## In Vivo T2 and T1rho Mapping of Rabbit Disc using Spin-Lock sequence at 3T

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**[Introduction]** Loss of proteoglycan (PG), a major component of intervertebral discs, is signified in early disc changes in degenerative disc disease (DDD) [1]. A non-invasive in vivo imaging technique to detect PG change would be useful in diagnosis of early DDD. A strong association between changes in  $T_{1rho}$  relaxation time and PG loss in discs was reported in a previous study [2]. In order to systemically assess this association in an animal model [3], we developed a dual-tuned (DT) RF coil and MR sequences to measure both  $T_2$  and  $T_{1rho}$  relaxation times in rabbit lumbar discs in vivo at 3T.

[Methods and materials] All scans were performed at a 3T human scanner (Siemens Medical Solutions, Germany). Twelve New Zealand white rabbits were studied:  $\leq 1$  year old, female, and  $5.2 \pm 0.4$  kg. MR imaging was performed using an in-house multi-channel DT RF coil (designed for

human knee imaging) that consisted of 4-channel transceivers with 120-mm diameter and 150-mm height [4]. Rabbits fit snuggly within the coil and were positioned supine at the center of coil. Form pads were inserted between the body and coil to minimize motion. T2-weighted TSE, DESS (data not shown), spin echo (SE), and spin-lock (SL) images with zero or nonzero B<sub>1</sub> SL preparation were acquired in the sagittal plane (**Fig. 1**): TSE – TR/TE = 3500/109 ms, resolution = 0.6×0.6×3 mm<sup>3</sup> (**Fig. 1A**); DESS – TR/TE = 14/5 ms, resolution = 0.6×0.6×3 mm<sup>3</sup> (**Fig. 1B**); SL multi-segmented SSFP - TR/TE/TSL(time of SL) = 7016/1.95/10 – 150 ms, B<sub>1</sub> = 387 – 473 Hz depending on SAR limit, resolution = 0.86×0.86×4 mm<sup>3</sup> (**Fig. 1C**). SL preparation consisted of two opposite polarity 90° RF pulses at the beginning and TSL, and low frequency SL B<sub>1</sub> pulse between them. A composite 180° RF pulse was also added before and after the

90°<sub>x</sub>. 180°<sub>y</sub>. 180°<sub>y</sub>. 90°<sub>x</sub>.

B<sub>1</sub>

y+ y+ y- y- y+

Spin-Lock preparation

[5]). MR signals were fitted using monotonic exponential curve, a\*exp(-TE/T<sub>2</sub>)+b or a\*exp(-TSL/T<sub>2</sub>(1rho))+b in pixel-by-pixel base; likewise for the segmented lumbar L2-L7 disc regions by intensity-threshold in ROI (white box

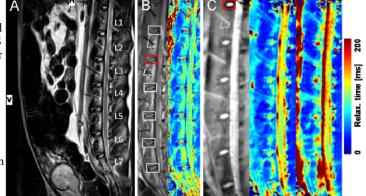
low frequency SL B<sub>1</sub> pulse (see left figure

in **Fig. 1B**) (**Figs. 2A** and **B**). Three different MR relaxation times were measured and compared across the lumbar spine discs from L2 to L7 (**Figs. 2C - E**).

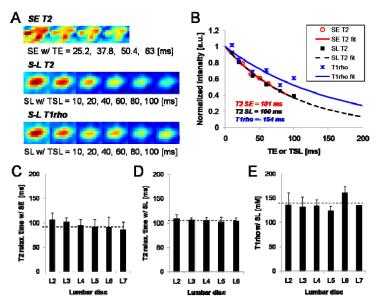
[Results and conclusions] High-resolution and high-contrast proton anatomy images of the rabbit disc were acquired in all 12 rabbits (Fig. **1A**). High T<sub>2</sub> contrast in the discs was evident in the T2-weighted TSE images. T<sub>2</sub> and T<sub>1rho</sub> maps of the spine were successfully acquired (Figs. 1B and C). The measured MR relaxation times in the lumbar discs were  $96.7 \pm 9.5$  (N = 12),  $105 \pm 3.9$  (N = 10), and  $138.3 \pm 15.8$  ms (N = 9) for  $T_2$  w/ SE,  $T_2$  w/ SL, and  $T_{1\text{rho}}$ , respectively. These measurements are similar to those reported in human lumbar discs  $(92.3 \ 6 \pm 27.2 \ \text{ms for T}_2 \ \text{and } 133.1 \ 6 \pm 13.8 \ \text{ms for T}_{1\text{rho}}) \ [6]. \ \text{In}$ conclusion, we successfully obtained high-resolution, high-contrast T<sub>2</sub> and T<sub>1rho</sub> images and measured T<sub>2</sub> and T<sub>1rho</sub> in normal rabbit lumbar spine discs. This was achieved using in-house dual-tuned RF coil and SL sequence at 3T. Further optimization of B<sub>1</sub> field homogeneity is needed for more stable T<sub>2</sub> and T<sub>1rho</sub> mapping (e.g., using volume excitation and multi-receiving RF coil array). Further study will also be required to demonstrate the difference in T<sub>2</sub> and T<sub>1rho</sub> between normal and degenerative disc models in rabbits in order to validate these imaging biomarkers for degenerative disc disease.

[Reference] 1, Kent et al., *Chiropr Osteopat* 13 (2005). **2**, Wheaton et al., *MRM*. 54:1087–1093 (2005). **3**, Sowa et al., *Spine*,33:1821-8 (2008). **4**. Kim et al., *ISMRM*, 2011 submitted. **5**, He et al., *ISMRM*, 2011 submitted. **6**, Blumenkrantz et al., *MRM*, 63:1193-1200 (2010).

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**Fig. 1** In-vivo MR imaging of rabbit disc. **A**, T2-weighted TSE (TR/TE = 3500/109 ms) of lumbar spine from L1 to L7. **B**, T2 mapping with SE; left-raw, right-T2 map. C, T2 and T<sub>1rho</sub> mapping with SL; left-raw, middle-T<sub>2</sub> map, right-T<sub>1rho</sub> map. Note high T<sub>1rho</sub> values in the discs.



**Fig. 2**  $T_2$  and  $T_{1rho}$  measurement in the rabbit lumbar disc. **A**, Disc images (red box in **Fig. 1B** and **C**) with varying TEs and TSLs; upper-SE  $T_2$ , middle-SL  $T_2$ , lower-SL  $T_{1rho}$  image. **B**, Time plot of disc image intensity with TE and TSL. **C** – **E**,  $T_2$  with SE,  $T_2$  with SL, and  $T_{1rho}$  measurement in lumbar disc;  $T_2$  w/ SE = 96.7 ± 9.5,  $T_2$  w/ SL = 105 ±3.9, and  $T_{1rho}$ =138.3 ±15.8 ms.  $T_{1rho}$  values were higher than the  $T_2$  values.  $T_2$  measurements were similar by SE and SL sequences.