

# Lab-on-chip for NMR microscopy: description and application for neurospheres imaging and spectroscopy studies

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## Introduction

Extensive and continuous monitoring is one of the main challenges in the study of cell cultures and organotypic tissues. We have developed a lab-on-chip device allowing monitoring different parameters, as temperature, pH and pO<sub>2</sub> or pCO<sub>2</sub> by using optical fibres (Patent: PCT\_ES2011\_070173\_ISRWO). Additionally, optical, fluorescence and confocal microscopy can be used together with NMR microscopy and spectroscopy. The aim of this communication is to describe the lab-on-chip and demonstrate the possibility of obtaining high resolution images and spectra at very low volumes by using NMR microscopy in a single neurosphere.

## Subjects and Methods

A set of neurospheres, from sub-ventricular zone (SVZ) of postnatal rats (P0-P5) and at passage 3, were introduced in the lab-on-chip system (Figure 1). The neurospheres were allowed to grow and differentiated by a continuous flow of DMEM growth medium (180 nl/minute) (Figure 2) the temperature was monitored by a fibre optic (LUXTRON 16521812). A surface coil of 500 µm of diameter (Figure 1, lower right part), allowing the insertion of the lab-on-chip device, and a set of gradients of 60 Amp were used for obtaining images and spectra in a Bruker 600 MHz instrument.

Gradient Echo sequences were used for obtaining NMR microscopy images: i) two with TE and TR 15 and 172 ms, respectively, and 64x64 (43s) and 128x128 (7m 16s) matrix with a resolution of 16 x 16 x 150 µm; and ii) one with a matrix of 256 x 256 (2h 43 s) with 8 x 8 x 50 µm resolution and TE = 7 ms and TR = 100 ms. PRESS with VAPOR water suppression was used for SV spectra acquisition, TE and TR were 12 and 1500 ms, respectively, and 0,2 x 0,2 x 0,2 mm dimensions with a nominal volume of 8 nl were used. (Figure 6)

## Results and Discussion

Highly resolved images were obtained for the single neurosphere developed in the lab-on-chip system by using a 500 µm surface coil (Figure 3). These images allowed to clearly differentiate heterogeneous parts, the surface and necrotic centre, of a ca 350 µm neurosphere located in the field of view of the coil. Although with limited S/N, different resonances were initially assigned in the growth medium and in the neurosphere (Figure 4). In particular, high levels of lipids were observed in the necrotic centre of the neurosphere.

## Conclusions

Different regions and metabolic information from a single neurosphere can be obtained by the combined use of a particular lab-on-chip developed for 500 µm surface coils and NMR microscopy and spectroscopy.

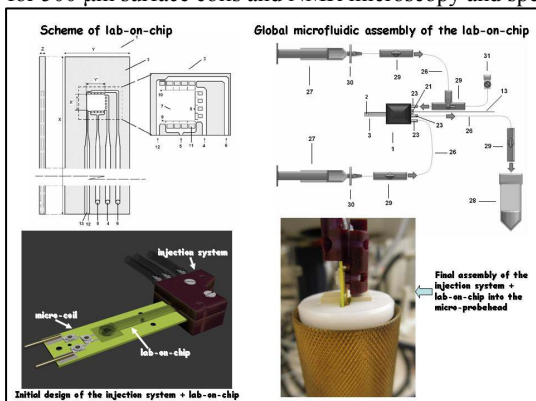


Figure 1.- Global description of the complete system lab-on-chip + injection device + microcoil + probehead

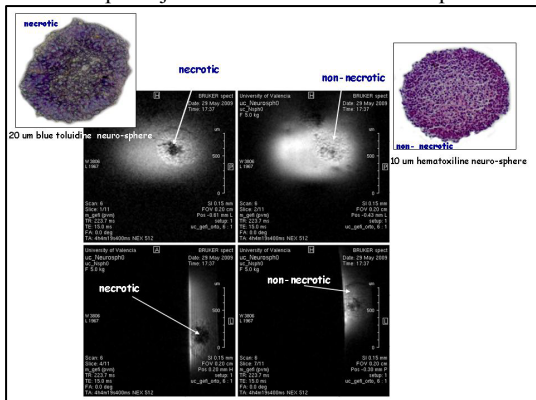


Figure 3.- Example of two slices (150 µm) axial (upper part) and sagittal (lower part) of a neurosphere of ca.350 µm growth inside of the lab-on-chip and obtained by using a 500 µm microcoil at 14 T.

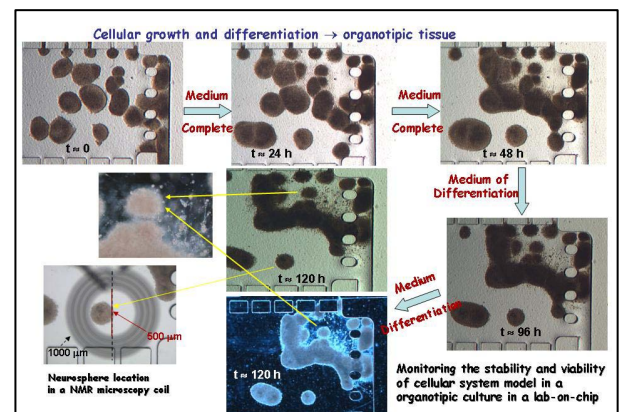


Figure 2.- Example of cellular growth and differentiation inside of the lab-on-chip under temperature control by optical fiber.

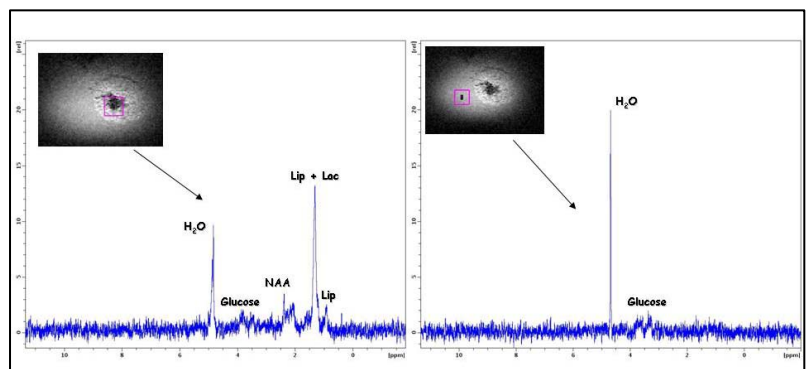


Figure 4.- Examples of spectra of two square SVs of 8 nl dimensions obtained by PRESS at 14T and using a 500 µm microcoil. Initial metabolite assignments are shown