

Two-pool compartmental modeling of balanced SSFP and CEST

K. L. Desmond¹, S. Deoni², S. Kolind², and G. J. Stanisz^{1,3}

¹Medical Biophysics, University of Toronto, Toronto, ON, Canada, ²Oxford University, Oxford, United Kingdom, ³Imaging Research, Sunnybrook Health Sciences Centre, Toronto, ON, Canada

Introduction: Chemical exchange saturation transfer has been used as a contrast mechanism for the imaging of cancer[1] and stroke[2], and is sensitive to the exchange rate between labile protons and unrestricted liquid water. Endogenously, several exchanging groups have been observed to contribute to the effect, mainly amide[1] and glycosaminoglycans[3] which have exchangeable groups that resonate within ~5ppm of the water frequency, and additionally contrast agents have been developed to exploit the CEST effect, including PARACEST[4] and which generally resonate on the order of 10-100 ppm shift from water. The standard CEST acquisition involves the application of a long RF saturation pulse such that a steady state is achieved between T1 recovery and saturation, with a resulting TR of 10s or longer. This is often referred to as the continuous wave (CW) CEST method. Several methods to reduce the scan time for CEST imaging have been introduced, including pulsed CEST[5-7]. This abstract simulates the utility of a balanced SSFP sequence for the characterization of chemical exchange, which promises a short TR and no waste of signal information due to spoiling.

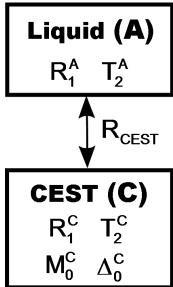
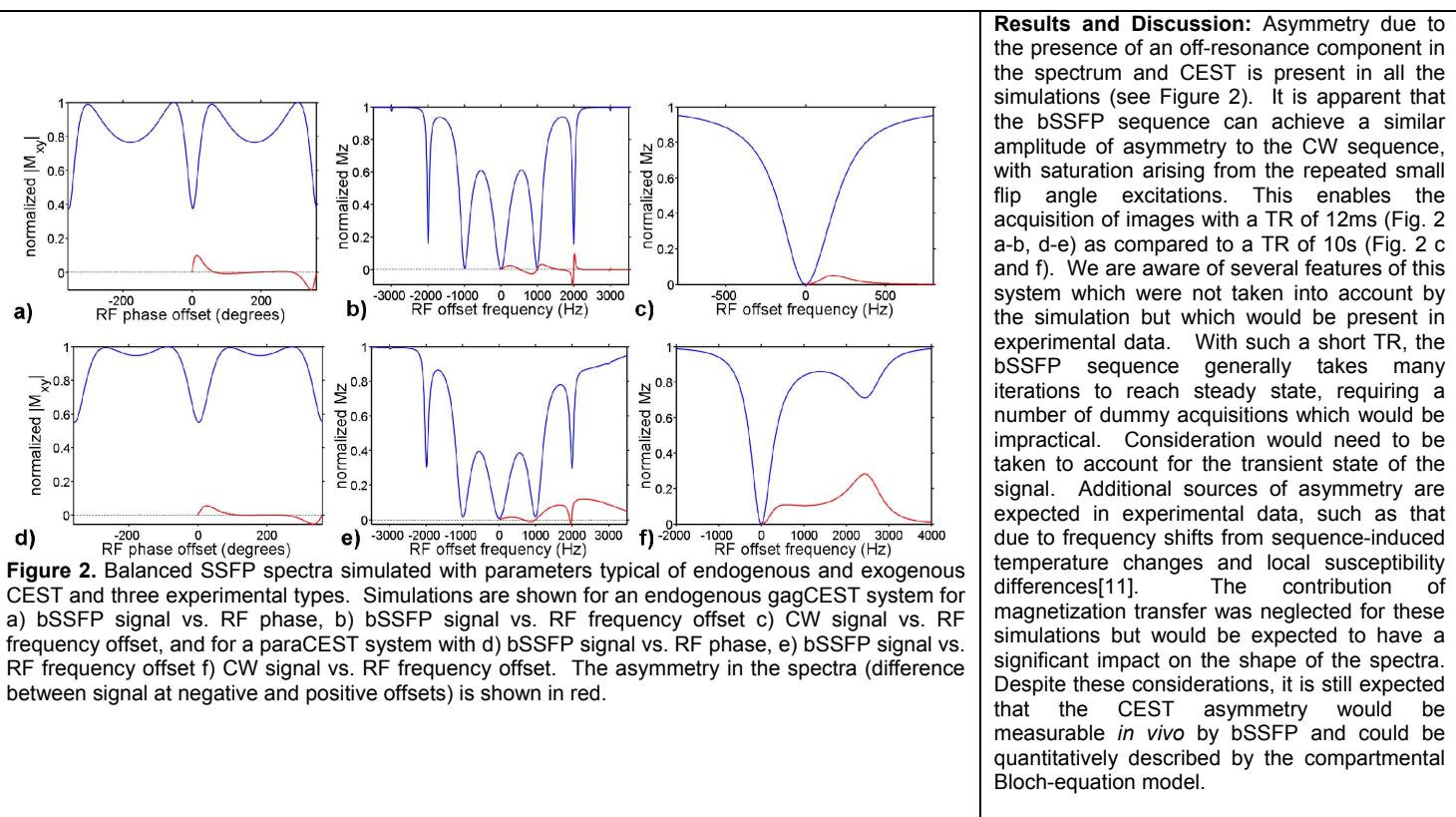


Figure 1.

Method: A simulation was developed which followed the behaviour of the liquid water and CEST proton magnetization vectors under the application of the balanced SSFP sequence. The spins were governed by a two-pool compartmental model of exchange (Fig. 1) in addition to effects of RF and T1, T2 relaxation[8]. The parameters which describe this model are as follows: M_{0c} (relative concentration of protons associated with the CEST pool), $T2_a$ and $T2_c$ ($T2$ of water and CEST pool respectively), Δ_{0c} (the frequency offset between the CEST pool and the free water pool) and R_{cest} (rate of chemical exchange between the CEST and free water pools). We modeled two different cases, simulating endogenous CEST from glycosaminoglycans (gagCEST) with $M_{0c} = 0.007$, $T2_c = 0.03s$, $\Delta_{0c} = 1\text{ppm} = 128\text{ Hz}$ at 3T, $R_{cest} = 1000\text{ Hz}$ and exogenous CEST from a hypothetical paraCEST agent with $M_{0c} = 0.01$, $T2_c = 0.08s$, $\Delta_{0c} = 20\text{ppm} = 2550\text{ Hz}$ at 3T, $R_{cest} = 2500\text{ Hz}$. $T1_a$ and $T2_a$ were assumed to be 0.8 and 0.08s respectively. Two different approaches to the collection of CEST spectra with balanced SSFP were investigated: 1. Magnitude of the transverse magnetization as a function of RF phase[9, 10] 2. Magnitude of the longitudinal magnetization as a function of RF offset frequency (flip angle = 15° and TR = 12ms for both approaches). These were compared to a third experiment type, the standard CW CEST experiment (saturation pulse amplitude = 0.4 μT for 10s). For all experiment types it was assumed that enough repetitions had been performed so that the magnetization was at steady state.



Conclusion: A quantitative model of CEST has been adapted to describe the signal observed with the bSSFP experiment. Balanced SSFP sequences show promise for the measurement of CEST asymmetry and can be performed with a TR a hundred times less than that required for traditional CW CEST experiments.

References: The authors would like to thank Karla Miller for helpful discussions.

1. Zhou, J., et al., Nat Med, 2003. **9**(8): p. 1085-90.
2. Sun, P.Z., et al., MRM, 2007. **57**(2): p. 405-10.
3. Ling, W., et al., Proceedings of the National Academy of Sciences, 2008. **105**(7): p. 2266.
4. Aime, S., et al., MRM, 2002. **47**(4): p. 639-48.
5. Desmond, K.L. and G.J. Stanisz, Proc. of the ISMRM, 17th Annual Meeting, Honolulu, United States, 2009.
6. Dixon, W.T., et al., MRM **63**(1): p. 253-256.
7. Sun, P.Z., et al., MRM, 2008. **60**(4): p. 834-841.
8. Woessner, D.E., et al., MRM, 2005. **53**(4).
9. Miller, K.L., MRM **63**(2): p. 385-395.
10. Scheffler, K. and S. Lehnhardt, European radiology, 2003. **13**(11): p. 2409-2418.
11. Miller, K.L., S.M. Smith, and P. Jezzard, MRM **63**(2): p. 396-406.