

Rapid Quantification of Arterial Input Function and Myocardial T₁ Changes in Mice during Contrast Agent Injection

W. Li^{1,2}, W. Li^{1,2}, C. Flask^{2,3}, M. Griswold^{2,3}, and X. Yu^{1,2}

¹Department of Biomedical Engineering, Case Western Reserve University, Cleveland, OH, United States, ²Case Center for Imaging Research, Case Western Reserve University, Cleveland, OH, United States, ³Department of Radiology, Case Western Reserve University, Cleveland, OH, United States

Introduction

Dynamic contrast enhanced MRI (DCE-MRI) has been used increasingly in cardiovascular MR (CMR) (1). The time course of contrast agent accumulation in myocardium and blood is required for quantitative analysis. A common practice is to obtain contrast agent concentration from the signal intensity of T₁-weighted gradient-echo images. However, this approach is subject to multiple artifacts (1). More accurate quantification can be obtained from T₁ measurements because of the linear relationship between R₁ and the contrast agent concentration (2). Previously, we developed an ECG-triggered saturation recovery Look-Locker (SRL) method that allowed T₁ mapping of mouse myocardium in 3 min (3). However, the longer T₁ value of the blood may require a longer imaging time. This leads to inadequate temporal resolution to track the dynamics of contrast agent in rapid circulating blood in mice. In the current study, we aimed to develop and evaluate a modified SRL (MSRLL) method with improved temporal resolution for T₁ quantification of both blood and myocardium in DCE-MRI experiments.

Methods

MRI Protocol The MRI study was performed on a horizontal 7T Bruker scanner (Bruker Biospin Co. Billerica, MA). The ECG-trigger SRL pulse sequence was described previously (3). Briefly, a 90° RF pulse was applied at the beginning of each phase encoding step, followed by a series of ECG-triggered FLASH acquisitions to track the longitudinal recovery process. In the current study, three nonselective 90° RF pulses were applied using a 18 cm long volume coil to achieve whole body saturation. To improve the temporal resolution of T₁ mapping, 32 phase encoding lines in the low spatial frequency range were acquired during the injection and washout periods. The images were expanded to a larger matrix size (128x64) offline with the high spatial frequency lines from the images acquired at baseline (Fig. 1). Such acquisition scheme effectively halved the imaging time.

Phantom Study A water-filled 2cm diameter and 8 cm long phantom was imaged using SRL and MSRLL methods, respectively. The following imaging parameters were used: TE, 1.7 ms; TR, 3.5 s; number of FLASH images, 16; flip angle, 10°; slice thickness, 1.5 mm; number of averages, 1; FOV, 3x3 cm². The equilibrium magnetization was measured at baseline with a TR of 5 s and a matrix size of 128x64.

Animal Preparation and MEMRI Protocol Four month old C57BL/6J mice (n = 5) were used. Animals were prepared as previously described (3). Heart rate was maintained at around 480 bpm with 0.8–2% isoflurane. To demonstrate the utility of MSRLL in DCE-MRI experiment, a 126 mM MnCl₂ solution was injected via tail vein at a dose of 14 nmol/min/g (body weight) for 30 min, followed by a 30 min washout period. ECG and respiratory signal was monitored by a physiological monitoring system (SA Instruments, Billerica, MA). 11–14 FLASH images were acquired at end-diastole, rendering a TR of ~3.5 s. Other imaging parameters were the same as for phantom study. Baseline T₁ was measured with both large (128x64) and undersampled (128x32) matrix sizes. During the injection and washout periods, images were acquired with undersampling in phase encoding direction for high temporal resolution (<2 min).

Image Analysis Composite images with a matrix size of 128x64 were generated by expanding the undersampled images with the high spatial frequency lines from the baseline image (Fig. 1). All images were further zero filled to 128 x 128. Parametric T₁ maps were generated using an in-house developed Matlab software described previously (3).

Statistic Analysis All results were expressed as mean ± SD. Paired student's *t*-test analysis were performed to compare T₁ values estimated using MSRLL method to those using SRL for both phantom and *in vivo* studies. P<0.05 was considered statistical significant.

