

# Effect of graded O<sub>2</sub> challenge on vascular and metabolic parameters

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**INTRODUCTION:** Calibrated fMRI relies on an iso-CMRO<sub>2</sub> challenge to obtain the calibration factor, M (1-3). Typically, the calibration experiment is performed using hypercapnia via inhalation of CO<sub>2</sub> gas mixture. However, recent evidence suggested that CO<sub>2</sub> may cause a suppression of neural activity by 10-15% due to acidosis in the brain parenchyma (4). Thus O<sub>2</sub> challenge became a preferred option for the calibration experiment (3). However, the assumption that O<sub>2</sub> challenge is iso-CMRO<sub>2</sub> has not been validated either. The present study will investigate whether physiologic manipulation of O<sub>2</sub> content in the arterial blood will change brain metabolism. We used a recently developed TRUST MRI technique (5-6) to monitor the subject's CMRO<sub>2</sub> while altering the O<sub>2</sub> concentration in the inspired air. Our data suggest that O<sub>2</sub> content changes brain metabolism and an inverse relationship was observed between CMRO<sub>2</sub> and arterial O<sub>2</sub> content which accounts for both hemoglobin-bound and dissolved O<sub>2</sub> in the blood.

**METHODS:** Nine young healthy subjects (age 28±5yrd, 3 F 4 M) were studied on a 3T Philips System. Each subject participated in a 50 min session with a graded O<sub>2</sub> challenge in which they breathed four levels of O<sub>2</sub> in the following order: 21%O<sub>2</sub> for 8min, 14%O<sub>2</sub> for 18min, 50%O<sub>2</sub> for 15min, and 98%O<sub>2</sub> for 12min (see color coding in Fig. 1). The duration of each breathing period was determined based on time needed to reach a new steady state after switching the gas (data from tests outside MRI). For example, after switching to 14% O<sub>2</sub> (hypoxia), it takes about 10 min to reach stabilization. The criteria for stabilization used Pulse-Oximeter-derived arterial oxygen saturation fraction (Y<sub>a</sub>) for hypoxia and oxygen-sensor-derived oxygen partial pressure (p<sub>a</sub>O<sub>2</sub>) for hyperoxia, as exhaled O<sub>2</sub> has reached plateau and does not change during hyperoxia. An index for total arterial oxygen content was calculated for each O<sub>2</sub>-breathing condition by accounting for both hemoglobin-bound and dissolved O<sub>2</sub>:  $[O_2]_a = (Y_a \times C_h + p_a O_2 \times C_d) / C_h$ , where C<sub>h</sub> (in unit of μmol O<sub>2</sub>/100ml blood) and C<sub>d</sub> (μmol O<sub>2</sub>/100ml blood/mmHg O<sub>2</sub> tension) are associated with hemoglobin oxygen-carrying capacity and plasma's oxygen-dissolving capacity, respectively. The term in the parenthesis was divided by C<sub>h</sub> so that the final index is in units of saturation fraction (%). Note that in this study  $[O_2]_a$  could reach a value greater than 100% because of the large amount of dissolved oxygen during 50% and 98% O<sub>2</sub> breathing periods.

At each level of O<sub>2</sub> breathing (after stabilizing), four physiologic parameters were determined, including venous oxygen saturation fraction (Y<sub>v</sub>), oxygen extraction fraction (OEF=[O<sub>2</sub>]<sub>a</sub>-Y<sub>v</sub>), CBF and CMRO<sub>2</sub> (i.e. OEF×CBF). These measurements would allow a complete characterization of vascular and metabolic responses to O<sub>2</sub> challenge, and the inter-relationship among these parameters are illustrated in Fig. 2. Y<sub>v</sub> was estimated using a T2-Relaxation-Under-Spin-Tagging (TRUST) MRI technique via the measurement of pure blood T2 in the major vein sagittal sinus (5). TRUST MRI uses the spin labeling principle to separate flowing blood signals from static tissue, thereby obtaining pure blood signal. T2-weighting was then applied in the sequence with non-slice-selective preparation pulses (inter-pulse interval τ<sub>CPMG</sub>=10ms). Other imaging parameters were: voxel size 3.44x3.44x5mm<sup>3</sup>, TR=8000ms, TI=1200ms, four TEs: 0ms, 40ms, 80ms and 160ms, duration 3.5 min. CBF across the sagittal sinus was measured using a phase-contrast (PC) flow velocity technique with the following parameters: voxel size 0.45x0.45x5 mm<sup>3</sup>, maximum velocity 80cm/s, duration 30sec. CMRO<sub>2</sub> was calculated using the Fick principle and was based on a previous study (6) except that the dissolved oxygen was also accounted in the present study.

In addition, PC MRI was performed continuously during the transit periods, providing a time course of CBF changes during the entire scan session.

For statistical analysis, correlation coefficients between  $[O_2]_a$  and the parameters described above were calculated for each subject, which were then converted to z-scores using a Fisher transform and the group z-scores were compared to 0 using t tests.

