

## Increasing diffusion time improves in vivo DWI sensitivity to liver fibrosis

Darwin S. Gao<sup>1,2</sup>, Jian Yang<sup>1,3</sup>, Matthew M. Cheung<sup>1,2</sup>, April M. Chow<sup>1,2</sup>, Shujuan Fan<sup>1,2</sup>, Kwan Man<sup>4</sup>, and Ed X. Wu<sup>1,2</sup>

<sup>1</sup>Laboratory of Biomedical Imaging and Signal Processing, The University of Hong Kong, Hong Kong SAR, China, People's Republic of, <sup>2</sup>Department of Electrical and Electronic Engineering, The University of Hong Kong, Hong Kong SAR, China, People's Republic of, <sup>3</sup>Department of Radiology, The First Affiliated Hospital of Medical School, Xi'an Jiaotong University, Xi'an, China, People's Republic of, <sup>4</sup>Department of Surgery, The University of Hong Kong, Hong Kong SAR, China, People's Republic of

**INTRODUCTION:** Hepatic fibrosis, which occurs in response to chronic liver injury from many causes such as hepatitis and alcohol intoxication [1], was historically regarded as irreversible process. In recent years, it is considered to be a wound-healing response that is much more reversible [2, 3]. Percutaneous liver biopsy has long remained as the gold standard for diagnosis and staging hepatic fibrosis. However, its utility is limited by the invasiveness of the procedure, likelihood of sampling errors and inter-observer variations [4, 5]. Therefore, noninvasive methods with sufficient sensitivity to identify liver fibrosis at early stage and monitor fibrosis progression and regression are of great clinical and therapeutic values. Diffusion time dependency of measured diffusion coefficient, being the evidence of restricted diffusion, has been widely observed in brain tissues [6, 7]. In liver fibrosis, the structural damage on cellular scale may contribute to a modification of the restricted diffusion behavior of water molecules in the intra- and extracellular space. In this study, we examined whether different diffusion times would yield different sensitivities in detecting the pathological alterations in tissue microstructure during liver fibrogenesis in an experimental rat model.

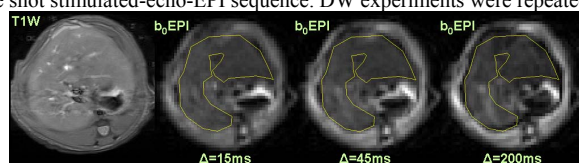
**METHODS: Animal preparation:** Sixteen male adult SD rats (220–260g, 6 weeks old) were assigned to two groups. The fibrotic group (N=12) received subcutaneous injection of 1:1 volume mixture of carbon tetrachloride (CCl<sub>4</sub>) in olive oil at a dose of 0.2 mL / 100 g of body weight twice a week for 8 weeks [8]. The control group (N=4) received no injection. DW MRI was performed in animals 1 day before, 2, 4, 6 and 8 weeks after CCl<sub>4</sub> administration. **MRI:** All MRI experiments were performed using a Bruker 7T scanner. DW images (DWIs) with 5 different b-values (0, 200, 400, 800 and 1000 s/mm<sup>2</sup>) along phase encoding direction (R-L) were acquired in one axial slice covering the liver with respiration-gated single shot stimulated-echo-EPI sequence. DW experiments were repeated with diffusion time  $\Delta = 15, 45$  and 200ms. Imaging parameters were TR/TE=2000/20ms,  $\delta = 3.1$ ms, slice thickness=3mm, FOV=51x51mm<sup>2</sup>, acquisition matrix=51x51, NEX=11.

