

# Non-contrast-enhanced Flow-independent Peripheral Angiography using a 3D Concentric Cylinders Trajectory

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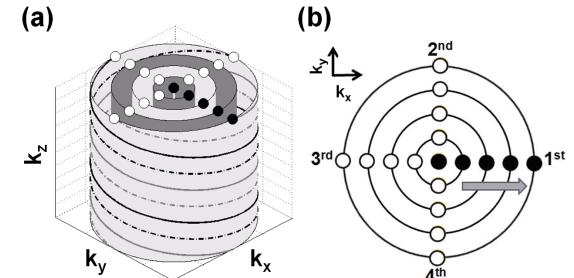
**Introduction:** Non-contrast-enhanced flow-independent angiography (FIA) exploits intrinsic tissue parameters such as T1, T2 and chemical shift to generate vessel contrast [1]. Unlike flow-dependent methods, FIA can generate stable vessel contrast even with slow flow potentially in the diseased leg. Magnetization-prepared 3D SSFP sequences have been of interest for FIA [2,3], but an important issue with these sequences is artery-vein contrast. Recently, an efficient and robust 3D SSFP imaging sequence using a concentric cylinders  $k$ -space trajectory was developed [4]. Concentric cylinders offer the following advantages: a) fewer excitations required than a comparable 3DFT sequence; b) robustness to off-resonance effects; and c) inherent centric view-ordering, crucial for magnetization-prepared imaging [4-6]. In this work, the 3D concentric cylinders trajectory was incorporated into a T2-prepared SSFP FIA sequence for scan-time reduction that can be traded off to improve the artery-vein contrast in the lower extremities.

**Methods:** 3D concentric cylinders: A 3D concentric cylinders trajectory derives from a 2D concentric rings trajectory [7] by adding a constant  $G_z$  gradient during the readout interval. The constant  $G_z$  gradient is a key feature that converts the off-resonance behavior from a blur (typical of non-Cartesian trajectories) into a benign geometric shift in the  $z$ -direction [4-6]. Once the radius ( $k_{x\max}$ ,  $k_{y\max}$ ) and the height ( $k_{z\max}$ ) of the outermost cylinder are chosen based on the desired spatial resolution, the number of uniformly-spaced concentric cylinders ( $N_c$ ) determines the in-plane FOV, while the number of helical interleaves per cylinder ( $N_{\text{intlv}}$ ) and the number of revolutions for each helical interleaf ( $N_{\text{rev}}$ ) determine the  $z$ -direction FOV and the number of slices ( $N_{\text{slice}} = N_{\text{intlv}} \times N_{\text{rev}}$ ) (Fig. 1a). This 3D trajectory is well-suited for T2-prepared SSFP sequence in two aspects. 1) It requires a factor of  $2N_{\text{rev}}$  fewer excitations than a comparable 3DFT acquisition, and 2)  $k$ -space can be easily divided into several centric-ordered segmentations by collecting helical interleaves at the same angular location from each cylinder (Fig. 1b).

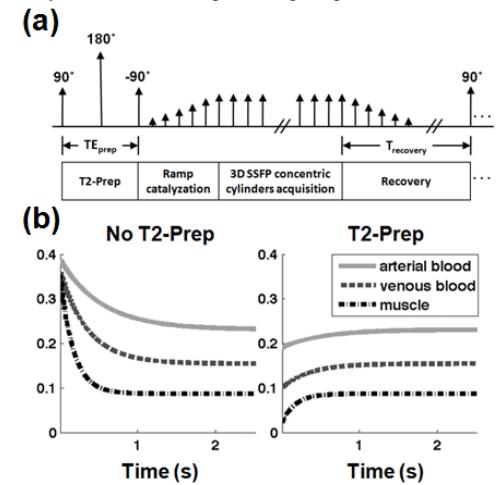
Pulse Sequence: Figure 2a shows a pulse sequence diagram for the T2-prepared 3D SSFP angiography. The T2-preparation augments the artery-vein and artery-muscle contrast inherent in the SSFP sequence (Fig. 2b). This transient contrast is effectively captured by the centric-ordered segmented acquisition using the 3D concentric cylinders trajectory. The factor of  $2N_{\text{rev}}$  fewer excitations can be exploited to increase the number of segments ( $N_{\text{seg}}$ ), thereby shortening the readout interval per segment to better capture the contrast created by T2-preparation. Increasing  $N_{\text{rev}}$  can be also beneficial for purposefully lengthening the TR to improve venous suppression [8].