**RESULTS and DISCUSSION:** Fig. 1 shows the time courses of arterial O<sub>2</sub> content ( $[O_2]_a$ ) and CBF in the sagittal sinus (from the continuous PC MRI measurement) during the graded O<sub>2</sub> challenge. Correlation analysis on the steady state data revealed an inverse relationship between CBF and  $[O_2]_a$  (P<0.001) (Fig. 2). Similarly, Y<sub>v</sub> was correlated with  $[O_2]_a$  (P<0.001) (Fig. 2). The arterial-venous O<sub>2</sub> difference, OEF, was not correlated with  $[O_2]_a$  (P=0.64), suggesting that tissue extracts identical amount of O<sub>2</sub> from each unit of blood flow regardless the level of arterial O<sub>2</sub> content. Interestingly, CMRO<sub>2</sub> was found to be inversely correlated with  $[O_2]_a$  (P<0.001) (Fig. 2). Fig. 3 shows a scatter plot between  $[O_2]_a$  and relative CMRO<sub>2</sub> with respect to the room air value for all O<sub>2</sub> conditions and all subjects. Note that different subjects had different  $[O_2]_a$  responses to hypoxia (breathing 14% O<sub>2</sub>), possibly due to variations in compensatory responses in respiration. This provides a sufficient dynamic range to allow us to compare  $[O_2]_a$  and CMRO<sub>2</sub> across subjects within the hypoxia condition (blue line in Fig. 3). An inverse correlation (r=0.81, P=0.01) was again observed. The  $[O_2]_a$  variations in hyperoxia across subjects were minimal, thus no further correlation was assessed.

In this study, we characterized brain vascular and metabolic responses to graded O<sub>2</sub> challenge. It appears that O<sub>2</sub> has a suppressive effect on CBF and oxygen metabolism. The reduction in CBF in response to O<sub>2</sub> has been reported previously (7). The suppression on metabolism has not been reported in the literature but appears to be consistent with the notion of a coupling between neural and vascular responses, i.e. reduced CMRO<sub>2</sub> may be the underlying reason for a lower CBF. The present study suggests that O<sub>2</sub>-based challenge may not be suitable for fMRI calibration experiment as it changes CMRO<sub>2</sub>. These findings may also be useful in understanding the effect of O<sub>2</sub> therapy in various neurologic diseases.

**REFERENCES:** 1) Davis et al. Proc Natl Acad Sci 95:1834 (1998); 2) Hoge et al. MRM 42:849 (1999); 3) Chiarelli et al. Neuroimage 37:808 (2007); 4) Zappe et al. Cereb Cortex 18:2666 (2008); 5) Lu and Ge MRM, 60:357 (2008); 6) Xu et al. MRM 62:141 (2009); 7) Sicard et al. Neuroimage 25:850 (2005).

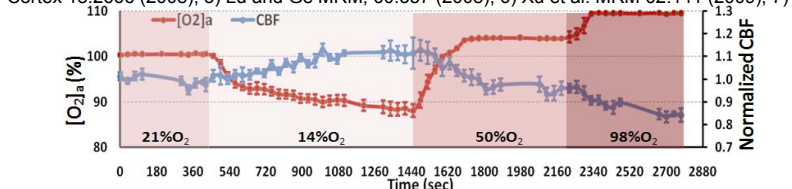


Fig.1 The time courses of arterial oxygen content ( $[O_2]_a$ , combining the hemoglobin-bound O<sub>2</sub> and dissolved O<sub>2</sub>) and CBF. Each blue dot indicates one CBF measurement. The error bars indicate standard errors across subjects (N=9).

Fig. 3 Scatter plot between  $[O_2]_a$  and relative CMRO<sub>2</sub> (with respect to the room air value).

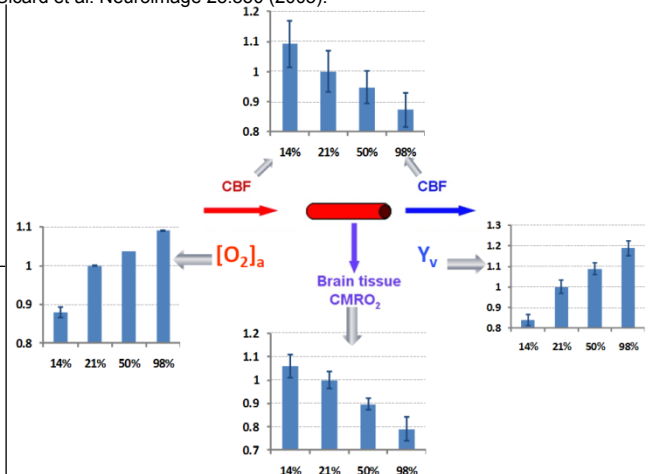
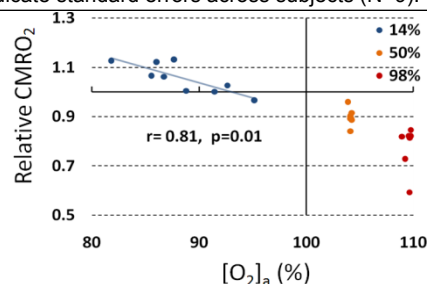


Fig. 2 The relationship among vascular and metabolic parameters in the brain and their fractional changes under various O<sub>2</sub> levels; the error bars indicate standard errors (N=9).