

# Measurement of apparent diffusion coefficients (ADC) and $^1\text{H}$ transverse relaxation times ( $T_2$ ) of human brain metabolites and water: insights on white matter microstructure

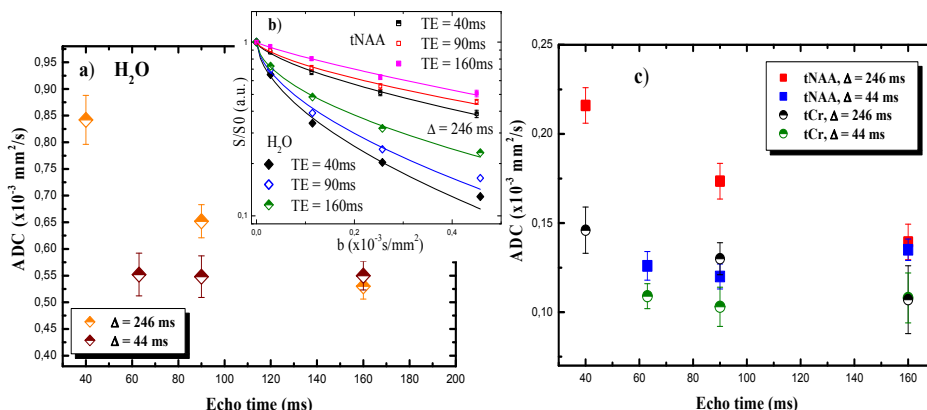
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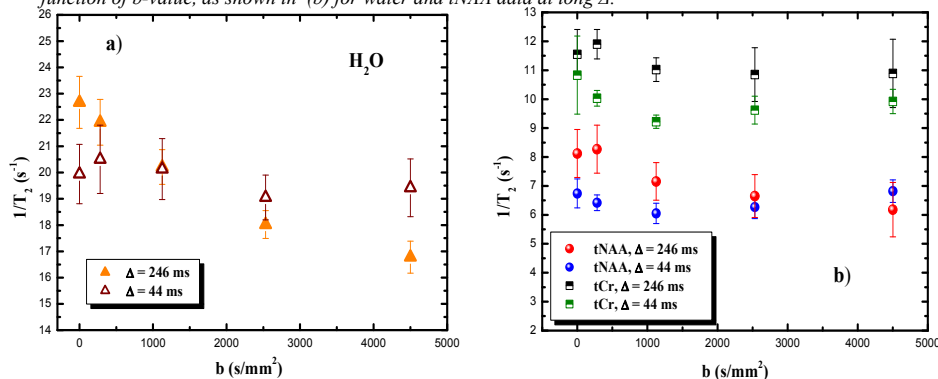
**Introduction:** The dependence of MR-measured apparent diffusion coefficients (ADCs) of molecules in biological tissues on a controllable, acquisition-specific timescale allows measurement of the characteristic length scales of tissue compartments and it has been recognized as a powerful mechanism for deriving information on tissue microstructure [1]. Metabolites are generally considered to be confined mainly in intracellular compartments and to exchange at a slower rate than water molecules between microscopic pools, making them better probes for compartmentalization in neural tissues [2,3] and therefore providing higher specificity in studying brain tissue microstructure and pathology. In addition, because of the differing molecular environments modulating dipolar interactions among proton spins, each micro-anatomical pool may also be characterized by a different  $^1\text{H}$  transverse relaxation rate ( $1/T_2$ ) [4]. The interaction between relaxation and diffusion characteristics has been thoroughly studied in porous media [5] and in excised nerves [4,6]. In this study, for the first time the interaction between relaxation and diffusion was studied for both water and metabolites in the human brain in vivo at 7T.

**Methods:** Scans were performed on a 7T scanner (Philips, Best, the Netherlands) equipped with a 32-channel receive coil and a quadrature transmit head coil (In Vivo, Florida). A single voxel, diffusion-weighted STEAM sequence was used to measure the diffusion of water and of the metabolites tNAA, tCr and tCho (not all data shown here). Two data-sets were obtained, from 7 young healthy volunteer each ( $29 \pm 8$  years), using two different diffusion times ( $\Delta$ ). Sequence parameters for the two protocols were:  $\Delta = 246/44\text{ms}$ ,  $\text{TM} = 230/14\text{ms}$ ,  $\delta = 10/24\text{ms}$ ,  $\text{TR} = 3$  cardiac cycles (triggering using PPU), spectral width 3kHz, 1024 sample points. Diffusion gradients were applied in 3 standard quasi-orthogonal directions with 5 increasing gradient strengths chosen in the range 0-3.6G/cm in order to obtain identical b-values of 0, 285, 1140, 2570, and 4575  $\text{s}/\text{mm}^2$  in both data-sets. A bipolar gradient scheme was employed to minimize eddy currents. Spectra were acquired for three echo times  $\text{TE} = 40/63, 90, 160\text{ms}$  with an increasing number of averages in order to keep similar SNR for each scan. A VOI of  $30 \times 20 \times 19 \text{mm}^3$  was positioned in parietal WM. The residual water peak was used to perform phase and frequency corrections on individual spectra before summation. Non-water suppressed spectra were also acquired to derive water diffusion and relaxation properties as well as for eddy current corrections.

**Results and discussion:** The water and metabolite normalized signals as a function of b-value show a non-monoexponential trend (Fig.1b), reflecting, among other factors, the presence of physical restriction hindering the diffusion of molecules. In order to extract the ADC values, the



**Fig.1:** Water (a), tNAA and tCr (c) ADCs measured as a function of echo time for two different diffusion times  $\Delta$ . ADCs have been estimated from fits to stretched exponentials of normalized signal decay plotted as a function of b-value, as shown in (b) for water and tNAA data at long  $\Delta$ .



**Fig.2:** Water (a), tNAA and tCr (b) proton transverse relaxation rates  $1/T_2$  measured as a function of b-value for two different diffusion times  $\Delta$ .

and “free” states affecting both metabolite relaxation rates and diffusion.

**References:**[1] K. Nicolay *et al.*, *NMR Biomed.*, **14**, 94–111 (2001). [2] Y. Assaf *et al.*, *NMR Biomed.*, **12**, 335–344 (1999). [3] Y. Cohen *et al.*, *NMR Biomed.*, **15**, 516–542 (2002). [4] S. Peled *et al.*, *MRM*, **42**, 911–918 (1999). [5] P.T. Callaghan *et al.*, *MRM*, **162**, 320–327 (2003). [6] M. D. Does *et al.*, *MRM*, **844**, 837–844 (2000). [7] K.M. Bennett *et al.*, *MRM*, **50**, 727–734 (2003).