

Sustained Cerebral Hypoxia Increases Cerebral O₂ Metabolism

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Introduction: Hypoxia decreases arterial oxygenation to the brain and increases resting cerebral blood flow (CBF). Previous studies suggest moderate global hypoxia does not influence resting cerebral oxygen metabolism (CMRO₂) [1-3], yet basal metabolic rate has been shown to increase with sustained hypoxia [4]. In this study, we examined the effects of 2 days and 7 days of sustained global hypoxia on CMRO₂ and Oxygen Extraction Fraction (OEF) in 8 healthy subjects (4M, mean 29 +/- 7 yrs).

Theory: CMRO₂ is the product of CBF, OEF and arterial O₂ content of blood, [O₂]_a:

$$CMRO_2 = CBF \cdot OEF \cdot [O_2]_a$$

Since [O₂]_a is the product of hemoglobin concentration, [Hb], and arterial O₂ saturation, Y_a, this can be rewritten as:

$$CMRO_2 = CBF \cdot \{ (1-Y_v)[Hb] - (1-Y_a)[Hb] \}$$

In this equation, (1-Y_v)[Hb] is the concentration of venous deoxyhemoglobin, dHb. In a different study (submitted as a separate abstract), we determined that dHb is related to venous T₂ by the equation:

$$dHb = k (5.6/T_2 + 0.0273).$$

Thus, CMRO₂ can be determined from venous T₂, scaling constant k, CBF, arterial O₂ saturation (Y_a), and Hb concentration.

Measurements: Venous T₂ was measured using a TRUST (T₂ relaxation under spin tagging) MRI technique with single shot spiral readout (TE=2.8ms, TR=8s, TI=1.2s, 4 echoes, 10 mm slice, 80 mm tag, 4 mins). Baseline CBF was measured using a PICORE QUIPSS2 ASL technique (TE=9.1ms, TR=2.5s, TI₁=700ms, TI₂=1500ms, 6 mm slices, 3.5 mins). O₂ saturation was continuously monitored with a Nonin 8600FO MRI-compatible pulse oximeter calibrated against an arterial blood sample. [Hb] was determined from the arterial blood sample using a IL-682 co-oximeter. Measurements were made during normoxia, and following 2 and 7 days sustained hypoxia at White Mountain Research Station (3,800m altitude, 90 Torr O₂, mean Y_a of 85.4% at 2 days, 87.88% at 7 days). Subjects remained hypoxic until after the MRI was completed. The order of the 2-day and 7-day exposure was random, with an interval of 2-13 months between measurements to ensure subjects were fully de-acclimatized. CBF measurements were corrected for the effect of O₂ desaturation on T₁ of blood. Differences in T₂ measured with TRUST and conventional spin echo T₂ were corrected from a separate phantom calibration measurement. Scaling constant k was adjusted during normoxia such that OEF=0.4 and CMRO₂=1.6 mmol/g/min. Data were analyzed with repeated measures ANOVA and Fisher PLSD (significant at p < 0.05).

Results: During normoxia, mean resting CBF was 44.4 ml/100ml/min (+/- 11.1), mean venous T₂ in superior sagittal sinus was 43.3 ms (+/- 6.4), mean [Hb] was 9.2 mEq/L (+/- 0.8), and mean SaO₂ was 98.5% (+/- 0.5). Proportionality constant k was determined to be 36.5 for CMRO₂=1.6 mmol/g/min, and OEF=0.4. Following 2 days sustained hypoxia, CBF increased to 53.7 ml/100ml/min (+/- 18.6, p = ns), mean venous T₂ in superior sagittal sinus decreased to 30.9 ms (+/- 2.3, p < 0.001), and mean SaO₂ was 85.4% (+/- 3.8, p < 0.01). CMRO₂ increased by 59% to 2.5 mmol/g/min (+/- 0.9, p < 0.01), and OEF by 23% to 0.5 (+/- 0.1, p = ns). Following 7 days sustained hypoxia, CBF was 53.3 ml/100ml.min (+/- 27.4, p=ns), mean venous T₂ was 32.5 ms (+/- 5.3, p < 0.001), and mean SaO₂ 87.9% (+/- 2.4, p < 0.001). CMRO₂ was increased 36% relative to normoxia to 2.2 mmol/g/min (+/- 0.8, p < 0.05), and OEF increased 18% to 0.5 (+/- 0.1, p = ns) relative to normoxia.

Conclusions: The combination of arterial O₂ saturation and [Hb] with two MRI measurements (venous T₂ and baseline CBF) provides a valuable method for determining CMRO₂, and extending the utility of TRUST MRI to hypoxic conditions. Our results indicate that there is a 59% increase in CMRO₂ after 2 days hypoxia, falling to 36% after 7 days. This trend is similar to increases in basal metabolic rate seen during sustained hypoxia.

References: [1] Nioka et al. 1990. J Appl Physiol 68:2527-35. [2] Shimojyo et al. 1968. Neurology 18: 127-33. [3] Moller et al. 2002. J Cereb Blood Flow Metab 22: 118-26. [4] Butterfield et al. 1992. J Appl Physiol 72:1741-1748. **Support:** NIH R01 NS053934

