

Multiparametric MRI Assessment of Early Osteoarthritis in a Rabbit Model of Anterior Cruciate Ligament Transection

Jari Rautiainen¹, Mikko J. Nissi^{1,2}, Timo Liimatainen³, Walter Herzog⁴, Rami K. Korhonen¹, and Miika T. Nieminen^{5,6}

¹Department of Applied Physics, University of Eastern Finland, Kuopio, Finland, ²Center for Magnetic Resonance Research, University of Minnesota, Minneapolis, MN, United States, ³Department of Neurobiology, A.I. Virtanen Institute for Molecular Medicine, University of Eastern Finland, Kuopio, Finland, ⁴Faculty of Kinesiology, Engineering and Medicine, University of Calgary, Calgary, AB, Canada, ⁵Department of Diagnostic Radiology, Oulu University Hospital, Oulu, Finland, ⁶Department of Medical Technology, University of Oulu, Oulu, Finland

INTRODUCTION

Anterior cruciate ligament (ACL) injury is a common cause for post-traumatic degenerative changes of articular cartilage that is characteristic of osteoarthritis (OA), thereby promoting ACL transection (ACLT) as a reliable method for inducing experimental OA [1]. In the present study, early OA changes were investigated in an ACLT rabbit model. The specific aim was to investigate the sensitivity of selected MRI techniques (Adiabatic $T_{1\rho}$ & $T_{2\rho}$ [2], continuous wave (CW) $T_{1\rho}$, CPMG T_2 , Adiabatic double echo (DE) T_2 , T_1 during saturation, and relaxation along fictitious field (RAFF) [3]) to early degenerative changes in articular cartilage following ACLT. For reference, biomechanical testing of the load bearing cartilage was conducted.

MATERIALS AND METHODS

ACLT was unilaterally induced in the knees of skeletally mature New Zealand White Rabbits ($n = 8$). The contralateral (CTRL) joints were used as a non-transected control group. Femoral condyles of the joints were harvested for investigation 4 weeks after ACLT. The procedure was carried out according to the guidelines of the Canadian Council on Animal Care and was approved by the committee on Animal Ethics at the University of Calgary.

MRI was performed at 9.4 T (Oxford instruments Plc, Witney, UK) with a 19 mm quadrature RF volume transceiver (RAPID Biomedical GmbH, Rimpur, Germany) and Varian DirectDrive console (Varian Inc. Palo Alto, CA, USA). After biomechanical testing, the samples were immersed in FomblinTM inside a TeflonTM test tube. The selected MRI parameters were measured using a global magnetization preparation block followed by a multi-slice fast spin echo readout (TR = 5 s, ETL = 4, TE_{eff} = 5 ms, 256x128 matrix size, slice thickness 1 mm and FOV = 16x16 mm). Two slices were acquired during the TR period immediately after the preparation; the slices were positioned at the lateral and medial condyles, respectively, including the site of biomechanical testing. The parameters for the preparation block for different contrasts were: (1) *adiabatic $T_{1\rho}$* : a train of 0, 4, 8, 12 and 24 adiabatic full-passage (AFP) pulses with duration of 4.5 ms per pulse ($\gamma B_{1,max} = 2.5$ kHz), (2) *adiabatic $T_{2\rho}$* : a train of 0, 4, 8, 12 and 24 AFP pulses between adiabatic half-passage (AHP) pulses with duration of 4.5 ms per pulse ($\gamma B_{1,max} = 2.5$ kHz), (3) *CW $T_{1\rho}$* : CW spin-lock pulses ($\gamma B_1 = 1$ kHz) with durations of 0, 10, 20, 40, 80 and 160 ms embedded between AHP pulses, (4) *RAFF*: a train of 0, 2, 4 and 6 RAFF pulses of 9 ms length [4], (5) *T_1 saturation*: off-resonance irradiation with duration of 0.1, 0.3, 0.8, 2 and 4 s at 10 kHz offset ($\gamma B_1 = 250$ Hz), (6) *Double echo T_2* : TE = 4, 8, 16, 32, 64, 128 ms, TR = 5 s, (7) *CPMG T_2* : TE = 4, 8, 16, 32, 64, 128 ms, TR = 5 s. Relaxation time maps were calculated using MATLAB. 3-mm wide full-thickness ROI was drawn to the site of biomechanical testing in both slices and the mean \pm standard deviation (SD) of each parameter was determined. The full-thickness ROI was further split into superficial and deep halves.

The biomechanical properties of the samples were determined using indentation tests with a plane-ended indenter (dia. 1 mm). A stepwise stress-relaxation test was performed to determine the equilibrium elastic modulus of cartilage. Dynamic elastic modulus was determined from sinusoidal dynamic tests with a frequency of 1 Hz. The Poisson's ratios for cartilage were assumed to be 0.1 and 0.5 in determination of equilibrium and dynamic modulus, respectively [5].

Statistical analysis of the MRI and biomechanical data was performed with Kruskal-Wallis post hoc test using IBM SPSS Statistics 19.0 (New York, NY, USA).

