

In Vivo Hyperpolarized ^{13}C -MRS Shows Abnormal Cardiac Metabolism in the PPAR α Knockout Mouse

Michael Dodd^{1,2}, Rosalind Bray¹, Vicky Ball¹, Mark Cole¹, Kieran Clarke¹, and Damian Tyler¹

¹Department of Physiology, Anatomy and Genetics, Oxford University, Oxford, OXON, United Kingdom, ²Department of Cardiovascular Medicine, Oxford University, Oxford, OXON, United Kingdom

Introduction: During fasting, plasma free fatty acids increase and stimulate their own catabolism through increased fatty acid oxidation. This is achieved, in part, via the activation of the peroxisome proliferator-activated receptors (PPARs), a group of nuclear response elements. PPAR α has a high binding affinity for fatty acids and functions to transcriptionally regulate the expression of a range of fatty acid oxidation genes. The PPAR α knockout mouse has been developed to investigate the role of PPAR α in metabolism and disease. The aim of this work was to assess the *in vivo* metabolic phenotype of the PPAR α -KO heart using hyperpolarized magnetic resonance spectroscopy. In particular the metabolism of [$1\text{-}^{13}\text{C}$]pyruvate was measured in fed and fasted mice.

Methods: *Animals* - Five 12-14 month old male PPAR α -KO mice and their littermate controls (129Sv, n = 5) received two hyperpolarized scans on two separate days. Before the first scan, the mice were provided with food and water *ad libitum* and scans were performed between 7 am and 11 am during the fed state. For the second scan, the mice were fasted overnight (minimum of 18 hours), with free access to water.

Hyperpolarized ^{13}C MRS Protocol - [$1\text{-}^{13}\text{C}$]pyruvate was hyperpolarized and dissolved as previously described [1,2]. An aliquot of 0.2 ml of 80 mM hyperpolarized [$1\text{-}^{13}\text{C}$]pyruvate solution was injected over 10 s via a tail vein catheter into an anaesthetised mouse positioned in a 7 T MR scanner. Spectra were acquired for 1 min following injection with 1 s temporal resolution, using a 15° RF excitation pulse. Signal was localised to the heart using a home-built ^{13}C RF surface coil. Quantified peak areas were input into a kinetic model described by Atherton *et al* [3]. The rate of exchange of the ^{13}C label between pyruvate and its metabolites was termed ^{13}C label incorporation. ^{13}C label incorporation into the bicarbonate pool is a sensitive measure of pyruvate dehydrogenase (PDH) flux in the rat [3].

Results: Fed PPAR α -KO mice had a 41% increase in PDH flux when compared to fed control animals (figure 1, p < 0.05), whilst incorporation of the ^{13}C label into lactate and alanine pools was unchanged between groups. Control animals in the fasted state had a $10 \times 10^{-4} \text{ s}^{-1}$ reduction (55%) in PDH flux when compared to the same animals in the fed state (p < 0.05). Interestingly the fasted PPAR α -KO mice also had a $10 \times 10^{-4} \text{ s}^{-1}$ decrease in PDH flux (45%) when compared to their fed state (p < 0.001). However, the fasted PPAR α -KO mouse had a 50% higher PDH flux than control fasted animals (p < 0.05).

Discussion: The normal cardiac response to fasting is a greater reliance on fatty acids and a shift away from glucose oxidation. The breakdown of fatty acid stores results in elevated plasma fatty acid levels which stimulate their own catabolism through increased fatty acid oxidation. This leads to decreased PDH flux that “spares” pyruvate for oxaloacetate production and gluconeogenesis [4]. The decrease in PDH flux is partly mediated by increased expression of PDH kinase (PDK) 4, the inhibitor of PDH [5]. In fasted wild type mice, PPAR α increases several key fatty acid oxidation proteins, as well as PDK4. In the fed PPAR α -KO mice there is a significant increase in PDH flux that is potentially due to decreases in PDK4 expression. Fasting led to a highly significant decrease in PDH flux in both control and PPAR α -KO mice. Interestingly the overall decrease in PDH flux in the control and PPAR α -KO mice was the same, indicating that this alteration is PPAR α independent and is a response to increased fatty acid supply and utilization. Previously it has been shown that PPAR α expression decreases with age [6], so future work will assess PDH flux in younger control and PPAR α -KO mice to determine whether this is an age related alteration in PDH flux control.

References: [1] Golman, K., *et al.* *PNAS*, 2006. **103**(30): p.11270-5 [2] Schroeder, M.A., *et al.* *PNAS*, 2008. **105**(33): p. 12051-6. [3] Atherton, H.J., *et al.* *NMR Biomed*, 2011. **24**(2): p. 201-8. [4] Huang, B., *et al.* *Diabetes*, 2002. **51**(2): p. 276-83. [5] Wu, P., *et al.*, *Biochem J.*, 1998. **329** p 197-201 [6] Iemitsu, M., *et al.*, *A.J.P Heart Circ. Physiol.*, 2002. **283**(5): p. H1750-60

Acknowledgements: This study was supported by the British Heart Foundation and GE Healthcare.

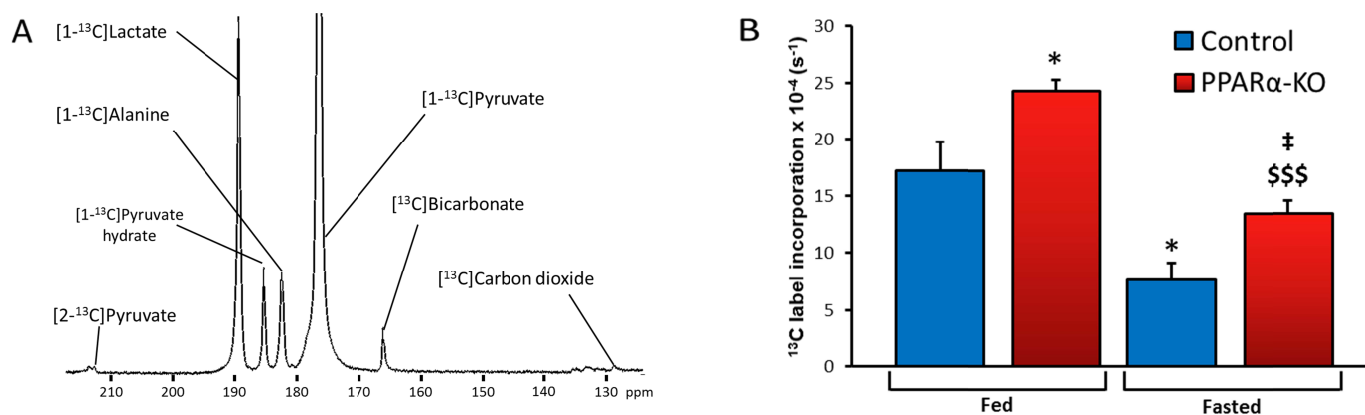


Figure 1 – A) Summed *in vivo* spectra from a fed PPAR α -KO mouse acquired over 45 seconds. B) ^{13}C label incorporation into bicarbonate (PDH flux). During the fed state there is a significant increase in PDH flux in PPAR α -KO mice. During fasting there is a significant reduction in PDH flux in both controls and PPAR α -KO mice. Values are mean data and error bars are S.E.M. Significant differences between mean values were determined by analysis of variance (ANOVA) followed by student two tailed t-test. * p < 0.05 compared to fed control, \$\$\$ p < 0.001 compared to fed PPAR α -KO and § p < 0.05 compared to fasted control