**Data analysis:** A large ROI excluding large blood vessels was drawn on liver parenchyma encompassing a large homogeneous liver region. Apparent diffusion coefficient (ADC) was obtained by fitting the equation:  $S_b/S_0 = \exp(-b \times ADC)$  with the ROI measurements of  $S_b/S_0$  at all five b values (0, 200, 400, 800, 1000 s/mm<sup>2</sup>) using a least-square nonlinear fitting in Matlab. True diffusion coefficient ( $D_{True}$ ) was estimated by fitting the signal decay in the ROI on the images of b values higher than 200 s/mm<sup>2</sup> (400, 800 and 1000 s/mm<sup>2</sup>) to the equation:  $S_b/S_0 = (1-f) \times \exp(-b \times D_{True})$ . One-way ANOVA with Turkey's multiple comparison tests was employed to compare ADC, and  $D_{True}$  measurements between different time points of liver fibrosis, as well as different  $\Delta$ , and  $p < 0.05$  was considered as statistical significant.

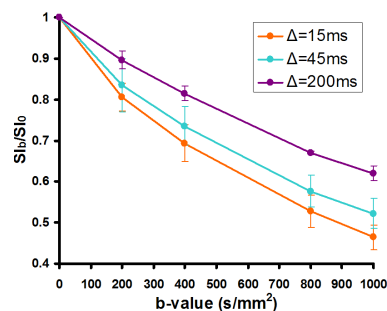
**RESULTS:** Fig.1 shows the representative T1W and B0 EPI images of  $\Delta = 15, 45$  and 200ms from one fibrotic animal. Typical ROI used for ADC and  $D_{True}$  measurement is also illustrated. Fig. 2 shows that the mean DW liver signal decay, computed as the average of  $S_b/S_0$  from 6 animals after 8-week CCl<sub>4</sub> insult, as a function of b-values at different times ( $\Delta$ s). Fig. 3 shows the liver ADC and  $D_{True}$  values measured with  $\Delta = 15, 45, 200$  ms at different time points of liver fibrosis. ADC,  $D_{True}$  generally decreased with  $\Delta$  in both normal and fibrotic liver, confirming their diffusion time dependency and spatially restricted diffusion in liver. Fig.4 compares the liver ADC and  $D_{True}$  values measured at five different stages of liver fibrosis for three different times. The comparisons demonstrate that  $D_{True}$  yielded higher sensitivity than ADC in detecting cellular changes during liver fibrosis. Specifically, greater statistical significances were generally observed when comparing  $D_{True}$  between 0-week, 2-week and 4-week CCl<sub>4</sub> insult at three different  $\Delta$  (Fig. 4 right). Moreover, the percentage changes of  $D_{True}$  from week 2 to week 4 after CCl<sub>4</sub> insult were greater at  $\Delta = 200$ ms (15±3.3%) than  $\Delta = 45$ ms (12±4.1%) and  $\Delta = 15$ ms (9±3.2%), indicating that long diffusion time improved  $D_{True}$  sensitivity. ADC and  $D_{True}$  of liver from age-matched control group exhibited no significant differences over different time points from liver fibrosis group at all  $\Delta$ s (not shown) as expected, confirming the robustness of the DW and analysis protocol employed in the current study. Note that the diffusion distance corresponding to diffusion time of  $\Delta = 15 \sim 200$ ms ranges approximately from 5 $\mu$ m to 16 $\mu$ m in normal liver as estimated using Einstein's equation. Also note that our histological results confirmed that the livers were at different stages of fibrosis during different periods after CCl<sub>4</sub> administration (not shown) as expected.

