

Hyperpolarized Helium Measurements of $P_{A}O_2$ Correlate with Neutrophil Inflammation in the Rat Bleomycin Model

P. Mongkolwisetwara¹, E. B. Arguiri², K. Emami¹, Y. Xin¹, N. N. Kuzma¹, S. J. Kadlecck¹, Y. Xu¹, H. Profka¹, M. Christofidou-Solomidou², M. D. Rossman², M. Ishii³, and R. R. Rizi⁴

¹Radiology, University of Pennsylvania, Philadelphia, Pennsylvania, United States, ²Pulmonary Division, University of Pennsylvania, Philadelphia, Pennsylvania, United States, ³Otolaryngology–Head & Neck Surgery, Johns Hopkins University, Baltimore, Maryland, United States, ⁴Radiology, University of Pennsylvania, Philadelphia, Pennsylvania

INTRODUCTION: Hyperpolarized (HP) ^3He MRI is a technique sensitive to both lung structure (airway size through Apparent Diffusion Coefficient, ADC) and function (alveolar PO_2 and fractional ventilation, r) through direct imaging of respiratory gas molecules. In this work, we investigated correlated changes of these metrics with bronchoalveolar lavage (BAL) measurements and biochemistry in a rat model of interstitial fibrosis secondary to bleomycin.

METHODS: Male Sprague Dawley rats ($n=9$, 300–350g body weight) were given intra-tracheal bleomycin and 7 (7BR) and 21 (21BR) days after intratracheal bleomycin, they underwent HP ^3He MRI to measure r , ADC and $P_{A}O_2$. The animals were intubated and connected to a custom-designed small animal ventilator. This ventilator is capable of delivering the breathing gas with an accuracy of $\pm 100\mu\text{L}/\text{breath}$ and real-time monitoring of peak inspiration pressure (PIP). For ventilation imaging, a series of 10 HP gas breaths ($^3\text{He}:\text{O}_2$, $\text{FIO}_2=20\%$) was delivered to the rat at the designated tidal volume $V_T = 1.0\text{ml}/100\text{g}$, and one image was acquired after each breath during a 350-ms breath-hold. The HP ^3He signal build up in the rat lung was then recursively solved for r to yield the fractional ventilation map, as described earlier [1]. For ADC imaging, rats were ventilated with five identical breaths of HP $^3\text{He}:\text{O}_2$ (4:1) at the designated inflation level followed by a 3-sec breath-hold during which five diffusion-weighted images were acquired corresponding to b -values = 0.00, 3.73, 2.18, 1.00 and 0.00 s/cm^2 . This procedure was repeated immediately with identical but reversed polarity diffusion gradient b -values. These 10 diffusion-weighted images were then combined to yield the ADC map of the imaged slice according to a double-acquisition diffusion imaging scheme described earlier [2]. For $P_{A}O_2$ imaging, rats inhaled a series of 5 breaths of $^3\text{He}:\text{O}_2$ followed by a short 6-sec breath-hold, during which images were acquired at a set of predefined delay times, and the resulting images corresponding to the same slice/delay combination were then averaged and fit to a model of O_2 -induced decay and respiratory gas redistribution as described earlier [3]. Images were acquired using a diffusion-weighted gradient echo imaging pulse sequence with centric phase-encoding in a 50-cm bore 4.7-T MRI scanner (Varian Inc) equipped with a 12-cm, 25-G/cm gradients and a 2-3/4"-ID quadrature 8-leg birdcage body coil (Stark Contrast). Images were acquired in the middle coronal slice of the rat lung with the following imaging parameters: $\text{FOV}=6\times 6\text{cm}^2$, $\text{ST}=6\text{mm}$, $\text{MS}=64\times 64$, $\alpha=4\sim 5^\circ$, $\text{TR}=6.6\text{ms}$, and $\text{TE}=4\text{ms}$. Diffusion sensitizing gradient was applied along the phase-encoding (L–R) direction with the following timing parameters: $\Delta=1\text{ms}$, $\delta=200\mu\text{s}$, and $\tau=180\mu\text{s}$ according to the naming convention of [4]. Upon conclusion of imaging, BAL was performed for measurement of white blood cell (WBC) numbers and differential, and BAL protein content as a measure of lung damage. The right lung was fixed for histology and the left lung for measurement of hydroxyproline, a measure of fibrosis. Healthy rats (HR) were similarly tested. Values were expressed as mean \pm SD and statistical significance was determined by pairwise t-tests.

