

MR Spectroscopy: The promise

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Despite important progresses in the last decades, the development of Magnetic Resonance Spectroscopy (MRS) approaches in the Clinic remains significantly slower than the widespread clinical applications of its imaging counterpart (MRI). Probably the most fundamental advantage of MRS is its chemical specificity, allowing for the non-invasive characterization of metabolite profiles and their changes with time, in normal and diseased tissues of animals and humans. The clinical implementation of this important advantage has been, however, hampered by important limitations in sensitivity and resolution demanding, progressively, drastic technological advances as higher field magnets, improved field homogeneity, advanced volume localization techniques and sophisticated spectral acquisition and processing algorithms. Many of these developments are not presently accessible in most clinical imaging centers, making it difficult to implement successfully the MRS approach widespread in the clinic. These limitations have not hampered, however, the impressive development of MRS approaches in preclinical research, where very high field magnets, specialized personnel and powerful software packages have endowed MRS with a prominent role in the evaluation of animal models of disease, drug development and therapy design.

Probably the most general landmark in the development and outcome of any pathology is the appearance of energy limitations, leading eventually to energy failure and death of the diseased tissues. These circumstances result always in important adaptive metabolic responses that constitute the metabolic fingerprints of the pathological development, as well as of its response to therapy. In the following, we shall support that MRS from different nuclei, is well endowed to detect these metabolic alterations, many times, much earlier than the morphological alterations observable later by MRI.

¹H MRS

Taking the brain as an example, ¹H MRS provides a robust fingerprint of metabolites revealing healthy or diseased metabolism in animal models and humans [1-6]. Relative decreases in N-acetyl-aspartic acid (NAA) are thought to reveal neuronal loss, increases in lactate (Lac) are associated to hypoxia and enhanced glycolitic contribution, lipid resonance (Lip) increases are normally observed in hypoxic tumor areas, alterations in the total creatine resonance (tCr) are thought to represent alterations in energy metabolism, changes the total Choline resonance (tCho) reflect changes in phospholipid metabolism, and alterations in the myo-inositol (Ino) content reveal inflammatory responses and osmolite volume regulation. In the prostate, alterations in the citrate levels have been correlated with malignity of the lesions [7, 8]. In addition to these metabolites, easily detectable even in low field clinical scanners (1,5T), up to fourteen cerebral metabolites may become detectable in higher field scanners, including the amino acids glutamate, glutamine, gaba and taurine among other metabolites. ¹H MRS provides thus, the most comprehensive, non invasive, metabolic profiling method available to date, useful in the diagnosis and prognosis of cancer, neurodegenerative diseases and ischemic episodes.

³¹P MRS

³¹P NMR was the earliest MRS implemented providing direct information on the bioenergetics of skeletal and cardiac muscle, brain and tumors, because of the direct detection of ATP, PCr, Pi, Phosphomester (PME) and Phosphodiester (PDE) resonances. ³¹P NMR was mainly used to investigate cerebral energetics in the healthy brain, as well as in cerebral tumors [9-11]. In the latter sense, ³¹P NMR pioneered the development of the pattern recognition algorithms implemented for the intelligent diagnosis of tumors [12-15]. An important clinical multicenter study merits special mention here, illustrating the diagnostic potential of ³¹P NMR in oncology [16, 17].

¹³C NMR (and hyperpolarization)

¹³C NMR has been probably the slowest MRS approach to be incorporated to the clinic. The low natural abundance of ¹³C required the use of relatively expensive ¹³C enriched isotopes, specialized volume localization and proton decoupling techniques and sophisticated mathematical modeling approaches. Despite its drawbacks, ¹³C NMR provides unique information on cerebral metabolism and its compartmentation, the neuronal and glial tricarboxylic acid cycle rates, and the transcellular glutamate-glutamine or gaba cycles and in vivo neurotransmission [18-22]. Nevertheless, the clinical applications of ¹³C MRS remained limited by the inherently low sensitivity of the technique [23].

The recent advent of ¹³C hyperpolarization strategies has improved spectacularly the previous signal to noise limitations, at the expense of very fast technologies for ¹³C acquisition, observing only the longest relaxation time hyperpolarized ¹³C carboxylic carbons [24-29]. Although mainly implemented in preclinical ¹³C NMR laboratories, the future impact of this technique in the clinic is considered feasible [30], providing information on tumor pH, pyruvate dehydrogenase flux and redox state, with comparable spatial resolution to the nuclear medicine PET or SPECT techniques.

Finally, other nuclei are conveniently available for MRS approaches including mainly ¹⁹F NMR [31] and ¹⁷O NMR [32]

Conclusion

The advance of clinical MRS approaches in the last decades cannot be underscored, but probably remains still insufficient for wide spread, routine, clinical applications except in frontier clinical research centers. The current limitations of clinical MRS appear to be mostly due to economic constraints and educational or translational difficulties, rather than to an insufficient supply of a wealth of relevant diagnostic and prognostic information. On these grounds, the main promise of MRS as the most powerful, specific, non-invasive tool, for personalized diagnosis and treatment remains intact.

Bibliography

1. Lee, M.R., et al., Preclinical (1)H-MRS neurochemical profiling in neurological and psychiatric disorders. *Bioanalysis*, 2012. **4**(14): p. 1787-804.
2. Choi, J.K., A. Dedeoglu, and B.G. Jenkins, Application of MRS to mouse models of neurodegenerative illness. *NMR Biomed*, 2007. **20**(3): p. 216-37.

