

# Field dependence of relaxivity of Gd chelates as a function of macromolecular content

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## Introduction

**Target Audience:** Researchers interested in the factors that affect the relaxivity of Gd chelates at high  $B_0$  and in vivo for a variety of applications.

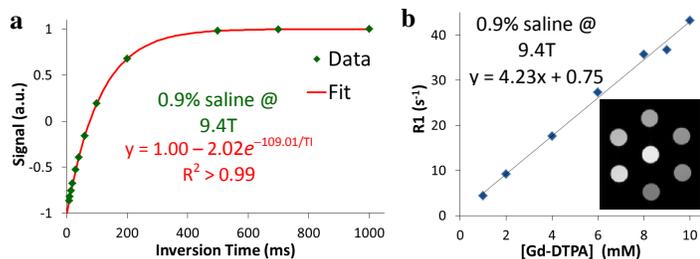
**Purpose:** To measure the  $T_1$  relaxivity of Gd-DTPA as a function of macromolecular content and different high  $B_0$  strengths (up to 15.2T).

Gd-DTPA is a common MRI contrast agent widely used in tumor imaging<sup>1</sup>, MR angiography<sup>2</sup>, modeling hemodynamic behavior<sup>3</sup>, and myocardial perfusion<sup>4</sup>.  $T_1$  relaxivity ( $r_1$ ) is a fundamental property of contrast agents that is important for modeling and quantitative analyses. Measurements of Gd-DTPA  $r_1$  have previously been extensively reported<sup>5,6,7</sup>, including field-cycling measurements over a range of frequencies. However, previous studies did not address the behavior of  $r_1$  at very high fields or how  $r_1$  may be modified in the presence of realistic media to different extents at different fields. The Solomon-Bloembergen-Morgan equations predict a frequency effect that depends on the relevant correlation time(s) of the paramagnetic interaction, which were investigated by simulation studies<sup>8</sup>. Here, we report Gd-DTPA  $r_1$  measurements using the same samples and analysis over a range of high  $B_0$ . We also investigate the  $r_1$  dependence on macromolecular content at different fields, which is known to modulate  $r_1$ <sup>6</sup>.

## Methods

Four sets of solutions were prepared with seven concentrations (0.6–10 mM) of Gd-DTPA (Magnevist, Schering): saline, 3% w/v milk powder in saline, 12% w/v milk powder in saline, 30% w/v milk powder in saline, and 24% w/v homogenized rat brain tissue in saline. All saline solutions had a concentration of 0.9%. The milk powder was fat-free. The brain tissues were harvested from healthy adult Sprague-Dawley rats. The 12% milk powder solution was chosen to approximate the reported protein content in rat brain<sup>9</sup>, while the 3% milk powder solution was chosen to approximate the expected protein content in the 24% homogenized rat brain tissue solution. Each set of solutions were transferred into 5mm NMR tubes and bundled together for MRI experiments.

MRI experiments were run on both Varian (4.7, 7, and 9.4T) and Bruker (15.2T) systems at room temperature (20°C). A single-slice 2D



**Fig 1.** (a) Sample IR three parameter fit ( $y(TI) = a+b \cdot e^{-TI/T_1}$ ) for single pixel. Note fitted  $a$  and  $b$  were close to the expected values 1 and -2, respectively. (b) Sample linear fit to measure  $r_1$  ( $R1 = 1/T_1 = r_1[\text{Gd-DTPA}] + 1/T_1^0$ ) where  $T_1^0$  is the  $T_1$  in the absence of Gd-DTPA. Inset: sample IR-SE image of NMR tube phantom.

inversion-recovery spin-echo (IR-SE) imaging sequence was used to measure  $T_1$  of each Gd-DTPA concentration simultaneously with the following parameters:  $128 \times 128$ , FOV/THK=30/3mm, TR = 1.5s, TE=8ms, and TI=8.5-1400ms. An adiabatic RF pulse was used for inversion. Magnitude images were generated and a phase-based polarity correction was used<sup>10</sup>. Pixel-wise inversion curves were fit to the standard three-parameter model (Fig.1) to generate a  $T_1$  map (Matlab, Mathworks). ROIs were drawn in each NMR tube to measure mean  $T_1$  values and  $r_1$  could be determined by a linear fit (Fig.1).

To estimate the error in  $r_1$  measurements, it was assumed that the dominant error was from volume measurements while preparing the solutions. With our protocol, an error of 0.1ml is reasonable and can be propagated through the various dilutions to calculate the maximum and minimum expected Gd-DTPA concentrations. These values, in turn, can be used to calculate the maximum and minimum expected  $r_1$  values.

## Results and Discussion

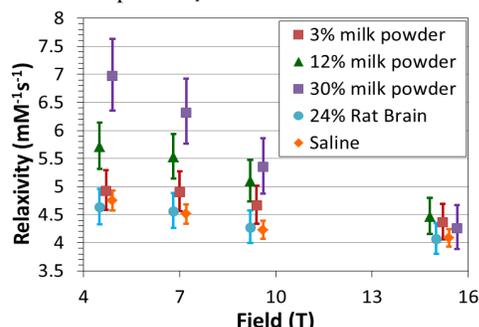
Fig. 1 shows example  $T_1$  and  $r_1$  fits. All fits had  $R^2 > 0.99$ . Note that fitted  $T_1$  in absence of Gd-DTPA ( $T_1^0$ ) for 0.9% saline at 9.4T was 1.34s. Fig. 2 shows plots of measured  $r_1$  vs  $B_0$  strength for various solutions. Error bars for saline  $r_1$  are smallest as saline solutions were prepared first and subsequently used to prepare the other solutions. The  $r_1$  of saline falls between 4–5  $\text{mM}^{-1}\text{s}^{-1}$  at all  $B_0$  which agrees with literature values<sup>5,6</sup>. All solutions show a trend of decreasing  $r_1$  with increasing  $B_0$  matching previous simulation results<sup>8</sup> and consistent with theory.

It is known that increasing macromolecular content increases  $r_1$  presumably due to the increased correlation times of the dipolar interaction of Gd-DTPA with water. At 4.7T,  $r_1$  increased 20% and 50% for 12% and 30% milk powder solutions similar to reported values<sup>6</sup>. Simulation studies have investigated the effect of correlation times on  $r_1$  of Gd-DTPA as a function of  $B_0$ <sup>8</sup>. Under saline solution conditions,  $r_1$  did not show a strong  $B_0$  dependence, while under conditions mimicking large macromolecular content there was a strong  $B_0$  dependence –  $r_1$  increased significantly at lower fields, but not at higher fields. The results in Fig. 2 agree well with the simulation studies and provide experimental confirmation.

The  $r_1$  for 24% rat brain solution and saline were observed to be almost the same at all  $B_0$ . This observation was repeated with 3% milk powder solution, which approximates the protein content in the rat brain solution, suggesting that it was the dilution of the rat brain homogenate that removed any macromolecular effect. 12% milk powder solution, which approximates the protein content in intact rat brain<sup>9</sup>, shows increased  $r_1$  at lower  $B_0$  but not at higher  $B_0$ . In fact, at 15.2T, there was no observed difference in  $r_1$  among all solutions. These results suggest that Gd-DTPA  $r_1$  in rat brain approaches that of Gd-DTPA in saline at higher  $B_0$ .

## Conclusion

This work is a study of Gd-DTPA  $r_1$  as a function of macromolecular content across a range of high  $B_0$ . The results suggest that modification of correlation times by viscosity and binding effects do not increase  $r_1$  at very high fields confirming simulation studies.



**Fig 2.** Plots of  $r_1$  vs  $B_0$  strength for various solutions. Markers at a given  $B_0$  are offset to make error bars clearly visible. Error bars represent the minimum and maximum expected  $r_1$  assuming dominant errors are from volume measurements in solution preparation.

**References:** 1. Padhani A, et al, *Clin Radiol* **56**:607 (2001). 2. Spinosa DJ, et al, *Radiology*, **223**:319 (2002). 3. Keston P, *Clin Radiol*, **57**:505 (2003). 4. Jeresch-Herold M, *JMRI*, **19**:758 (2004). 5. Donahue KM, et al, *MRM*, **32**:66 (1994). 6. Stanisz, GJ, et al, *MRM*, **44**:665 (2000). 7. Rohrer M, et al, *Invest Radiol*, **40**:715 (2005). 8. Caravan P, et al, *Contr Med Mol Imag*, **4**:89 (2009). 9. Banay-Schwartz M, *Age*, **15**:51 (1992). 10. Gowland PA, et al, *MRM*, **18**:224 (1991). **Acknowledgements:** NIH grant R01 CA109106-06A1