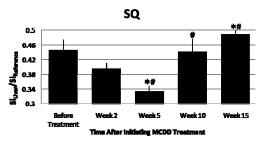
Evaluation of Nonalcoholic Liver Disease using ²³Na MRI and Shift Reagent-aided ²³Na and ³¹P MRS

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Introduction

The ability to noninvasively detect diffuse chronic liver diseases, especially nonalcoholic fatty liver disease (NAFLD), is greatly needed since currently the gold standard for diagnosis is biopsy. One possible imaging modality that takes advantage of an altered transmembrane Na⁺ gradient in diseased tissue is ²³Na MR. The current study uses single quantum (SQ) and triple quantum-filtered (TQF) ²³Na MR techniques to show variations in the observed signal intensity (SI) that correlates with disease progression. To quantify these changes observed with ²³Na MRI, shift reagent (SR)-aided ²³Na and ³¹P MRS techniques were employed to determine the relative intra- and extracellular spaces (rICS and rECS, respectively) and intra- and extracellular Na⁺ concentrations ([Na_i⁺] and [Nae⁺], respectively). To determine whether the TQF SI came from the intracellular space or whether increased extracellular matrix (ECM) proteins contributed to the signal, TQF SR experiments with T₁ and T₂ assessment were conducted before and after SR infusion.



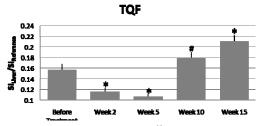


Fig. 2 SQ (top) and TQF (bottom) ²³Na MRI SI before initiating MCDD and 2, 5, 10, and 15 weeks after initiating MCDD (left-to-right).

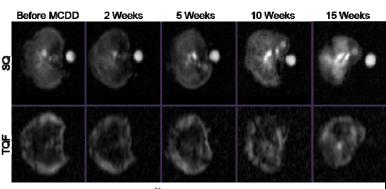


Fig. 1 SQ (top) and TQF (bottom) ²³Na MRI before initiating MCDD and 2, 5, 10, and 15 weeks after initiating MCDD (left-to-right).

Methods

Wistar rats (~250 g) were placed on methionine- and choline-deficient diet (MCDD), after baseline SQ and MQF 23Na MRI. Additional data were collected at 2, 5, and either 10 weeks or 15 weeks after initiating MCDD treatment. MR data were acquired with a Varian 9.4 Tesla horizontal bore system. 3D SQ transaxial ²³Na MRI were obtained with a home-built loop-gap resonator tuned to 106 MHz. The SQ 23Na MRI were collected using a gradient-echo (GE) imaging sequence and following imaging parameters: TR = 50 ms, TE = 4.5 ms and 10 min total imaging time. TQF ²³Na MRI employed the same parameters as used for SQ ²³Na MRI except TR = 100 ms and 50 min total imaging time. Rats were surgically prepared for infusion of TmDOTP⁵ through the external jugular vein. A 2 cm diameter surface coil tunable to 106 MHz for ²³Na and 163 MHz ³¹P spectra was placed over the exposed liver with proper placement being confirmed by minimal if any observable phosphocreatine peak in the 31P ATP spectra. SQ ²³Na MRS were collected with a simple one-pulse sequence. ²³Na T₁ was measured using a pulse-burst saturation recovery pulse sequence consisting of 10 saturation pulses followed by an incremental delay (16 values ranging from 0.05 to 200 milliseconds), a 90° observed pulse and acquisition with Cyclops phase cycling. 23 Na T_{2f} and T_{2s} were measured using a Hahn SE sequence consisting of a composite 180° pulse. The TE was varied from 0.06 to 40 milliseconds. The instrument dead time of 10 microseconds was included as a part of the TE. The relaxation times were computed by fitting a plot ²³Na resonance area versus TR or TE to a mono-exponential function for T_1 and a bi-exponential function for T_2 . The TQF ²³Na MRS sequence employed a MQ preparation time (τ) and evolution time $(\bar{\delta})$ of 3 μ s in order to maximize the SI [1]. Due to the dependence of the TQF signal on T_{2s} and T_{2f}, a second TQF experiment was performed by varying τ from 0.06 to 40 milliseconds. Histologic samples were fixed in formalin and stained with H&E and trichrome stains.

Results

SQ and TQF ²³Na MRI SI decreased to a minimum at week 5 (0.33±0.01 and 0.11±0.01, respectively) and then peaked at week 15 (0.44±0.04 and 0.18±0.01, respectively) becoming

significant to baseline. These data negatively correlate with the lipid content observed with 1H MRI/S (data not shown). For the SR experiments, an increase in [Na_i†] was observed at week 15 (58.6±3.34) compared to baseline (28.9±8.6). No change was observed between intra- and extracellular T_1 measurements. T_{2s} did not vary between timepopints, however, T_{2f} did increase slightly (~50%) at week 15 compared to baseline. Although the [Na_i†] did increase, the TQF 23 Na MRS data showed that a large component of the TQF signal arose from the extracellular space. Therefore, the increase in TQF 23 Na MRI SI is due to not only increased [Na_i†], but also a large increase in the macromolecules associated with fibrosis development. No variations were observed in pH, [Mg²¹], or β -ATP/P_i between timepoints. Histological data correlated with the data (i.e., increasing fibrosis with time and cirrhosis development at week 15).

Conclusion

SQ and TQF ²³Na techniques are sensitive to cellular and tissue damage caused by diffuse liver diseases, prompting their further development and clinical translation. TQF ²³Na MRI may be more useful in detecting the severity and progression of liver damage than SQ ²³Na MRI since it depends on [Na_i⁺] and the accumulation of ECM macromolecules [2].

References: [1] Seshan *et al. MRM* 38:821 -27 (1997) [2] Weber *et al.* Crit Rev Toxicol. 33:105-36 (2003)

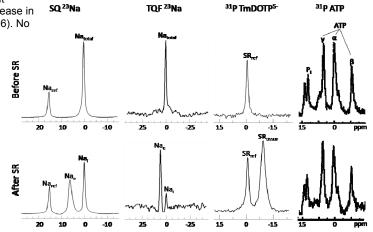


Fig. 3 SQ (left) and TQF (left-middle) ²³Na MRS and TmDOTP⁵⁻ (right-middle) and ATP (right) ³¹P MRS before (top) and after (bottom) infusing the shift reagent, TmDOTP⁵⁻.