

Multi-Parametric Classification of fMRI-Activated Voxels Using Venous Vessel-Size, BOLD Latency and Susceptibility-Weighted Imaging

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Introduction

In order to increase the spatial specificity of *functional magnetic resonance imaging* (fMRI) based on the *blood oxygenation level dependent* (BOLD) effect by excluding macrovascular contributions, several approaches have been presented to identify large veins [1-10]. In the present study, some of the techniques mentioned above have been combined into a single set of experiments by the *multi-gradient-echo single-shot-sampling of spin-echo refocusing* (MESSER) sequence [10, 11] which is a single-shot parallel-imaging variant of *gradient-echo sampling of FID and echo* (GESFIDE) [12]. MESSER allows fast mapping of several parameters which are supposed to indicate the presence of large veins: From the transverse *gradient-echo* (GE) and *spin-echo* (SE) relaxation rates, $\cdot R_2^*$ and $\cdot R_2$, the average venous vessel size, $\cdot r_v$, the change in resonance frequency, $\cdot \Delta\omega$, and the BOLD latency $\cdot t_D$ can be estimated. To compare the results with a well-established external standard, MESSER was combined with high-resolution venograms based on susceptibility-weighted imaging to obtain a measure for the 'veininess', $\cdot V_{SWI}$, of a voxel. The purpose of this approach is twofold: Firstly, it was possible to study different parameters of activated voxels and their interrelationships. Secondly, based on these findings, the effectiveness of these parameters to detect large veins in order to increase specificity of BOLD-based fMRI could be evaluated.

Materials and Methods

Measurements were performed on a Siemens 3T Trio system with the MESSER sequence and an acceleration factor of three; a limited FOV of 154. × 154-mm; 64. × 64 matrix; SE echo time, $\cdot TE = 121$ ms; repetition time, $\cdot TR = 1050$ ms; 110-kHz receiver bandwidth; phase encoding direction from left to right; 7 slices (3-mm thickness). Eight healthy subjects were examined using visual stimulation with a short block duration of 10 s and a total of 15 blocks. fMRI analysis was performed by a linear correlation of the $\cdot -R_2^*$ and $\cdot -R_2$ time course to emulate an GE and SE experiment, respectively. A Fourier transformation of the time course yielded $\cdot t_D$ [13] and $\cdot r_v$ was estimated from the combined change in $\cdot R_2^*$ and $\cdot R_2$ [10, 14-16]. Finally, $\cdot \Delta\omega$ was obtained by linear regression of the change in image phase as a function of $\cdot TE$. To obtain the venograms, a three-dimensional first-order fully flow-compensated spoiled GE sequence ($\cdot TE = 28.8$ ms, $\cdot TR = 50$ ms) with an isotropic resolution of 0.6 mm was used to obtain high-resolution phase images which were high-pass filtered to obtain maps of $\cdot V_{SWI}$. Activated voxels were classified by a $\cdot K$ -means clustering algorithm using $\cdot r_v$, $\cdot t_D$ and $\cdot V_{SWI}$. In addition, the interrelationship between these parameters among each other and with $\cdot \Delta R_2^*$, $\cdot \Delta R_2$ and $\cdot \Delta\omega$ was studied by a rank-correlation analysis (Kendall's $\cdot \tau$).

Results

Fig. 1 displays representative maps of $\cdot r_v$ and $\cdot t_D$ overlaid onto $\cdot V_{SWI}$. Most of the venous structures, i.e. regions with a high $\cdot V_{SWI}$, correspond well with a high $\cdot r_v$ and a late $\cdot t_D$. These qualitative findings could be verified by the cluster analysis which divided the ensemble into a macro- and a microvascular voxel population (Table 1). Besides $\cdot r_v$, $\cdot t_D$ and $\cdot V_{SWI}$, $\cdot \Delta R_2^*$ is large for the cluster 'macro'. Both the microvascular and macrovascular $\cdot \Delta\omega$ have a mean value of almost zero indicating that large veins are not accompanied by detectable non-zero $\cdot \Delta\omega$ in our experiments. In the SE-activated ensemble, the mean $\cdot r_v$ is lower than that of the $\cdot R_2^*$ -activated voxels in the ensemble 'all', but higher than $\cdot r_v$ of the microvascular ensemble. However, unexpectedly, $\cdot V_{SWI}$ is slightly higher than in the GE-activated ensemble and $\cdot t_D$ is relatively high. Thus, it is questionable whether the SE ensemble reflects microvascular effects in general which is commonly assumed. The correlation analysis revealed highly significant mutual correlation of 0.17 between $\cdot V_{SWI}$ and $\cdot r_v$ as well as $\cdot t_D$. In contrast, correlation of these three parameters with $\cdot \Delta\omega$ and $\cdot \Delta R_2$ revealed much lower correlation coefficient below 0.1.

Discussion and Conclusions

The relatively good mutual correlation of venous vessel size, latency and venographic intensity suggests that these parameters can be used equally well to discriminate large veins. In contrast, a consistent change in resonance frequency for the macrovascular population was not observed in our experiments. Finally, it is questionable whether SE-BOLD contrast can be exclusively attributed to microvascular effects.

References

- [1] Lai S, et al. *Magn Reson Med*, 30(1993):387. [2] Lee AT, et al. *Magn Reson Med*, 33(1995):745. [3] Frahm J, et al. *NMR Biomed*, 7(1994):45. [4] Segebarth C, et al. *NeuroReport*, 5(1994):813. [5] