

T1 and T2* responses to hypercapnic and hyperoxic gases in normal tissue are independent of the order of gas delivery

J. D. Winter^{1,2}, M. Estrada¹, and H-L. M. Cheng^{1,3}

¹Physiology and Experimental Medicine, The Hospital for Sick Children, Toronto, Ontario, Canada, ²Research and Development, IMRIS, Winnipeg, Manitoba, Canada, ³Medical Biophysics, University of Toronto, Toronto, Ontario, Canada

Introduction: Quantitative MRI measures of T₁ and T₂* relaxation times offer a noninvasive means to indirectly monitor tissue O₂ content. T₁ oxygen-dependence is primarily based on the presence of weakly paramagnetic molecular O₂ dissolved in blood plasma and interstitial space; whereas, T₂* oxygen dependence is based on the signal loss induced by local magnetic field inhomogeneities generated by paramagnetic deoxy-hemoglobin. The clinical rationale for gas challenge studies originate from radiotherapy strategies aimed at augmenting tissue oxygenation via increased fraction of inspired O₂ (FiO₂) and/or CO₂ (FiCO₂). Inclusion of CO₂ in the inhaled O₂ gas mixture was proposed to reduce O₂-induced blood flow reductions, and previous studies have demonstrated differential T₁ and T₂* responses to breathing carbogen and 100% O₂^{1,2}. However, in these studies the underlying tissue oxygenation and perfusion status were not investigated, and gases were delivered in a fixed order. The objective for the current study was to characterize T₁ and T₂* responses to various combinations of FiO₂ and FiCO₂ in rabbit abdominal tissues. Potential blood flow and partial pressure of O₂ (pO₂) dependence on previous gas challenges were examined by randomizing the gas challenge order in each subject. Following the imaging sessions, pilot measurements of tissue pO₂ and perfusion were conducted to assess temporal dynamics that may contribute to MR relaxation time changes.

Methods: All animal studies were approved by our institutional animal care committee. Quantitative T₁ and T₂* MRI parameters were collected from five New Zealand white rabbits (4.0 - 4.5 kg) in six imaging sessions with various combinations of FiO₂ and FiCO₂. Each rabbit was induced with 5 % isoflurane, and maintained at 1 % for the MR experiments and 2 % for the invasive studies. The following gas challenges were delivered in random in 5 - 8 different steps: room air (21% O₂, balanced N₂), 10% CO₂ (balanced air), 100% O₂, and carbogen (10% CO₂ and 90% O₂). MR imaging was performed on 1.5 T GE scanner (Signa EXCITE TwinSpeed; General Electric Healthcare, Milwaukee, WI, USA) using a transmit/receive quadrature knee coil, with coronal images slices positioned to encompass the kidney, liver, and paraspinal muscle. T₁ quantification was achieved using a variable flip angle approach, with a series of 3D fast spoiled gradient recalled echo scans with the following imaging parameters: TR = 7.2 ms, TE = 3.1 ms, FA = 2, 10 and 21°, FOV = 160 mm, matrix = 256 × 160, slice thickness (SL_{TH}) = 3 mm, number of slices (N_{SL}) = 10, N_{AVG} = 4. T₂* quantification was achieved using a 2D multi-echo gradient-echo sequence with the following parameters: TR = 100 ms, 16 equally spaced TEs = [2.1 - 47.1] ms, FA = 30°, FOV = 160 mm, matrix = 256 × 192, SL_{TH} = 3 mm, N_{SL} = 6, and N_{AVG} = 4. Pixel-wise T₁ and T₂* maps were generated using in-house Matlab (V.7.0, Mathworks Inc., Natick, MA, USA) scripts. T₁ parameter maps were generated using the signal equation for the SPGR steady state magnetization with analytical-based flip angle correction using B₁ field maps acquired separately³. T₂* parameter maps were computed by fitting T₂* signal intensity versus echo time to a monoexponential signal decay function⁴. Regions-of-interest were outlined in the liver, kidney and muscle to extract mean T₁ and T₂* values for each gas challenge.

Invasive tissue measurements were conducted in a separate session with OxyLite and OxyFlo (Oxford Optonics, Oxford, UK) fibre optic probes that were inserted into the three tissue types (liver, kidney, and muscle). The OxyLite uses fluorescence lifetime measurements to quantify absolute tissue pO₂ and OxyFlo uses laser Doppler to provide relative measures of tissue perfusion.

Relative ΔT₁ and ΔT₂* values were computed for each transition between the different gas challenges, and a series of one-tailed one-sample t-tests were performed to determine significant differences from zero (null hypothesis).

Results: Figure 1 provides the ΔT₁ and ΔT₂* values for each of the transitions from air to one of the three inspired gas challenges. All transitions exhibited expected trends, especially in liver and kidney. Key observations include trends toward positive ΔT₂* for air → 100% O₂, negative ΔT₂* on air → 10% CO₂, and equivocal ΔT₂* on air → carbogen. Significance for ΔT₂* transitions existed for air → 10% CO₂ in liver (p < 0.01) and air → 100% O₂ in kidney (p < 0.05). Muscle ΔT₂* were very small and insignificant, and air → carbogen transitions produced inconsistent ΔT₂* in all tissues. ΔT₁ values exhibited variability, with only kidney air → 100% O₂ (p < 0.05) transition exhibiting significance. Figure 2 provides ΔT₁ and ΔT₂* for transitions between the three different gas challenges. Expected trends were also observed: positive ΔT₂* and negative ΔT₁ on 10% CO₂ → 100% O₂/carbogen, and reverse changes for 100% O₂ → carbogen. ΔT₂* significance was achieved in both liver and kidney on all transitions: 10% CO₂ → 100% O₂/carbogen (p < 0.05) and 100% O₂ → carbogen (p < 0.01). For muscle, only the negative ΔT₂* for 100% O₂ → carbogen (p < 0.05) was significant. ΔT₁ did not reach statistical significance for any gas transitions, except muscle transitions from 10% CO₂ → 100% O₂ and 100% O₂ → carbogen (p < 0.05).

Figure 3 provides a representative OxyFlo tissue pO₂ and OxyLite perfusion time series in the liver. The invasive measurements demonstrated consistent trends in tissue perfusion and oxygenation changes but considerable variability in their absolute values and temporal dynamics.

Discussion: Although the principle clinical application for MR-based oxygenation monitoring is in oncology, there is still much to learn from normal tissue changes. In this study, we found that T₂* changes followed predicted patterns: ↑ T₂* on 100% O₂ (due to higher pO₂) and ↓ T₂* on 10% CO₂ (due to higher Hb and blood volume). Similarly, T₁ changes followed predicted trends toward a ↓ T₁ on 100% O₂ (due to higher pO₂ and lower blood volume) and ↑ T₁ on 10% CO₂ (due to greater blood volume). However, T₁ changes were much less predictable, compared with T₂*. The air → carbogen transition generated the most variable T₁ and T₂* transitions, possibly attributed to opposing influences from observed perfusion and pO₂ changes. A previous study in normal human abdominal tissues found results which are inconsistent with the T₁ and T₂* transitions in our study¹. This cited study reported T₁ decreases and non-significant T₂* changes for air → 100% O₂ and T₁ increases and T₂* decreases for air → carbogen¹. This distinction suggests physiological differences between the two studies may exist. For the invasive measures, we found large variability in the temporal dynamics of tissue perfusion and pO₂, which is a consideration for future studies. In conclusion, we found that T₂* provided greater sensitivity to changes in tissue pO₂ and perfusion compared with T₁, and we also found independence of relaxation time changes from order of gas administration. Variability in our study and the literature may be partly attributed to organ-specific pO₂ and perfusion dynamics exhibited in this study.

References: 1. O'Connor JP *et al.*, Magn Reson Med 2009,61:75-83; 2. O'Connor JP *et al.* Magn Reson Med 2007,58:490-496; 3. Cheng HL *et al.*, Magn Reson Med 2006,55:566-574; 4. Beaumont M, *et al.*, J Magn Reson Imaging 2009,30:313-320.

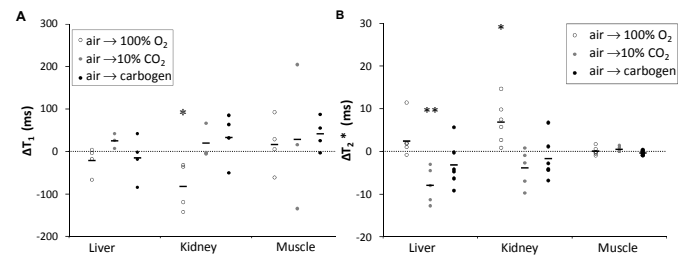


Figure 1. ΔT₁ (a) and ΔT₂* (b) values for each gas transition from air in three tissues. * p < 0.05, ** p < 0.01

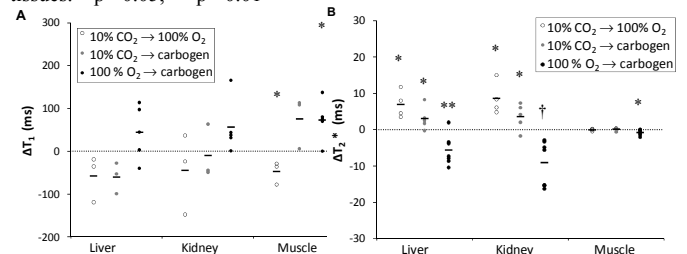


Figure 2. ΔT₁ (a) and ΔT₂* (b) values for transitions between gas challenges in three tissue types. * p < 0.05, ** p < 0.01, † p < 0.005

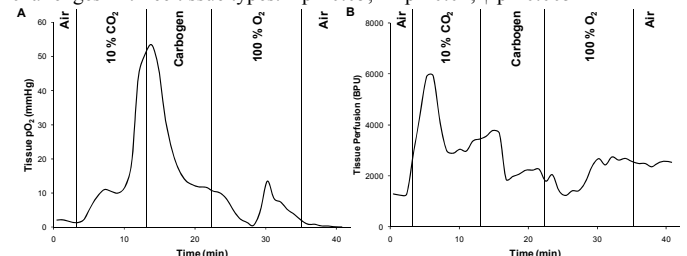


Figure 3. Representative OxyLite pO₂ values (a) and OxyFlo relative perfusion measurements (b) in the rabbit liver.

decreases and non-significant T₂* changes for air → 100% O₂ and T₁ increases and T₂* decreases for air → carbogen¹. This distinction suggests physiological differences between the two studies may exist. For the invasive measures, we found large variability in the temporal dynamics of tissue perfusion and pO₂, which is a consideration for future studies. In conclusion, we found that T₂* provided greater sensitivity to changes in tissue pO₂ and perfusion compared with T₁, and we also found independence of relaxation time changes from order of gas administration. Variability in our study and the literature may be partly attributed to organ-specific pO₂ and perfusion dynamics exhibited in this study.