RESULTS

Adiabatic $T_{1\rho}$, CW $T_{1\rho}$ and DE T_2 parameters showed a significant elongation in the superficial half of the cartilage in the ACL transected lateral femoral condyle (Figure 1). Furthermore, a significant elongation in superficial adiabatic $T_{1\rho}$ was seen in the medial compartment. For full thickness ROIs, statistically significant changes were observed in adiabatic $T_{1\rho}$, $T_{2\rho}$, and CW $T_{1\rho}$ between lateral condyles. The equilibrium elastic modulus, which is mostly affected by the proteoglycan content [6], was significantly reduced on the medial and lateral sides of the ACLT experimental joints as compared to the contralateral reference joints (Table 1). The dynamic elastic modulus that is sensitive to collagen integrity was also reduced at the lateral and medial compartments of the ACLT samples. Pooling all the samples, significant correlations were observed between MRI and biomechanical parameters. Most notably, for equilibrium modulus, the highest correlation was found with DE T_2 ($r = -0.465$, $p < 0.05$) and for dynamic modulus with CW $T_{1\rho}$ ($r = -0.462$, $p < 0.01$).

DISCUSSION

According to the present results rotating frame of reference (RFR) techniques are sensitive in detecting early cartilage degeneration, as detected by the slight yet significant change in the biomechanical properties. Moreover, RFR techniques appeared more sensitive in detecting early cartilage degeneration as compared to conventional techniques, namely T_2 and MT. Our findings are in concordance with a previous study reporting that $T_{1\rho}$ was superior to T_2 in differentiating OA patients from healthy subjects [7]. These findings may be due to the increased sensitivity of RFR techniques to very slow molecular motion, which is altered during the course of degeneration of the extracellular matrix. Unexpected finding in the present study is the low sensitivity of RAFF to the changes in cartilage after ACLT. While tissue degradation was detected by both MRI and biomechanical testing, the MRI parameters were not strongly associated with mechanical properties in this model. In conclusion, the high sensitivity of RFR techniques to early cartilage degeneration encourages to further investigate their implementation for the detection of early OA.

REFERENCES

- Nelson et al. OsteoArth Cart 14:114-119, 2006.
- Michaeli et al. J Magn Reson 169:293-299, 2004.
- Liimatainen et al. Magn Reson Med 64: 983-994, 2010.
- Liimatainen et al. Proc. ISMRM 2010, 836.
- Korhonen et al. J Biomech 35:903-909 2002.
- Korhonen et al. J Biomech 36:1373-1379, 2003.
- Stahl et al. Eur Radiol 19:132-143, 2009.

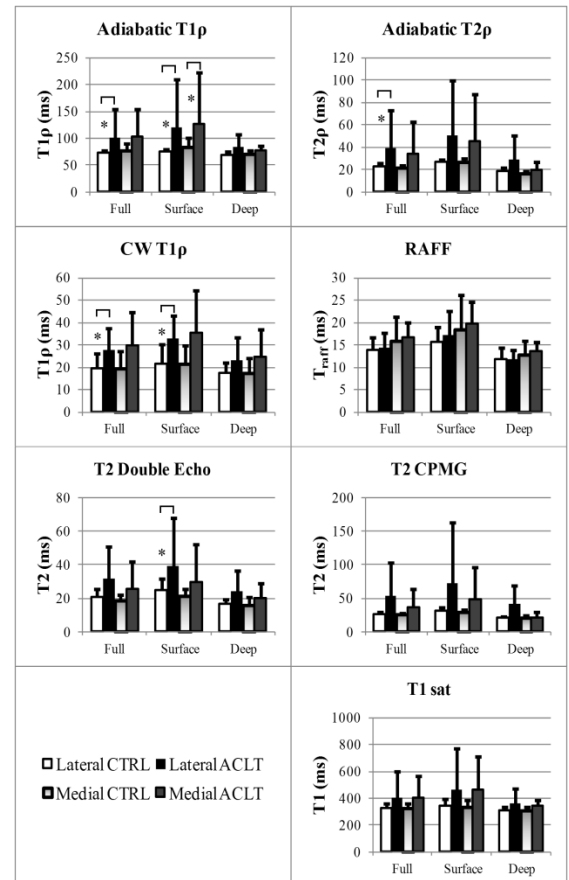


Figure 1. Mean \pm SD values of MRI parameters for full-thickness, surface and deep ROIs. * $p < 0.05$, comparison between ACLT and contralateral joint.

Table 1. Mean values (\pm SD) of cartilage equilibrium and dynamic elastic modulus. * $p < 0.05$, compared to contralateral.

		ACLT	CTRL
Equilibrium elastic modulus	Medial	0.41 \pm 0.30*	0.75 \pm 0.27
	Lateral	0.31 \pm 0.16*	0.55 \pm 0.18
Dynamic elastic modulus	Medial	5.15 \pm 1.80*	6.58 \pm 1.72
	Lateral	4.76 \pm 1.55*	6.87 \pm 1.47