Results

Each T₁ map was completed within 2 min using MSRLL method. The composite images exhibited comparable SNR and edge information as the images acquired with twice phase encoding lines (Fig. 2). The accuracy of MSRLL method was demonstrated by its strong agreement in T₁ values measured by SRL method (Fig. 2h). Further, the estimated T₁ values of both myocardium and blood were consistent with the literature (4). Fig. 3 shows that during the course of MnCl₂ injection, R₁ values of blood and myocardium increased from 0.57±0.06 s⁻¹ and 0.75±0.03 s⁻¹ to 1.39±0.28 s⁻¹ and 2.04±0.31 s⁻¹, respectively. During the washout period, blood R₁ decreased rapidly to the baseline level, while myocardial R₁ remained unchanged, suggesting Mn²⁺ retention in the myocardium.

Conclusion

The MSRLL method allows the measurement of both myocardium and blood T₁ in mouse within 2 min. The accuracy was validated in both phantom and *in vivo* studies. The practical utility of this method was demonstrated in a dynamic manganese-enhanced MRI study. In summary, this study suggests that MSRLL may provide a rapid and robust method for simultaneous quantification of the arterial input function and myocardial contrast agent accumulation in DCE-MRI studies.

References

1. Edelman RR. Radiology 2004. 2. Bokacheva L et al. MRM 2007. 3. Li W et al. ISMRM Proc. 2009, No.442. 4. Lu et al. MRM 2004;

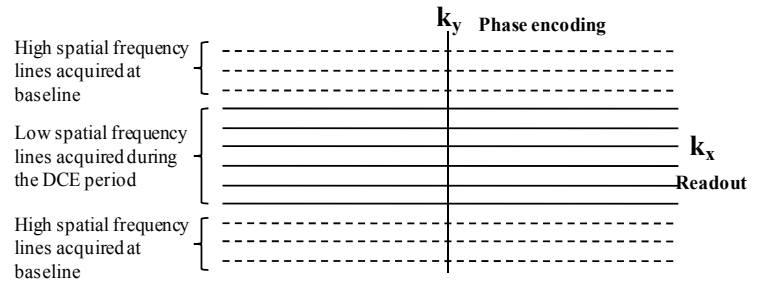


Figure 1. The expansion scheme of undersampled images acquired during the DCE period.

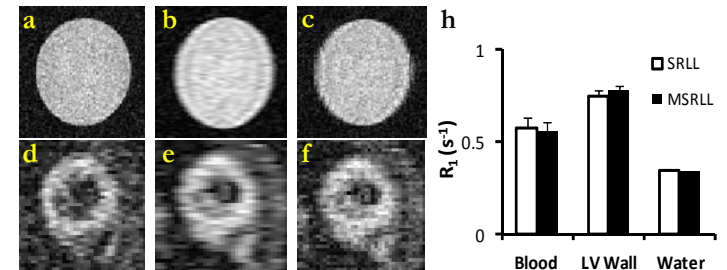


Figure 2. Water phantom and *in vivo* mouse heart. (a) & (d) Images acquired with large matrix size. (b) & (e) Images acquired with small matrix size. (c) & (f) Composite images of (b) & (e) expanded with high spatial frequency lines from (a) & (d), respectively. (h) T₁ values estimated from SRL and MSRLL methods.

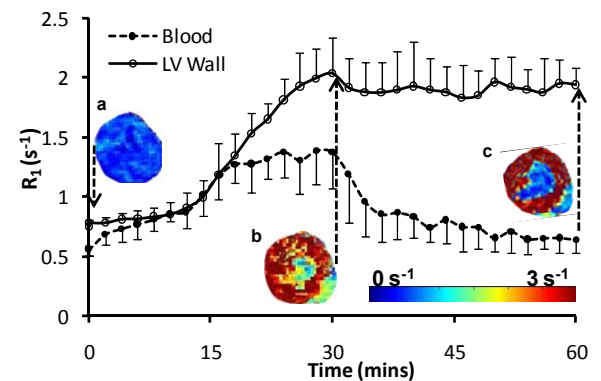


Figure 3. Dynamics of LV and blood R₁ in response to Mn²⁺ injection. (a) – (c) R₁ maps acquired at (a) baseline, (b) end of 30 min Mn²⁺ infusion and (c) end of 30 min washout period.