RESULTS AND DISCUSSION: Figure 1 shows a representative map of $P_{A}O_2$, ADC, and r in HR, 7BR, and 21BR rats. Figure 2 shows that the overall mean of ADC were not significantly reduced in 7BR ($0.25\pm 0.10\text{ cm}^2/\text{s}$) and 21BR ($0.24\pm 0.07\text{ cm}^2/\text{s}$) compared to HR ($0.31\pm 0.11\text{ cm}^2/\text{s}$). The means of r were also not significantly reduced in 7BR (0.23 ± 0.15) compared to the HR (0.33 ± 0.17) and returned towards normal in 21BR (0.31 ± 0.16). In contrast, Figure 3 shows the $P_{A}O_2$ was significantly ($p < 0.05$) increased in 7BR ($175.0\pm 24.8\text{ mbar}$) compared to HR ($108.2\pm 4.4\text{ mbar}$) and returned toward HR in 21BR ($99.07\pm 21.55\text{ mbar}$). Both the number ($r = 0.868$) and percent ($r = 0.833$) of neutrophils in the BAL fluid was significantly ($p < 0.01$) correlated with the $P_{A}O_2$ but there was no significant correlation between BAL cells or protein with ADC or r . Hydroxyproline was unchanged in 7BR but was significantly increased in 21BR (HR = 64.3 , 7BR = 53.1 , 21BR = $95.7\text{ ug}/\text{ug}$ lung tissue, $p < 0.05$).

CONCLUSION: In the rat model of pulmonary fibrosis due to bleomycin, $P_{A}O_2$ correlated with the extent of neutrophil inflammation in the lung. This suggests that $P_{A}O_2$ measured by

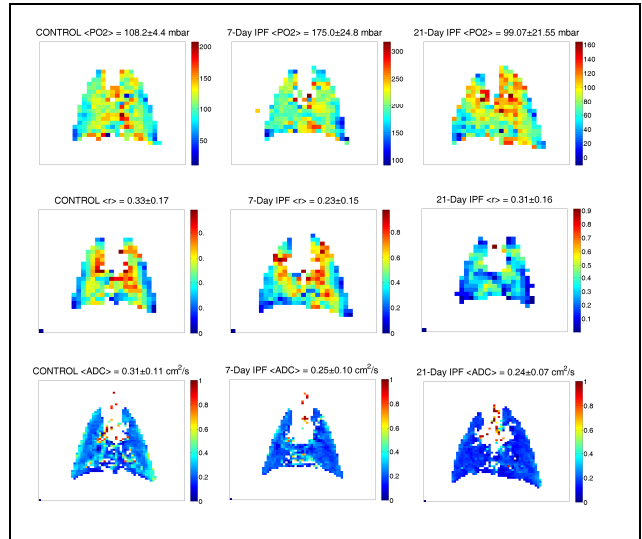


Figure 1. Representative maps of $P_{A}O_2$, ADC, and r in HR, 7BR, and 21BR rats.

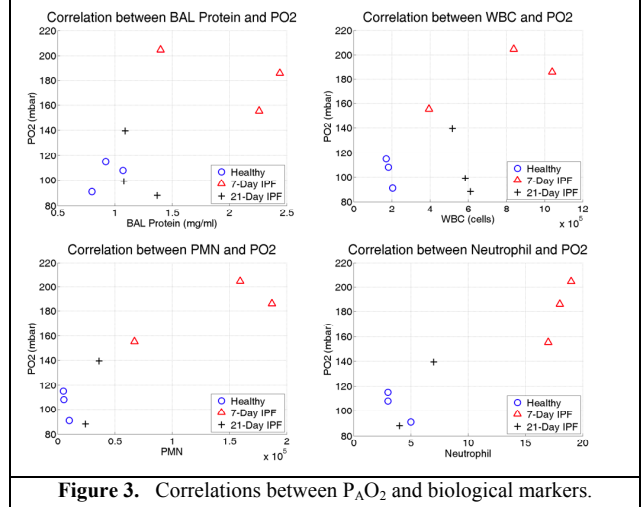


Figure 3. Correlations between $P_{A}O_2$ and biological markers.

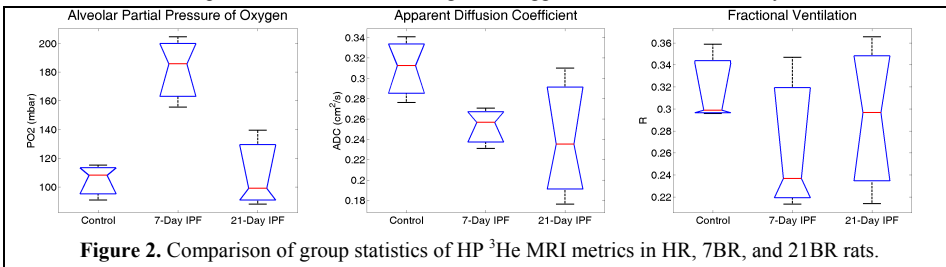


Figure 2. Comparison of group statistics of HP ^3He MRI metrics in HR, 7BR, and 21BR rats.

HP ^3He MRI can be a sensitive indicator of pulmonary inflammation and may help with predicting prognosis, and helping with drug development as a non-invasive measure of lung function.

REFERENCES: [1] Emami K, *et al.*, Magn Reson Med. 2010; [2] Emami, K, *et al.* Proc Intl Soc Mag Reson Med 2007; [3] Kadlecck *et al.*, Proc Intl Soc Mag Reson Med 2009; [4] Yu, J, *et al.* J Magn Reson Med 2007.