequent injections of three different D-glucose doses, and the dose-dependence of the  $R_{1\rho}$  increases. With 1 g/kg D-glucose, the peak  $\Delta R_{1\rho}$  is  $0.3 \text{ s}^{-1}$  and a 50-60 min to return to baseline, while injection of 1 g/kg 2DG (Fig. 2C), gives a much larger peak  $\Delta R_{1\rho}$  of  $0.75 \text{ s}^{-1}$  and a much slower return to baseline vs. D-glucose. This indicates that 2DG accumulates within the cells, while D-glucose metabolizes quickly [3]. With a higher spin-lock power  $\gamma B_1 = 2000$  Hz,  $\Delta R_{1\rho}$  is much smaller than with 500 Hz, as expected from phantom data (Fig. 1B).

**Discussion** Previous rat brain studies at 9.4 T indicated that gluco-CEST signal changes could not be detected with a 0.5 g/kg D-glucose injection and were very weak even with 1 g/kg injection [3]. Our results show this detection threshold may be significantly lower with the spin-lock approach. This sensitivity advantage may be partly because the hydroxyl-water proton exchange is in the intermediate to fast exchange regime, where spin-lock sensitivity is better vs. CEST. Since  $k/\delta$  increases at lower magnetic fields, this advantage will be more prominent at clinical fields such as 3 T (Fig. 1A). Another important advantage of spin-lock is its insensitivity to any  $B_0$  shift much smaller in magnitude than the spin-lock pulse power, while CEST is highly susceptible to  $B_0$  shifts of only a few Hertz. In addition, the sub-minute temporal resolution of spin-lock MRI (Fig. 2) provides a much higher statistical power for glucose detection as compared to the low temporal resolution of gluco-CEST (~10 minutes in previous studies [1-3]). Unlike  $\text{MTR}_{\text{asym}}$ , which is coupled to other relaxations including  $T_1$ ,  $T_2$  and magnetization transfer effects [4-6] and usually used in CEST, our phantom results indicate that  $R_{1\rho}$  changes are independent of  $T_1$  and  $T_2$ , and provide a quantitative index to glucose concentration changes.

**Conclusion** Our results show that spin-lock MRI is highly sensitive to the administration of D-glucose and 2DG, provides a quantitative index to glucose concentration changes, and therefore may have significant advantages over gluco-CEST for imaging of glucose transport and metabolism.



**Fig. 1.** (A) Bloch-McConnell simulations comparing optimal spin-lock and CEST contrast from 10 mM of labile protons as a function of  $k/\delta$ . (B) Spin-lock  $R_{1\rho}$  dispersion of 5 and 20 mM glucose without and with  $\text{MnCl}_2$ .

**Fig. 2.** Rat brain spin-lock  $R_{1\rho}$  changes. (A) Calculated t-maps (2-slices) are due to 0.5 g/kg D-glucose injection with  $\gamma B_1 = 500$  Hz. Averaged  $R_{1\rho}$  time courses represent cortical responses during (B) subsequent injection of three different D-glucose doses with  $\gamma B_1 = 500$  Hz and (C) during injection of 1 g/kg 2DG with  $\gamma B_1 = 500$  and 2000 Hz.

**References** [1] Walker-Samuel S et al, Nat Med 19:1067 (2013). [2] Chan K W Y et al., MRM 68:1764 (2012). [3] Nasrallah FA et al., JCBFM 33:1270 (2013). [4] Jin T et al., MRM 68:1056 (2012). [5] Sun PZ, MRM 67:936 (2012). [6] Vinogradov E et al., JMR 229:155 (2013). [7] Jin T et al., NeuroImage 78:385 (2013). [8] Trott O et al., JMR 154:157 (2002).