**Results:** In-vivo experiments were performed on a GE Excite 1.5 T whole-body scanner with a quadrature birdcage coil. Gradients were designed to provide resolution =  $1 \times 1 \times 2$  mm $^3$  and FOV =  $26 \times 26 \times 13$  cm $^3$  (matrix size =  $256 \times 256 \times 64$ ) using  $N_c = 128$ ,  $N_{\text{slice}} = 64$ ,  $N_{\text{intlv}}/N_{\text{rev}} = 32/2$ . Readout bandwidth =  $\pm 125$  kHz was used, which resulted in  $N_{\text{amp}} = 1312$  points per interleaf. For water-fat separation, an extended 2-point Dixon method [9] was used with TE = 3.8/6.0 ms, TR = 10 ms, and flip angle = 60°. Adiabatic BIR-4 pulses were used for T2-preparation [10].  $k$ -space was divided into  $N_{\text{seg}} = 32$  segments with 1 s recovery time between segments, which yielded 1.3 s for each segment. Two phase-cycled datasets were acquired for banding artifact reduction [11]. Total scan time was 6 min. Images were reconstructed by 3D gridding followed by the extended 2-point Dixon method and a maximum-intensity-projection (MIP) with a factor of 2 zero-padding in all three dimensions. Figure 3 shows the coronal MIP images of the calf obtained with no T2-Prep, T2-Prep with TE<sub>prep</sub> = 80 ms (maximizing artery-muscle contrast), and T2-Prep with TE<sub>prep</sub> = 160 ms (maximizing artery-vein contrast). The image with T2-prep of TE<sub>prep</sub> = 80 ms (Fig. 3b) shows a significant suppression of background muscle signal compared to the image without T2-Prep (Fig. 3a). T2-Prep of TE<sub>prep</sub> = 160 ms further suppressed the residual venous signal (white arrow) as well as the muscle signal with only marginal decrease of the arterial signal (Fig. 3c).

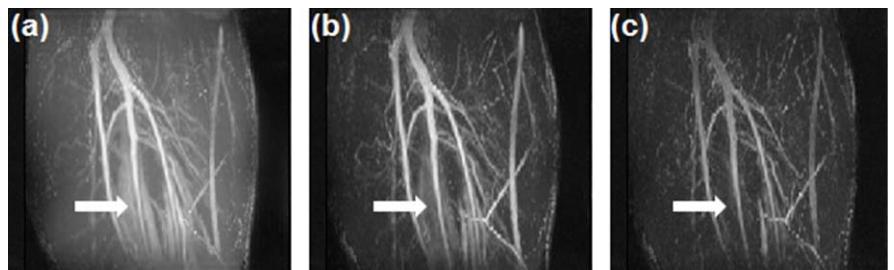
**Discussion:** We demonstrated that the 3D concentric cylinders trajectory is well-suited for the 3D magnetization-prepared FIA method. Compared to other non-Cartesian sequences, implementation is more straightforward and off-resonance correction is not needed. The reduction in number of excitations afforded by the sequence facilitates the use of more imaging segments to better capture the artery-vein contrast created by the T2-Prep. The sequence can be adapted for imaging at 3 T to further improve the contrast by taking advantage of the decreased effective T2 of venous blood. [12]. It can also be combined with other scan-acceleration methods such as variable-density sampling, parallel imaging, and compressed sensing.



**Fig. 1.** (a)  $k$ -space trajectory of 3D concentric cylinders with  $N_c = 5$  cylinders (the radius of the innermost cylinder is 0).  $N_{\text{intlv}} = 4$  helical interleaves with  $N_{\text{rev}} = 2$  revolutions are shown on the side of the outermost cylinder. (b) Centric-ordered scheme ( $N_{\text{seg}} = 4$ ) viewed from  $k_z$  axis. Dots are starting points of interleaves. The collection of black dots represents the 1st segment and the arrow shows the center-out acquisition direction for the 1st segment, which yields a smooth  $k$ -space weighting.



**Fig. 2.** (a) Pulse sequence diagram. (b) Simulated regular SSFP (left) and T2-prepared SSFP (right, TE<sub>prep</sub> = 160 ms) transient signal for arterial blood (T1/T2 = 1000/220 ms), venous blood (1000/100 ms), and muscle (870/47 ms).



**Fig. 3.** Coronal MIP images of the calf: (a) No T2-Prep (b) T2-Prep with TE<sub>prep</sub> = 80 ms (c) T2-Prep with TE<sub>prep</sub> = 160 ms. White arrows show the degree of venous suppressions.

**References:** [1] Wright, MRM 17:126, 1991. [2] Brittain, ISMRM, p.1710, 2003. [3] Stafford, MRM 59:430, 2008. [4] Kwon, ISMRM, p.4973, 2010. [5] Mugler, SMR, p.483, 1995. [6] Ruppert, ISMRM, p.208, 2003. [7] Wu, MRM 59:102, 2008. [8] Dharmakumar, MRM 53:574, 2005. [9] Skinner, MRM 37:628, 1997. [10] Nezafat, MRM 61:1326, 2009. [11] Bangerter, MRM 51:1038, 2004. [12] Brittain, ISMRM, p.1710, 2003.