

# Phase-Adjusted Echo Time (PATE)-Averaging: Application for glutamine resolution at 3.0 Tesla

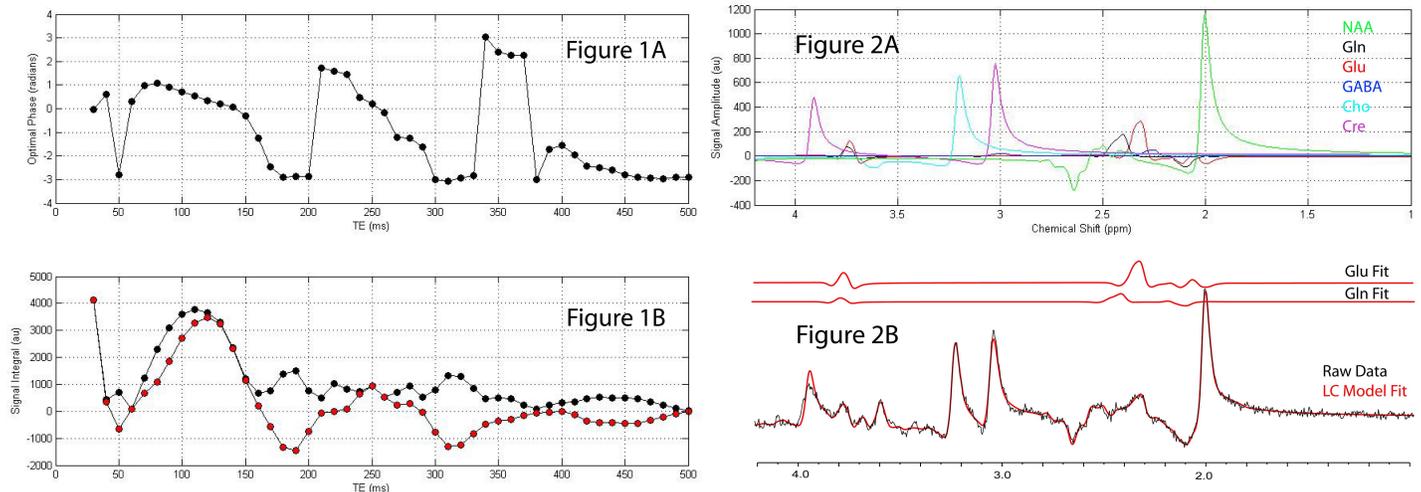
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**Introduction:** Several proton (<sup>1</sup>H) magnetic resonance spectroscopy (MRS) studies have reported abnormal glutamatergic neurotransmission in psychiatric disorders including schizophrenia (1) and acute mania (2). For such studies, the reliable separation of Gln from Glu resonances is of critical importance as impaired glutamatergic neurotransmission may be associated with altered glutamate (Glu) and/or glutamine (Gln) concentration. However, substantial spectral overlap of Gln peaks with Glu and N-acetyl aspartate (NAA) resonances occurs with conventional <sup>1</sup>H MRS performed at clinical static magnetic field strengths ( $B_0 \leq 3.0$  T). Glu resolution can be improved through an approach known as echo-time (TE)-averaging <sup>1</sup>H MRS (3), which involves the averaging of multiple TEs. That method retains the central line of a Glu 2.35 ppm pseudo-singlet at the expense of strongly suppressing the phase-modulated Gln resonances. Here we report a processing method that we have termed phase-adjusted TE (PATE)-averaging for retrieving Gln signals from a TE-averaged <sup>1</sup>H MRS dataset. PATE-averaging applies an optimal zero-order phase factor to each TE step prior to signal averaging. Preliminary simulation and *in vivo* data are presented here to demonstrate the potential utility of the technique.

**Methods:** MATLAB (The MathWorks, Natick, MA) was used to simulate Gln, Glu,  $\gamma$ -amino butyric acid (GABA), choline (Cho), creatine (Cre) and NAA spectra using density matrix theory ( $B_0 = 3.0$  T; TE = 30 - 500 ms;  $\Delta TE = 10$  ms). Simulations assumed ideal spin excitation (-ly) followed by a non-selective double spin echo. Chemical shifts and spin-spin ( $J$ ) coupling constants were taken from precedent literature (4). Spectra were generated using physiologic concentration ratios ([Glu]:[Gln]:[Cho]:[Cre]:[NAA] = 8:4:2:8:10) and a realistic *in vivo* line broadening factor was applied to all spectra. In addition, a T<sub>2</sub>-weighting filter was applied along the TE dimension using previously reported metabolite T<sub>2</sub> relaxation times (5,6). The Gln T<sub>2</sub>-relaxation time filter was assumed to match that of Glu (6). All 48 TE-stepped Gln spectra were subjected to a zero-order phase loop (-179 to 180 degrees) with peak integration over the 2.41 - 2.49 ppm chemical shift region calculated for each phase factor. The optimal TE-specific phase yielded maximum signal area. ***In vivo* Measurements:** Six subjects were scanned twice on consecutive days using a 3.0 Tesla Siemens (Erlangen, Germany) Trio whole body MRI system. TE-stepped <sup>1</sup>H MRS data were acquired from a 22.5 mL voxel positioned bilaterally on the anterior cingulate cortex (TR/TE = 2000/30 - 500 ms;  $\Delta TE = 10$  ms; NEX = 8). *In vivo* data were processed using phasing parameters determined from simulation and fitted using Linear Combination (LC)-Model (7) with a simulated PATE-averaged basis set. The PATE-averaged test-retest performance was compared with standard PRESS data acquired from the same voxel (TR/TE = 2000/30 ms; NEX = 128). The PRESS Cre signal area was used to normalize metabolite peak areas derived from both techniques and test-retest reliability was determined by the group mean coefficient of variation (CV;  $100 \times \text{standard deviation (SD)} / \text{mean}$ ).

**Results and Discussion:** The TE-specific optimal phase for maximizing the Gln peak integral (2.41 - 2.49 ppm) is plotted in Figure 1A. The Gln peak area following application of the optimal phase at each TE is plotted in Figure 1B (black dots) together with the corresponding phase-unadjusted Gln peak area (red dots). Figure 2A shows simulated PATE-averaged data for six metabolites with color-coding used to identify each separate spectrum. That data were reconstructed using the TE range of 50 - 230 ms as the averaging of these phase-adjusted TE steps yielded efficient suppression of the overlapping Glu and NAA methylene resonances. A PATE-averaged dataset acquired *in vivo* is presented in Figure 2B, together with the composite LC Model fit and individual Glu and Gln fits. Gln measurement test-retest reliability was calculated as  $19 \pm 25\%$  and  $8 \pm 7\%$  for PRESS and PATE-averaging, respectively. Glu measurement test-retest reliability was calculated as  $7 \pm 7\%$  and  $9 \pm 6\%$  for PRESS and PATE-averaging, respectively. Processing of TE-averaged <sup>1</sup>H MRS datasets using PATE-averaging approaches may help improve Glu quantification while maintaining Glu measurement reliability. Gln resolution is achievable in ~10 minutes using the parameters outlined in the present study (TE = 50 - 230 ms). PATE-averaging and the targeting of other metabolite resonances (e.g. GABA) currently is under investigation, and comparison with existing methods (8) will help put the findings into context.



**References:** [1] Theberge et al. (2002) *Am J Psychiatry*. **159**:1944-6. [2] Ongur et al. (2008) *Biol Psychiatry*. **64**:718-26. [3] Hurd et al. (2004) *Magn Reson Med*. **51**:435-40. [4] Govindaraju et al. (2000) *NMR Biomed*. **13**:129-53. [5] Traber et al. (2004) *J Magn Reson Imaging*. **19**:537-45. [6] Choi et al. (2006) *Magn Reson Med*. **56**:971-77. [7] Provencher et al. (1993) *Magn Reson Med*. **30**:672-79. [8] Schulte et al. (2006) *NMR Biomed*. **19**:255-63.

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