**DISCUSSION AND CONCLUSION:** First, the decrease in ADC and  $D_{True}$  observed after CCl<sub>4</sub> insult (2-week to 8-week) likely resulted from the excessive extracellular matrix (ECM) especially collagen fiber deposition, one of four central features in the process of lipocyte activation induced by chronic liver injury [2, 9], leading to higher content of diffusion barriers in the extracellular space in the scar tissue of fibrotic liver. ADC and  $D_{True}$  decrease at early phase after CCl<sub>4</sub> insult (week 2 to week 4) might also be related to the proliferation of hepatic lipocyte which is another feature during lipocyte activation [2, 9]. Cell proliferation causes an increase in cell density and cell membranes. Cell membranes hinders and highly restricts the water diffusion process inside the cells [10]. In addition, high cell density results in intracellular water fraction increase and extracellular water fraction decrease in tissue, leading to more restricted water diffusion and lower  $D_{True}$  and ADC. In fact, it is widely believed that diffusion decreases in the tissue undergoing cell proliferation [11, 12]. Secondly, the mean  $D_{True}$  reductions between  $\Delta = 15$  and  $\Delta = 45$  ms, 45 ms and 200 ms, 15 ms and 200 ms in liver after 2-week CCl<sub>4</sub> insult were 8±2.2%, 14±3.4% and 21±4.1%, respectively; while those in liver after 4-week CCl<sub>4</sub> insult were 10±3.2%, 18±3.8% and 26±6.4%, respectively (Fig.3). The substantial decreases in  $D_{True}$  with  $\Delta$  at week 4 might be due to that more barriers, such as increased cell membranes and accumulated collagen fibers, were encountered by water molecules diffusing in intra- and extracellular space in fibrotic liver. Note that ADC and  $D_{True}$  at different stages of fibrosis varied with  $\Delta$  to different extents, suggesting that it may be desirable to use two different  $\Delta$ s to detect cellular changes caused by fibrosis. Finally, liver ADC and  $D_{True}$  of age-matched control group showed no significant differences over different time points of liver fibrosis at all  $\Delta$ s, confirming that the changes in ADC and  $D_{True}$  were caused by fibrosis rather than liver maturation. To conclude, true diffusion coefficient ( $D_{True}$ ) measured with long diffusion time is highly sensitive in detecting and accessing subtle changes in tissue microstructure during liver fibrogenesis.

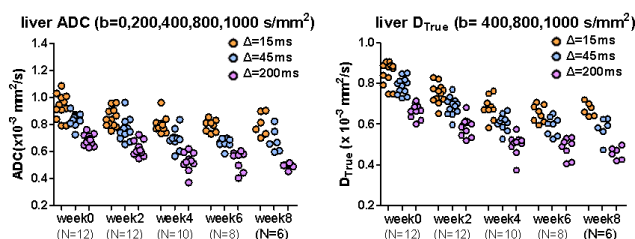
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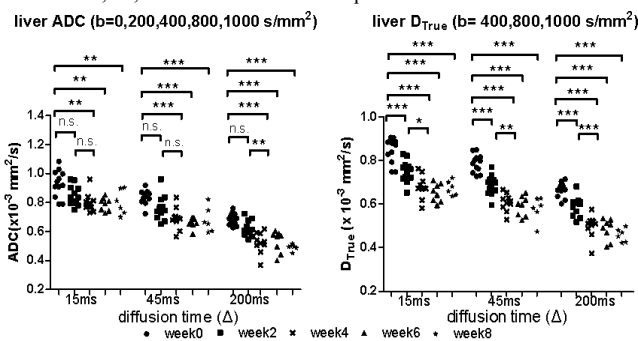
**Fig. 1** Representative T1W and B0 EPI images with diffusion time  $\Delta = 15, 45$  and 200ms from one adult SD animal after 8-week CCl<sub>4</sub> insult.



**Fig. 2** Typical mean DW signal decays, computed as the average of  $S_b/S_0$  from 6 fibrotic animals, for different diffusion times ( $\Delta$ s).



**Fig. 3** ROI measurements of liver ADC and  $D_{True}$  obtained from DWI with  $\Delta = 15, 45, 200$  ms at different time points of liver fibrosis.



**Fig. 4** Comparison of liver ADC and  $D_{True}$  measured at five different stages of liver fibrosis with three diffusion times using one-way ANOVA with \* for  $p < 0.05$ , \*\* for  $p < 0.01$ , \*\*\* for  $p < 0.001$ .