## Investigating $\Delta R_1$ and $\Delta R_2^*$ as Biomarkers of Tumour Oxygenation

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Introduction: Tumour hypoxia is associated with aggressive, malignant phenotypes and is a major determinant of treatment response and patient outcome. A range of imaging methods are being investigated that may provide a means of assessing tumour hypoxia *in vivo* [1]. Molecular oxygen is paramagnetic, and therefore in solution shortens the MR longitudinal relaxation time  $T_1$  of surrounding water protons [2]. *In vivo*, the level of molecular oxygen dissolved in blood plasma has been shown to cause tissue  $T_1$  to change proportionally to the variation in  $O_2$  concentration [3]. Controlled perturbation of blood plasma  $O_2$  through breathing a hyperoxic gas mixture such as carbogen (95%  $O_2$ , 5%  $CO_2$ ) results in a reduction in  $T_1$  ( $\Delta T_1$ ), providing information about changes in tissue oxygenation status.  $\Delta T_1$  has therefore been proposed as a novel imaging biomarker of tissue and tumour oxygenation [3, 4]. This approach is distinct to intrinsic susceptibility MRI, which relies on the dependence of the tissue transverse relaxation rate  $R_2^*$  on the ratio of oxy- to deoxyhaemoglobin in blood. Baseline  $R_2^*$  and carbogen-induced  $\Delta R_2^*$  are also being investigated for the provision of imaging biomarkers of tumour hypoxia [5, 6]. Given the dependence of both  $T_1$  and  $R_2^*$  on blood oxygenation, we hypothesised that parallel use of the two biomarkers may provide a more informative index of tumour oxygenation than either does individually. To this end, changes

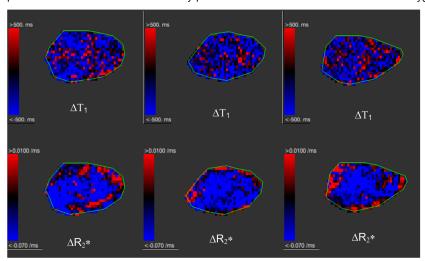


Figure 1. Representative maps of  $\Delta T_1$  and  $\Delta R_2^*$  from one GH3 prolactinoma

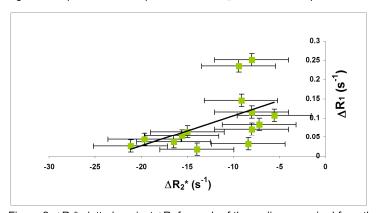


Figure 2.  $\Delta R_2^*$  plotted against  $\Delta R_1$  for each of three slices acquired from the six GH3 prolactinomas.

Baseline R <sub>2</sub> * (s <sup>-1</sup> )	$\Delta R_2^* (s^{-1})$	Baseline R <sub>1</sub> (s <sup>-1</sup> )	$\Delta R_1 (s^{-1})$
118.6 (±4)	-20.2 (±4)	0.61 (±0.02)	0.083 (±0.02)

in oxygenation in GH3 prolactinomas during carbogen breathing was investigated using  $\Delta R_1$  (1/T<sub>1</sub>) and  $\Delta R_2^*$ . Methods: GH3 prolactinomas were propagated by subcutaneous injection of 2.5 × 10<sup>6</sup> cells into the flanks of six female NCr nude mice. The tumours were imaged at a diameter of approximately 1cm. All images were acquired on a 7T horizontal bore Bruker system using a 3cm coil. The mice were anaesthetised and restrained using dental paste in order to limit motion artefacts [8]. A nosepiece was positioned for delivery of air or carbogen. TurboRARE images were acquired for tumour delineation, followed by two sets of baseline multi gradient echo (MGE) images (TR=200msec, TE=6-28ms, 4ms echo spacing, 8 averages, 2min 30s AQ), and one set of inversion recovery TrueFISP images (TE=1.2ms, TR=2.4ms, scan TR=10s,  $\alpha$ =60°, 20 averages, 8 min AQ) from 3 contiguous 1mm slices acquired from a 3x3cm FOV and 128x128 matrix whilst the host breathed air. The gas supply was then switched to carbogen, and following a two minute transition time, further identical MGE and TrueFISP image sets acquired. Data Analysis: Data were fitted using a Bayesian maximum a posteriori approach. This approach takes into account the data's Rician noise distribution, which was used in the calculation of a log-likelihood function which incorporated the Rice probability density function [9]. Tumour T<sub>1</sub> and R<sub>2</sub>\* values were estimated on a pixel-by-pixel basis. The MGE signal magnitude was modelled as a single exponential decay, enabling estimates of ΔR<sub>2</sub>\* uncertainty  $(\sigma_{\Lambda R2^*})$  to be defined and the probability that a given  $\Delta R_2$ estimate was significantly greater than or less than zero.

Results and Discussion: Representative  $\Delta T_1$  and  $\Delta R_2^*$  maps from one GH3 tumour are shown in Figure 1. The  $\Delta T_1$  and  $\Delta R_2^*$  response to carbogen were independently spatially heterogeneous. Carbogen breathing significantly increased  $R_1$  (p=0.02) and reduced  $R_2^*$  (p<0.05) in all six tumours (see Table). The positive median  $\Delta R_1$  is consistent with an increase in dissolved plasma  $O_2$  concentration. The reduction in  $R_2^*$  indicates increased blood oxyhaemoglobin concentration.

A weak yet statistically significant correlation (r=0.54,

p<0.05) was determined between  $\Delta R_1$  and  $\Delta R_2^*$  (Figure 2). This correlation suggests that tumours exhibiting a relatively smaller reduction in  $R_2^*$  exhibited a greater increase in  $R_1.$  This suggests an oxygenated tumour region with saturated haemoglobin, which therefore exhibited greater blood  $O_2$  concentration during carbogen

breathing. Conversely, tumour regions which exhibited relatively large reductions in  $R_2^*$  showed a less pronounced increase in  $R_1$ , consistent with a hypoxic yet erythrocyte perfused tumour region in which the deoxyhaemoglobin binds any dissolved molecular  $O_2$  in the blood.

**Conclusions:** In a controlled preclinical setting, the GH3 tumour model exhibits a significant and positive  $\Delta R_1$  and significantly negative  $\Delta R_2^*$  during carbogen breathing. Large negative  $\Delta R_2^*$  and small  $\Delta R_1$  may indicate hypoxic tumour tissue, whereas small negative  $\Delta R_2^*$  and large  $\Delta R_1$  suggest oxygenated tumour regions. The combined use of  $\Delta R_2^*$  and  $\Delta R_1$  may prove more informative for the assessment of tumour hypoxia.

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References: 1.)Tatum, J.L. et al., Int J Radiat Biol, 2006. 82(10): p. 699-757.2).Mirhej, M. CJC, 1964. 43: p. 1130-1138.3).O'Connor, J.P. et al., Mag Res Med, 2007. 58(3): p. 490-6. 4).O'Connor, J.P. et al., Int J Radiat Oncol Biol Phys, 2009. 75(4): p. 1209-15. 5).McPhail, L. and S. Robinson, Radiology, 2009: p. in press. 6).Alonzi, R., et al.Br J Cancer, 2009. 7).Robinson, S.P., et al. J Magn Reson Imaging, 2003. 17(4): p. 445-54.8).Landuyt, W., et al. J Magn Reson Imaging, 2002. 16(2): p. 224-7. 9).Walker-Samuel, S. in *ISMRM Cancer Workshop*. 2008. Nice.