

## Comparison of Arterial Spin Labelling and R2\* as Predictive Response Biomarkers for Vascular Targeting Agents in Liver Metastases

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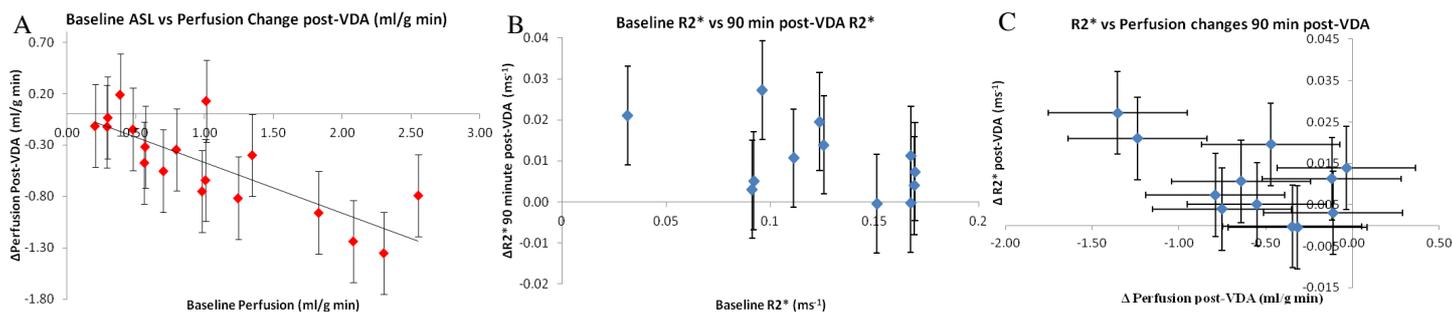
**Introduction:** Metastatic liver disease is the main cause of mortality in colorectal carcinoma (CRC) patients, with a 5 year survival rate of 40% following surgical resection of metastases<sup>1</sup>. Surgery with curative intent is only possible in 10-20%<sup>1</sup> of patients, demonstrating the need for alternative therapeutic approaches. The vascular disrupting agent OXi4503 is a compound that targets tumour vasculature and causes central tumour necrosis<sup>2</sup> leaving a small viable rim of tumour cells<sup>3</sup>. The acute (within 4 hours) accumulation of paramagnetic deoxyhaemoglobin resulting from vascular disruption has allowed R<sub>2</sub>\* changes to be used as a biomarker of therapeutic effect<sup>4</sup>. However, Arterial Spin Labelling (ASL) could offer an alternative quantifiable technique for assessing response, by measuring acute changes in tumour perfusion using wholly endogenous contrast mechanisms<sup>5</sup>. The current study therefore aims to compare changes in R<sub>2</sub>\* and ASL following OXi4503 treatment in a preclinical liver metastasis model.

**Method: Animal model:** The CRC cell line SW1222 was injected intrasplenically at a concentration of 1x10<sup>6</sup> cells in 100 µl in serum free media into n=6 MF1 nu/nu mice. Cells were allowed to wash through to the liver for 1 minute followed by splenectomy. Solid tumour deposits developed within the liver at ≈4 weeks following surgery.

**MRI:** A 9.4T Agilent VNMRS 20cm horizontal bore system with a 39mm birdcage coil was used, with a warm air blower to maintain animal temperature. Respiratory gating (SA instruments, New York, USA) was used on all scans. Fast spin echo images were used to define a suitable imaging slice within the liver followed by a segmented FAIR Look-Locker ASL sequence with a single slice spoiled gradient readout<sup>5</sup>. R<sub>2</sub>\* values were assessed by a multi-gradient echo (MGE) image sequence covering the entire liver. *FAIR Look-Locker ASL sequence parameters:* 30 x 30mm FOV, 128x128 matrix, TE: 1.18 ms, TI: 110 ms, TR<sub>RF</sub>: 2.3 ms, TR<sub>i</sub>: 13 s, 50 inversion recovery readouts. Localised inversion thickness: 6 mm, imaging readout slice thickness: 1 mm, 4 lines per segmented acquisition. *MGE sequence parameters:* 8 echoes, TE<sub>1</sub>=2ms, echo spacing=2ms, TR=280ms; 128x128 matrix, 40x40mm FOV, 1mm slice thickness.

