A novel technique for correlating between in vivo and ex vivo whole body mammary glands: Comparison of apparent coefficient values

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Target audience: This study will benefit the people who are interested in the mouse model of breast cancer, and correlation between *in-vivo* and *ex-vivo* mammary glands.

Introduction: Transgenic female mouse models are often used in pre-clinical research to understand progression of breast cancer. Correlations between *in vivo* and *ex vivo* MRI, as well as with histology, are often performed to confirm the type and stage of cancer ^[1-3]. Previous studies in transgenic mouse models demonstrated that not all ductal carcinoma *in situ* (DCIS) lesions may progress into invasive cancers, and tumorigenesis is accelerated in upper glands, i.e., at cervical and thoracic glands for SV30Tag mice ^[2]. Diffusion weighted imaging (DWI) has been increasingly used to detect breast cancer. However, correlation of whole body mouse mammary glands between *in vivo* and *ex vivo* DWI MRI has not been done and remains a challenge. Here, we report a novel approach to imaging whole body mouse mammary gland to correlate *in vivo* and *ex vivo* MRI and evaluate the ADC values. The polyoma middle T oncoprotein (PyMT) mice, which have been proved to be an excellent model to understand the biology of tumor progression in humans, are used in this study.

Methods: Four adult PyMT mice were imaged at 9.4T; whole-body scanning was used to study all of the mammary glands. For in vivo experiments, multi-slice RARE (Rapid Acquisition with Relaxation Enhancement) spin echo T2-weighted images with fat suppression were acquired to identify abnormal regions (TR/TE_{effect} = 4000/26 ms, FOV = 30 mm, matrix size = 256^2 , slice thickness = 0.5 mm, NEX = 2, RARE factor = 4) for upper and lower mammary glands separately. Then DWI was performed (TR/TE = 4000/26 ms, b-value = 0, 1000, and 3000 s/mm², FOV = 30 mm, matrix size = 128^2 , slice thickness = 1.0 mm, NEX = 1) for nine slices selected based on the T2W images. To facilitate correlation of in vivo and ex vivo images, two standards (1 mm diameter tube filled with water) were placed at 5 mm from the 1st nipple from top and the last nipple from the bottom for both *in vivo* and *ex vivo* scans. For *ex vivo* experiments, the skin and glands were taken by carefully excising the skin from the mouse. A midline incision along the ventral abdomen was made from the tail to the head; and then the skin, glands, and tumors were gently peeled from the body muscle so that the hide remained intact. Once formalin fixed, the tissue was placed around a fomblin filled tube and sutured together back along the midline to resemble the round shape of a mouse. This skin and tube were then placed in a larger tube, filled with fomblin, and sealed before being placed into the 35 mm diameter quad coil. The same pulse sequences used for *in vivo* experiments were also used for ex vivo experiments.

Results: The figure shows fat suppression T2-weighted images (left panel) and corresponding ADC maps (right panel) for both *in vivo* and *ex vivo* set-ups at the cervical and abdominal glands. The abnormal mammary gland regions are circled in red on the T2-weighted images. As we can see, most abnormal regions had ADC values between ~0.25 to ~ 0.75×10^{-3} mm²/s in both *in vivo* and *ex vivo* experiments, which are close to reported values from other clinical studies. There are also small regions with ADC values > 1.0×10^{-3} mm²/s. Due to a lack of landmarks and the potential for the skin to be slightly stretched causing deformation in the *ex vivo* images, it is hard to visually directly compare *in vivo* and *ex vivo* images. Nevertheless, small, distinct features, such as lymph nodes and small tumors, correlated well in full size images between *in vivo* and *ex vivo* images.



Discussion: To our knowledge, this is the first report of correlation between *in vivo* and *ex vivo* MRI of whole body mouse mammary glands. With this technique, features found on *in vivo* slices can be found on *ex vivo* slices and then accurately identified in fixed tissue, and correlated with histopathology. The ADC values calculated from both *in vivo* and *ex vivo* DWI were reasonable. The motion-free *ex vivo* ADC maps may assist us in finding values that characterize DCIS, invasive tumor and lymph node *in vivo*. More quantitative analysis and correlation with histopathology are under way to evaluate the ADC for detection and diagnosis mammary cancer, and to understand cancer progression.

Acknowledgement: This research is supported by NIH 1R01CA133490-01A2 and The University of Chicago Cancer Center. [1] Jansen et al. Phys Med Biol. 2008; 53:5481-93. [2] Jansen et al. Breast Cancer Res. 2009; 11:R65. [3] Jansen et al. NMR Biomed. 2011; 24:880-7.