

Cancer Imaging with Succinate Hyperpolarization

P. Bhattacharya¹, E. Y. Chekmenev¹, S. Wagner¹, H. R. Chan¹, W. H. Perman², A. Epstein³, and B. D. Ross⁴

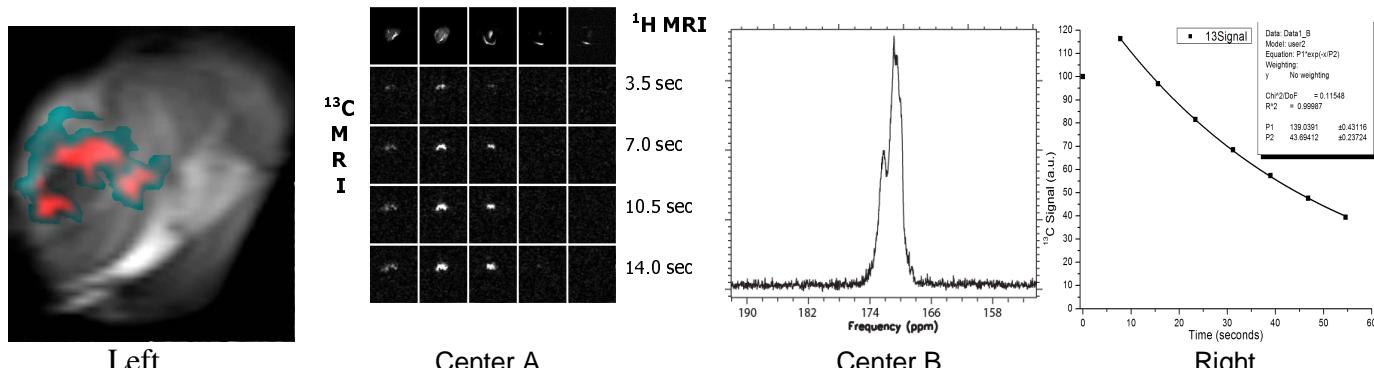
¹Enhanced MR Laboratory, Huntington Medical Research Institutes, Pasadena, CA, United States, ²School of Medicine, St. Louis University, St. Louis, MO, United States, ³Department of Pathology, University of Southern California, Los Angeles, CA, United States, ⁴Enhanced MR Laboratory, Huntington Medical Research Institutes, Pasadena, CA, United States

Background: The Enhanced MR laboratory at HMRI is at the forefront of hyperpolarizing ¹³C sodium succinate [1,2] and its derivatives like ethylsuccinate by PASADENA method of hyperpolarization. Succinate can be now be hyperpolarized at over 20% level of polarization routinely and reproducibly [3,4]. Succinate is biomedically interesting as it can potentially assess the *in vivo* activity of succinate dehydrogenase (SDH), the enzyme that was recently tagged as an oncogene due to its crucial role in cell energetics [5].

Purpose: The goal of research is to apply hyperpolarized succinate for imaging cancer in different tumor models in mice.

Methods: We utilized PASADENA to hyperpolarize 1-¹³C-succinate-d2 at pH=10 in a home-built polarizer and the hyperpolarized solution is injected via the tail vein of a BALB/c mouse bearing a Colon 26 tumor (N=5). A home made ¹H/¹³C dual resonance two turn loop coil and a ¹H/¹³C dual resonance Litz volume coil (Doty Scientific, Inc., Columbia, SC) are utilized for ¹³C hyperpolarized *in vivo* imaging and spectroscopy. Gradient echo and simple pulse and acquire sequences are used for hyperpolarized imaging and spectroscopy at Bruker Avance 4.7T animal scanner at HMRI.

Results: We have demonstrated that succinate is delivered to the tumor (Colon-26) of a mouse by tail vein injection (Fig. left). A ¹³C image was acquired with a large flip angle (40°) every 3.5 seconds with polarization re-supplied by inflowing blood. The hyperpolarized succinate signal from the inflowing blood allowed for imaging up to 1 minute after injection. The 2-D ¹³C image acquired was five 3.2 mm slices, 32 x 32 matrix, with an in-plane resolution of 1.9 mm. Proton images of the tumor verified that hyperpolarized succinate was localized primarily in the tumor space (Fig. center A). ¹³C MR spectroscopy of the tumor localized by the coil reveal a broad peak at hyperpolarized succinate resonance and the presence of metabolites is not confirmed in this mouse model where the succinate oncogene is absent (Fig. center B). *In vivo* ¹³C T₁ of succinate. Note that the first data point is excluded from T₁ simulation due to ongoing saturation of tumor by the succinate arriving to the tumor area. ¹³C *in vivo* T₁*=43.7±0.3 s. (Fig. right)



Conclusions: Succinate is taken up by the Colon-26 mice model of cancer. No metabolites are detected in this model where the succinate oncogene is absent. *Ex vivo* solid state ¹³C NMR spectra of the tumor reveal resonances from Glutamine and Citrate. Hyperpolarized succinate can be used to test permeability and uptake in a tumor as well as tissue heterogeneity. Experiments are underway with various other models of rodent cancer (e.g. 4T1 breast carcinoma). *In vivo* trials using hyperpolarized succinate derivatives like ethylsuccinate are also in progress in our laboratory.

Acknowledgements: We thank the following for funding: Tobacco Related Disease Research Program 16KT-0044, NIH/NCI R01 CA 122513, 1R21 CA118509, 1K99CA134749-01, Rudi Schulte Research Institute, James G. Boswell Fellowship, American Heart Association, American Brain Tumor Association and Prevent Cancer Foundation. Prof. Daniel P. Weitekamp & Valerie A. Norton at Caltech with help with the polarization transfer sequences.

References:

1. Bhattacharya P., et al. (2007) J. Magn. Reson. 186, 108-113.
2. Chekmenev, E.Y., et al. (2008) J. Am. Chem. Soc. 130: 4212-4213.
3. Hövener J.B., et al. (2009). Magn. Reson. Mater. Phys. in press.
4. Hövener J.B., et al. (2009). Magn. Reson. Mater. Phys. in press.
5. Rustin P., et al. (2002) Eur. J. Hum. Genet. 10 289-291.