

Nuclear spin properties of hyperpolarized solid-state MRI agents.

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Introduction – Hyperpolarization of solid-state nanoparticles such as ²⁹Si nuclei in silicon nanoparticles (SiNP) [1,2] and ¹³C nuclei in diamond [3] are emerging medical imaging modalities for non-invasive targeted diagnostics using MRI. These nanoparticles are attractive due to their negligible *in-vivo* toxicity [4] and lack of *in-vivo* background signal, while their surface chemistry and variable porosity makes them ideal for targeted drug delivery [5]. The unique nuclear spin properties of these solid state systems leads to multi hour spin relaxation (T_1) times that are unaffected by changes in magnetic field or temperature [6], and depend only on the particle size, isotopic concentration and materials purity [1]. Here we investigate the ²⁹Si nuclear spin properties of bare and functionalized SiNPs suitable for targeted gastrointestinal imaging agents.

Methods – SiNPs (approximately 2 μ m in diameter) were sourced commercially from Alfa Aesar. A two-step surface functionalization process with aminopropyltriethoxysilane (APTES) and polyethylene glycol (PEG) was carried out to enhance biocompatibility and hydrostability. Hyperpolarization of the ²⁹Si nuclear spins was undertaken at low temperatures using a home-built dynamic nuclear polarization (DNP) polarizer operating at 81 GHz and 3 K located next to a 4.7 T animal imager. *In-vivo* experiments were carried out on normal murine animal models with hyperpolarized SiNPs (~50 mg) diluted in saline (0.5 ml) and delivered into the gastrointestinal tract via a catheter. ²⁹Si spectroscopy was undertaken with a custom built surface coil using a variable flip angle sequence. Signal quantification was obtained via a co-located trimethoxysilane sample (1 ml) positioned above the coil. ²⁹Si imaging was performed using a fast spin-echo sequence with parameters (1 slice, slice thickness = 60 mm, $\alpha = 90^\circ$, TR = 1.5 ms, TE = 0.74 ms, FOV = 40 x 40 mm, 64 x 64 pixel resolution), with a total imaging time of 300 ms.

Results -

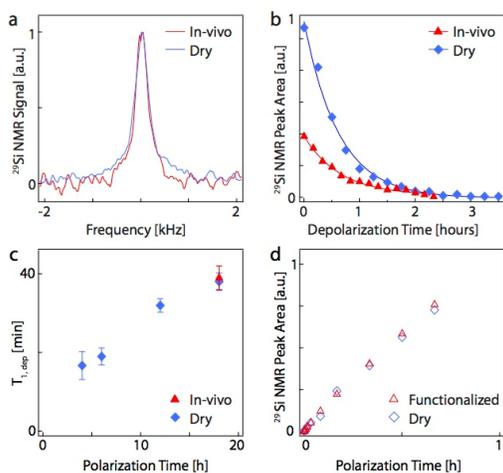


Figure 1 – (a) ²⁹Si NMR spectrum of HP SiNPs recorded *in-vivo* (red) and dry (blue) 30 min after delivery to the imaging magnet (b) Decay of ²⁹Si nuclear polarization of hyperpolarized SiNPs *in-vivo* (red) and dry (blue) after 18 h of DNP. Both show depolarization times of approximately 40 min. The difference in signal is attributed to sample loss in the dilution and injection process. (c) Characteristic ²⁹Si depolarization time constants in dry particle phantoms recorded in the imager for polarization times of 4, 6, 12, and 18 hours. (d) Time evolution of the ²⁹Si nuclear polarization under DNP conditions for particles with and without surface functionalization with PEG.

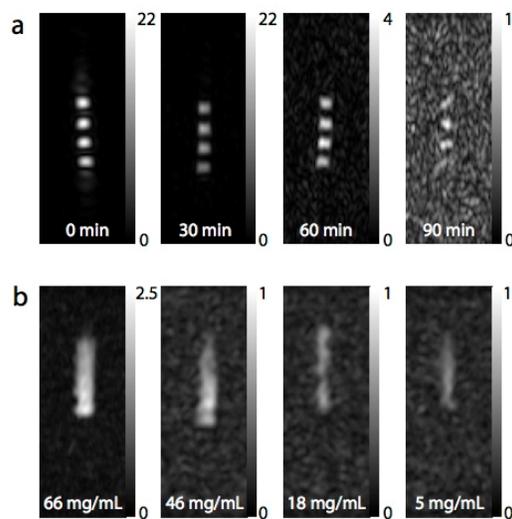


Figure 2 – (a) ²⁹Si MRI of a pixelated phantom containing silicon particles (15 mg in each pixel) with a delay of 0, 30, 60 and 90 min from when the phantom was loaded into the magnet until imaging. (b) ²⁹Si MRI of a cylindrical concentration phantom with particle concentrations of 66 mg/mL, 46 mg/mL, 18 mg/mL and 5 mg/mL. Images have been cropped from 40 mm to 15 mm in the horizontal dimension after processing.

Discussion - DNP in silicon particles is driven by electronic defects that exist at the particle surface and the nuclear hyperpolarization is carried into the center of the particles by nuclear spin polarization. Increasing the polarization time results in an increase in ²⁹Si signal together with longer depolarization times. After long periods of hyperpolarization, the polarized ²⁹Si nuclei in the core of the particle are protected from relaxation sources located at the particle surface such as those due to surface functionalization or the *in-vivo* environment. Single shot ²⁹Si images can be obtained up to 90 min after delivery to the imaging magnet, and down to concentrations of 5mg/mL, corresponding to sub-picomolar SiNP concentrations.

References – [1] Aptekar et al. ACSNano (2009) [2] Cassidy et al (2012) [3] Rej et al (2011) [4] Park et al Nature Materials (2009) [5] Tanaka et al, Cancer Research (2010) [6] Lee et al. PRB (2010).

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