Human Cardiac ³¹P Metabolite Concentrations at 3T

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INTRODUCTION. Phosphocreatine (PCr), is the main energy reservoir in both skeletal and cardiac muscles, supplying ATP energy for contraction via the creatine kinase (CK) reaction. Cardiac ³¹P MRS measurements of absolute ATP and PCr concentrations could

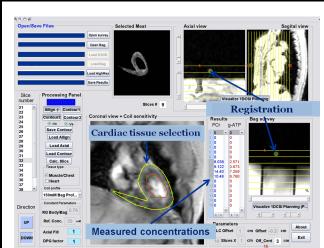


Fig. 1 Screen shot of the concentration quantification tool illustrating the different steps used.

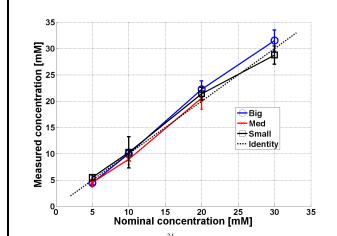


Fig. 2 Mean ±SD of measured ³¹P concentrations as a function of nominal concentration values in three cylindrical phantoms of diameters 3.7cm (small), 5.7cm (medium) and 9.4cm (big).

benefit from higher magnetic field strengths, but is challenged by T₂-decay, increased RF power deposition, and field inhomogeneities. While these have been specifically addressed at 3T [1,2], approaches to concentration measurements employing water-referencing and ¹H signal detection with a single tuned ³¹P coil [3] are compromised at 3T due to differences in spatial sensitivity at the higher field strengths.

This work presents a new method and semi-automated tool for measuring PCr and ATP concentrations from 1D chemical shift imaging (CSI) ³¹P spectra that compensates for the nonuniform sensitivity. It is validated in phantoms and applied to the human heart. **METHODS.** The method utilizes a single 1D CSI acquisition with a 5ms duration, 7kHz-sweep, frequency-cycled adiabatic half passage pulse [2]. Bloch equation analysis shows that this yields >95% of the magnetization [1]. Using a 17cm/8cm transmit/receive ³¹P coil set in a Philips 3T Achieva scanner, this permits cardiac measurements at depths of up to 8-9cm [1,2]. A 2cm diameter sphere filled with NaH₂PO₄ and a vitamin pill were both fixed to the coil housing to compensate for loading differences and to register the subject with a concentration reference, respectively. A ~6cm diameter cylindrical phantom containing 30mM NaH₂PO₄ was used for the concentration reference. A single 3DCSI scan of a 32x32cm² 600mM NaH₂PO₄ phantom was used to map the coil's sensitivity profile.

Multi-slice high-resolution ¹H MRI was used to identify the target tissue. Metabolite concentrations were calculated from:

$$[Metabolite]_{muscle}^{n} = \frac{S_{muscle}^{n} \cdot (1 - \exp(-TR_{ref} / T1_{ref})) \cdot \sum_{i} \sum_{j} w_{ij}^{ref} C_{ij}^{n}}{S_{ref}^{n} \cdot (1 - \exp(-TR_{muscle} / T1_{muscle})) \cdot \sum_{i} \sum_{j} w_{ij}^{muscle} C_{ij}^{n}} \times F_{g} \times F_{m} \times [^{31}P]_{ref}^{n}$$

where S is the fitted spectral area, n is the slice number, W is the selected area from images, C is the coil sensitivity, F_g is the measured loading factor, F_m is a measured motion correction factor for the heart, determined from free breathing axial cine images, and *ref* denotes the reference phantom. Peak areas were quantified using the CFIT routine [4]. Fig. 1 shows the *Matlab*-based software interface to perform all of the described operations for obtaining concentration values.

EXPERIMENTS AND RESULTS. The method was validated in 11 cylindrical phantoms of different diameters (3.7, 5.7 and 9.4 cm) and concentration values (5, 10, 20 and 30 mM). The mean measured ³¹P

concentration values agreed with the known phantom concentrations with a root-mean-square error of ~8% on average (Fig. 2).

Institutional Review Board-approved human cardiac studies were performed on 10 consenting healthy volunteers. Cardiac-triggered 16-step 1DCSI ^{31}P data were acquired with subjects prone on the coil set (field of view=16cm; repetition time \approx 16s; 2 averages; scan-time <10min). We measured an average cardiac [PCr] of 9.3 ± 1.7 and $[\gamma-ATP]$ of 5.2 ± 1.0 µmol/g wet wt, in agreement with prior measures at values at 1.5T ([PCr]/[ATP]= $10\pm2/5.8\pm1.6$ µmol/g [3], and $9\pm1.2/5.3\pm1.2$ µmol/g [5]).

CONCLUSION. A new semi-automated, image-guided method for measuring ³¹P concentrations in the human heart was developed that overcomes a number of challenges at 3T. The approach was validated in phantom studies, and provides PCr and ATP concentrations in healthy human hearts that closely agree with prior measures at 1.5T. The measurements can be performed in ~9min within a patient exam and thus can be part of a practical, quantitative cardiac ³¹P clinical research protocol.

ACKNOWLEDGMENTS. We thank Ronald Ouwerkerk for fruitful discussions and Tricia Steinberg for assistance with human studies. REFERENCES. 1. El-Sharkawy et al. *Magn Reson Med* 2009;61(4):785-795. 2. Schär M et al. Magn Reson Med 2010;63 (6):1493-1501. 3. Bottomley PA et al. *Magn Reson Med* 1996;35(5):664-670. 4. Gabr RG et al. *J Magn Reson* 2006;179(1):152-163. 5. Meininger M et al. *Magn Reson Med* 1999;41(4):657-663. Supported by NIH R01 HL56882, R01 HL61912 and a grant from the D.W. Reynolds Foundation.