3. Mountford, C.E., et al., Proton MRS can determine the pathology of human cancers with a high level of accuracy. *Chem Rev*, 2004. **104**(8): p. 3677-704.
4. Jones, R.S. and A.D. Waldman, ¹H-MRS evaluation of metabolism in Alzheimer's disease and vascular dementia. *Neurol Res*, 2004. **26**(5): p. 488-95.
5. Petroff, O.A., R.H. Mattson, and D.L. Rothman, Proton MRS: GABA and glutamate. *Adv Neurol*, 2000. **83**: p. 261-71.
6. Maton, B.M. and R.I. Kuzniecky, Proton MRS: N-acetyl aspartate, creatine, and choline. *Adv Neurol*, 2000. **83**: p. 253-9.
7. Celda, B., et al., MRS as endogenous molecular imaging for brain and prostate tumors: FP6 project "eTUMOR". *Adv Exp Med Biol*, 2006. **587**: p. 285-302.
8. Decelle, E.A. and L.L. Cheng, High-resolution magic angle spinning (¹ H MRS in prostate cancer. *NMR Biomed*, 2014. **27**(1): p. 90-9.
9. Negendank, W. and R. Sauter, Intratumoral lipids in ¹H MRS in vivo in brain tumors: experience of the Siemens cooperative clinical trial. *Anticancer Res*, 1996. **16**(3B): p. 1533-8.
10. Negendank, W., et al., Phospholipid metabolites in ¹H-decoupled ³¹P MRS in vivo in human cancer: implications for experimental models and clinical studies. *Anticancer Res*, 1996. **16**(3B): p. 1539-44.
11. Negendank, W., Studies of human tumors by MRS: a review. *NMR Biomed*, 1992. **5**(5): p. 303-24.
12. Wright, A.J., et al., Pattern recognition of MRSI data shows regions of glioma growth that agree with DTI markers of brain tumor infiltration. *Magn Reson Med*, 2009. **62**(6): p. 1646-51.
13. Maxwell, R.J., et al., Pattern recognition analysis of ¹H NMR spectra from perchloric acid extracts of human brain tumor biopsies. *Magn Reson Med*, 1998. **39**(6): p. 869-77.
14. Tate, A.R., et al., Lipid metabolite peaks in pattern recognition analysis of tumour in vivo MR spectra. *Anticancer Res*, 1996. **16**(3B): p. 1575-9.
15. Howells, S.L., et al., Pattern recognition of ³¹P magnetic resonance spectroscopy tumour spectra obtained in vivo. *NMR Biomed*, 1993. **6**(4): p. 237-41.
16. Arias-Mendoza, F., et al., In vivo ³¹P MR spectral patterns and reproducibility in cancer patients studied in a multi-institutional trial. *NMR Biomed*, 2006. **19**(4): p. 504-12.
17. Arias-Mendoza, F., et al., Methodological standardization for a multi-institutional in vivo trial of localized ³¹P MR spectroscopy in human cancer research. In vitro and normal volunteer studies. *NMR Biomed*, 2004. **17**(6): p. 382-91.
18. Rodrigues, T.B., et al., ¹³C NMR tracers in neurochemistry: implications for molecular imaging. *Q J Nucl Med Mol Imaging*, 2009. **53**(6): p. 631-45.
19. Garcia-Espinosa, M.A., et al., Cerebral glucose metabolism and the glutamine cycle as detected by in vivo and in vitro ¹³C NMR spectroscopy. *Neurochem Int*, 2004. **45**(2-3): p. 297-303.
20. Cruz, F. and S. Cerdan, Quantitative ¹³C NMR studies of metabolic compartmentation in the adult mammalian brain. *NMR Biomed*, 1999. **12**(7): p. 451-62.
21. Rothman, D.L., et al., ¹³C MRS studies of neuroenergetics and neurotransmitter cycling in humans. *NMR Biomed*, 2011. **24**(8): p. 943-57.
22. Ramadan, S., A. Lin, and P. Stanwell, Glutamate and glutamine: a review of in vivo MRS in the human brain. *NMR Biomed*, 2013. **26**(12): p. 1630-46.
23. Ross, B., et al., Clinical experience with ¹³C MRS in vivo. *NMR Biomed*, 2003. **16**(6-7): p. 358-69.
24. Gallagher, F.A., et al., Hyperpolarized ¹³C MRI and PET: in vivo tumor biochemistry. *J Nucl Med*, 2011. **52**(9): p. 1333-6.
25. Brindle, K.M., et al., Tumor imaging using hyperpolarized ¹³C magnetic resonance spectroscopy. *Magn Reson Med*, 2011. **66**(2): p. 505-19.

26. Gallagher, F.A., et al., Magnetic resonance imaging of pH in vivo using hyperpolarized ¹³C-labelled bicarbonate. *Nature*, 2008. **453**(7197): p. 940-3.
27. Day, S.E., et al., Detecting tumor response to treatment using hyperpolarized ¹³C magnetic resonance imaging and spectroscopy. *Nat Med*, 2007. **13**(11): p. 1382-7.
28. Nelson, S.J., et al., Strategies for rapid in vivo ¹H and hyperpolarized ¹³C MR spectroscopic imaging. *J Magn Reson*, 2013. **229**: p. 187-97.
29. Hu, S., et al., ¹³C-pyruvate imaging reveals alterations in glycolysis that precede c-Myc-induced tumor formation and regression. *Cell Metab*, 2011. **14**(1): p. 131-42.
30. Kurhanewicz, J., et al., Current and potential applications of clinical ¹³C MR spectroscopy. *J Nucl Med*, 2008. **49**(3): p. 341-4.
31. Chen, H., et al., ¹⁹F NMR: a valuable tool for studying biological events. *Chem Soc Rev*, 2013. **42**(20): p. 7971-82.
32. Zhu, X.H., et al., In vivo ¹⁷O NMR approaches for brain study at high field. *NMR Biomed*, 2005. **18**(2): p. 83-103.