**Dosing:** Cannulation of the tail vein was performed prior to baseline scans. Dosing of 40mg/kg via this remote i.v. line was performed in the scanner bore after baseline scans, and data acquired at 90min post dose.

**Data analysis:** n=18 metastases were evaluable across the n=6 mice for ASL and n=12 were available for R<sub>2</sub>\* analysis. Perfusion maps were generated using the Belle model<sup>7</sup>, (T<sub>1 blood</sub>=1.9 s<sup>8</sup>, blood-tissue partition coefficient λ=0.95 ml/g<sup>9</sup>) in MATLAB and R<sub>2</sub>\* maps were created using IDL.



**Fig 1:** Plots showing the liver metastases response 90 minutes post OXi4503 against baseline measurements for perfusion (A) and R<sub>2</sub>\* (B). A significant trend can be seen in the perfusion changes compared to the initial perfusion, however no trend can be seen in the post dose vs baseline R<sub>2</sub>\*. No significant correlation can be seen between the change in R<sub>2</sub>\* and perfusion post dose (C).

**Results:** A significant decrease was measured in ASL measurements of tumour perfusion at 90 mins following OXi4503 administration ( $P < 0.01$ , Mann-Whitney U test), with a mean change of -0.49 ml/g/min (-43%). A significant correlation was observed between baseline perfusion and the change in perfusion following therapy (Fig. 1A), suggesting that tumours better perfused at baseline responded better to the therapy. A significant increase in R<sub>2</sub>\* was also measured ( $P < 0.01$ , Mann-Whitney U test), with a mean change of 0.010 ms<sup>-1</sup> (13%), but with no significant correlation with initial R<sub>2</sub>\* (Fig.1B). There was no significant correlation between ASL and the R<sub>2</sub>\* responses (Fig.1C).

**Discussion:** We were able to detect acute changes in tumour pathophysiology caused by OXi4503 with both ASL and R<sub>2</sub>\*, with a significant decrease in mean perfusion and increase in R<sub>2</sub>\*. This is consistent with the mechanism of action of VDAs: cessation of blood flow leads to a reduction in tumour perfusion and an increase in paramagnetic deoxygenated haemoglobin. Changes in R<sub>2</sub>\* and perfusion were not correlated, indicating a complex relationship between changes in flow and accumulation of deoxyhaemoglobin, which may be specific to individual tumours.

The data presented here shows that ASL can be a predictor of vascular targeting agent efficacy in liver metastases, suggesting that tumour deposits better perfused at baseline display a greater acute response. R<sub>2</sub>\* response was not suggestive of any prognostic ability, but did respond positively. Given the mechanism of action of vascular disrupting agents, ASL provides response biomarkers that afford a less ambiguous interpretation than intrinsic susceptibility (R<sub>2</sub>\*) measures. However, an approach combining the two may provide deeper insights in to the mechanics of tumour response *in vivo*, by relating flow changes to changes in blood oxygen saturation.

The detection of a variable response, even in tumour deposits within the same liver highlights the need for robust assessment of response within individual patients. ASL sequences are non-invasive and do not require the administration of a contrast agent and so could be performed serially, soon after therapy to inform on drug efficacy. Given that brain and kidney FAIR ASL is commonplace in clinical scanners we anticipate a translation of hepatic ASL should be straightforward. Further work will characterise the response at later time points post OXi4503 and assess changes in perfusion and R<sub>2</sub>\* in other tumour lines in preclinical metastases models.

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**References:** (1) Penna C and Nordlinger B, *British Medical Bulletin* (2002) 64:127-140, (2) Chan LS et al, *Anticancer Drugs* (2008) 19(1):17-22, (3) Pedley RB et al, *Cancer Research* (2001) 61(12):4716-22, (4) Zweifel M et al, *Proc. Intl. Soc. Mag. Res. Med.* 19 (2011) 339, (5) Ramasawmy R et al, *Proc Brit Chap ISMRM* (2011) poster 62, (6) Chan LS et al *Anticancer Res* (2007) 27:2317-2324. (7) Belle, et al. *J Magn Reson Imaging* 1998;8:1249-1245. (8) Campbell A, et al. *Magn Reson Med.* 2012; doi: 10.1002/mrm.24243. (9) Rice G et al, *J Pharmacol Methods.* 1989 Jul;21(4):287-97.