

Global intravascular and local hyperoxia contrast phase-based blood oxygenation measurements

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Target: The measurement of venous blood oxygenation (Y_v) is important in cases where oxygen extraction fraction (OEF) may be perturbed, e.g. multiple sclerosis, traumatic brain injury and carotid stenosis; and also for hyperoxia-based BOLD calibration^{1,2}. **Purpose:** Phase-based measurements of Y_v provide a simpler data acquisition to R_2^* - and R_2 -based methods^{3,4,5}. Previous phase-based approaches measure the intravascular (IV) phase of a large draining vein^{6,7}, recently we have described a technique to compare the change in phase distribution local to a vein between normoxia (NO) and hyperoxia (HO)² that does not require knowledge of the vein's geometry. This approach assumes the phase distribution on NO is dominated by the effect of IV deoxyhaemoglobin and thus provides a template for the field change on HO (applying Eq.1 over a limited range of r) due to a change in Y_v .

Aim: To assess three approaches for measuring Y_v , using: the IV phase in the superior sagittal sinus and an infinite cylinder model (*Method A*) or a forward field calculation (*Method B*) of the IV field shift; or the phase distribution change on hyperoxia to measure localised values of Y_v (*Method C*).

Methods: 8 healthy volunteers (7M/1F, 22-32y.o.) were scanned, using a Philips Achieva 7T system with volume transmit/32-channel receive head coil. For 6 subjects, 3D whole-head FLASH data (0.65mm isotropic voxels, TR/TE=9.5/5ms; SENSE=2.25(AP); flow compensated; T_{A0} =3.5 min) were acquired at NO and at HO, controlling end-tidal O_2 ($P_{ET}O_2$) and CO_2 ($P_{ET}CO_2$) partial pressures using a Respiract™ (Thornhill Research Inc., Toronto, Canada). $P_{ET}O_2$ was targeted between the subject's baseline (~110 mmHg; NO) and 500 mmHg (HO), whilst $P_{ET}CO_2$ was held constant. To assess the reproducibility of Methods A&B, two further subjects were scanned 6 times at 5.5 min intervals while breathing room air. For all subjects, a multi-echo FLASH scan was acquired at NO to identify veins (3 echoes, TR/TE₁/ΔTE=21/5/5ms; SENSE=2.25/1.4(AP/FH)). Phase images were unwrapped, magnitude images were co-registered using FLIRT (FSL) and a homodyne filter (FWHM 8mm) was applied to phase images. **Method A & B:** In 5 slices in which the sagittal sinus was parallel to B_0 , the phase difference ($\Delta\phi_{IV}$) between IV and extravascular (EV) ROIs (*Fig1*) was found. Eq.2 was used to calculate Y_v from $\Delta\phi_{IV}$, assuming a susceptibility difference between tissue and deoxygenated haemoglobin of $\Delta\chi_{do} = 0.247$ ppm(cgs)⁸, and $\beta=1/3$ (*Method A*) or β calculated using a forward field calculation⁹ for the sagittal sinus (*Method B*). **Method C:** A vein mask was formed by thresholding the R_2^* map created from the multi-echo data ($R_2^* > 100s^{-1}$). For each vein, this mask was dilated by 1 voxel to include the EV field perturbations (*Fig1*). HO induced increases in Y_v (ΔY_h) reduce the amplitude of the EV field perturbation (*Fig2*). The HO phase distribution ($\Delta\phi_{HO}(r)$) was fitted to the NO phase distribution ($\Delta\phi_{NO}(r)$) for α (*Eq1*) over all voxels in the dilated mask of each vein. The value of α was then used in Eq. 3 (derived in²) to calculate Y_v for each vein, where $\Delta\chi_{oxy} = -0.017$ ppm(cgs)⁸ is the susceptibility difference between tissue and oxygenated haemoglobin. ΔY_h was then estimated from $P_{ET}O_2$, assuming $P_{ET}O_2 \approx$ arterial PO_2 (validity discussed in²). Monte Carlo simulations were performed, to assess how noise propagates through Eqs.1&3.

Results: *Method A:* $Y_v = 0.70 \pm 0.01$ (mean \pm SEM, 8 subjects). *Method B:* $Y_v = 0.60 \pm 0.02$, since $\beta = 0.257 \pm 0.005$ ($< 1/3$ in all subjects), with reproducibility $< 1.5\%$ for both subjects. *Method C:* $P_{ET}O_2$ increased by 320 ± 20 mmHg (mean \pm SEM, 6 subjects) giving¹ $\Delta Y_h = 0.066 \pm 0.003$ and $Y_v = 0.60 \pm 0.01$ averaged across veins. *Fig3* shows a map of Y_v for veins identified on the R_2^* map; visual inspection of these maps showed no apparent bias due to vessel orientation. *Fig4* shows the distribution of Y_v measured across veins of all subjects.

