

Exchange and T2-relaxation effects in double pulsed field gradient experiments

Casper Kaae Sønderby¹, Henrik Lundell¹, and Tim B. Dyrby¹

¹Danish Research Centre for Magnetic Resonance, Copenhagen University Hospital, Hvidovre, Denmark

Introduction: Double Pulsed Field Gradient (dPFG) sequences have recently been used for novel tissue contrasts such as the Apparent Exchange Rate (AXR) and Compartment eccentricity^{1,2}. Both stimulated echo (STE) and spin echo (SE) type sequences have been used for these kinds of experiments¹⁻³. However, the effects of differences in compartment specific T1/T2 relaxation between these types of sequences are currently unknown. Here we compare effects on the mixing time (TM) dependent Apparent Diffusion Coefficient (ADC'(TM)) of either T1- or T2-relaxation during TM in dPFG type experiments using two variants of a dPFG SE (dSE) sequence and dPFG STE (dSTE) sequence. Our results indicate that differences in T2-relaxation between signal components originating from compartments with different diffusivities cause a decrease in the estimated ADC as TM or echo time (TE) are increased.

Method: A single scan session was performed in an experimental 4.7 T MRI Agilent scanner with a maximal gradient strength of 600 mT/m. A yeast cell suspension constituting a well-defined isotropic two-compartment system of densely packed cells was used for all scans. Three days prior to scanning fresh baker's yeast was mixed with tap water (3:1 weight ratio). Before scanning, the yeast was centrifuged at 2800 rpm for 15 minutes in 15-mL plastic tube and the tube was positioned in a quadrature RF transmit/receive coil. The following imaging parameters were used: Six slices with no gaps, matrix size: 128x128, voxel size: 1³ mm³, and a TR of 2500 ms. ADC'(TM) was estimated using three types of sequences as shown in figure 1: i) dSTE, ii) dSE with fixed echo time (TE) of 107ms (dSE_{fix}) and iii) dSE with minimal TE increasing with TM (dSE_{var}). How TM was varied depended upon the type of sequence as seen in Fig 1.: For i) the *dSTE sequence* by increasing the delay between the 2nd and 3rd 90°RF pulse, ii) the *dSE_{fix} sequence* by using a fixed TE and increasing the separation between the filter and detection gradients, and iii) the *dSE_{var} sequence* by using the minimum possible TE while increasing the separation between the filter and detection gradients. For all sequences a diffusion weighted image with both the filter and detection block applied and an image with only the filter block applied i.e. a *filtered b₀* image, was acquired and from these the ADC'(TM) was calculated as the regular ADC. The following settings was used: b-value in both filter and detection block: 2720 s/mm² (gradient strength [G]: 490 mT/m, pulse duration (□): 5 ms, pulse separation (□): 8 ms), mixing time (TM): [10, 15, 20, 25, 30, 40, 50, 60, 70] ms. To further demonstrate potential effects of compartment specific T2-relaxation effects, the regular ADC was estimated at different TE using a standard diffusion weighted Pulsed Gradient Spin Echo (PGSE) sequence with the diffusion gradient position right before and after the 180°RF pulse. By increasing the delays before and after the diffusion encoding gradients TE was varied between [23, 40, 60, 80, 100] ms while keeping the diffusion encoding gradients identical for all experiments. The following settings were used: b-value: 1037 s/mm² (G: 300 mT/m, □: 5 ms, □: 8.11 ms). The temperature was kept at 18.4±0.7°C during all scans. The ADC'(TM) and ADC values was calculated using the mean signal from a ROI covering the central part of the yeast phantom.

Results: ADC'(TM) values for the three sequences in Fig. 1 are seen in Fig 2.

For short TM the ADC'(TM) values are comparable for dSE_{var} and dSTE whereas the values of dSE_{fix} are lower. As TM is increased, ADC'(TM) increases in a comparable way for both the dSTE and dSE_{fix} sequences due to exchange processes as previously shown^{1,3}. However, for dSE_{var} at TM 30 ms ADC'(TM) start decreasing with increasing TM and gradually approaches the values of dSE_{fix}, and at TM=70 ms the two measurements are comparable. ADC values measured at different TE using a standard diffusion weighted PGSE sequence are seen in Fig 3. The measured ADC values decreases more than 30% as TE is increased from 23 to 100 ms. The ADC was not expected to be dependent on TE. Note that the higher ADC in Fig. 2 are likely explained by the lower b-values used in the dwSE compared to those used in the dSTE and dSE sequences.

Discussion & Conclusion: We speculate that the decrease of ADC'(TM) in TM is explained by shorter T2-relaxation of compartment specific fast diffusion signal components, often assumed to originate from the extracellular space, effectively filtering the fast diffusing signal components as TE is increased. This explains: 1) dSTE and dSE_{fix} show comparable ADC'(TM) values at short TM where dSE_{fix} is subject to similar T2-relaxation as dSTE, 2) the ADC'(TM) values of dSE_{fix} approaches the lower values of dSE_{var} as TM increased since the T2-relaxation of dSE_{var} increases with TM, and 3) the decrease of ADC as TE is increased with otherwise identical PGSE sequence settings. Previous T2-relaxation studies in rat spinal cord, frog peripheral and frog sciatic nerve have found either the shortest T2-times in extra-axonal compartment⁴, the shortest T2-times in the intra-axonal compartment⁵ or non-identical T2-relaxation between the two compartments⁶. Hence the origin of the multiexponential T2-decays is not well established. If our yeast phantom is interpreted as a simple two-compartment system where the extracellular signal components are suppressed by the filter-gradients and subsequently restored due to exchange processes during TM, our data suggested that the extra-cellular signal component have shorter T2-relaxation compared to the intracellular compartment. Lastly we note that variability of ADC in TE suggests that both dPFG and regular single-shell diffusion weighted experiments with PGSE should consider the effects of differences in compartment specific T2-relaxations in the experimental design. Furthermore our results indicate that multi-shell experiments should adopt the same TE in all shells to maintain the relative T2-relaxation weighting of the individual compartments.

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