Discussion: Methods A&B use large vessels and so provide high SNR, but require knowledge of the vein's size, shape and orientation, restricting the application to larger vessels to avoid partial voluming effects. Furthermore, Method A is limited to long, straight vessels. The HO phase contrast method (Method C) does not require knowledge of the vein's size, shape or orientation and can be used on smaller veins, but does require an estimate of ΔY_h . The distribution in *Fig4* is centred on $Y_v \approx 0.6$, but skewed to lower Y_v . Monte Carlo simulations produced a much narrower (but also skewed) distribution of Y_v (*Fig4*-white histogram), suggesting that real Y_v heterogeneity was detected *in vivo*, although there was no obvious change in Y_v between brain regions. An 8 mm spatial filter width was used to preserve the phase contrast in the sagittal sinus, whilst removing large scale differences between NO and HO phase images. However, the systematically higher Y_v values produced by Method A suggest that the filter attenuated the sagittal sinus phase, whilst Method B is less sensitive to this effect. Decreased tissue-vein $\Delta\chi$ due to increased tissue PO_2 on HO¹⁰, will have a negligible effect here (tissue PO_2 increase of 50 mmHg, giving $\Delta\chi \approx 10^{-4}$ ppm).

Conclusion: Y_v measured from phase in the superior sagittal sinus gave repeatable results, but this approach is limited in utility since it can only be applied to large veins. Method C measures Y_v from the reduced field perturbation around a vein on HO, and is not limited to large veins, allowing the variation in Y_v across the brain to be examined which will be important clinically.

References: 1. Chiarelli et al. NeuroImage 37:808-820 (2007); 2. Driver et al. NeuroImage 63:1178-1187 (2012); 3. He and Yablonskiy MRM 57:115-126 (2007); 4. Lu and Ge MRM 60:357-363 (2008); 5. Bolar et al. MRM 66:1550-1562 (2011); 6. Haacke et al. HBM 5:341-346 (1997); 7. Jain et al. JCBFM 30:1598-1607 (2010); 8. Spees et al. MRM 45:533-542 (2001); 9. Marques, J.P., Bowtell, R. Concepts Magn Reson 25B, 65-78 (2005); 10. Schwarzbauer and Deichmann NeuroImage 59:2401-2412 (2012). **Acknowledgement:** Funded by the UK MRC.

$$\Delta\phi_{HO}(r) = \alpha \cdot \Delta\phi_{NO}(r) \quad - \text{Eq1}$$

$$(1 - Y_v) = \frac{\Delta\phi_{IV}}{\gamma \cdot TE \cdot B_0 \cdot \Delta\chi_{do} \cdot Hct \cdot \beta} \quad - \text{Eq2}$$

$$Y_v = \frac{\left[\frac{-\Delta Y_h (\Delta\chi_{oxy} - \Delta\chi_{do})}{(1-\alpha)} - \Delta\chi_{do} \right]}{(\Delta\chi_{oxy} - \Delta\chi_{do})} \quad - \text{Eq3}$$

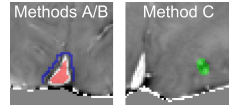


Figure 1: ROIs used.

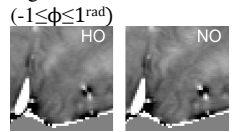


Figure 2: Effect of HO. (-0.5 ϕ \leq 0.5 rad)

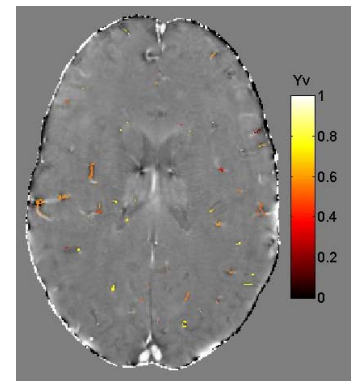


Figure 3: Variation in Y_v between veins for a single subject, overlaid on the NO phase image. (-1 ϕ \leq 1 rad)

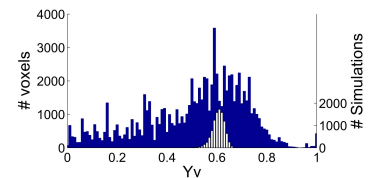


Figure 4: Histogram of venous Y_v values (weighted by # voxels in each vein) including all 6 subjects. Monte Carlo simulation results (white) indicate the expected noise distribution.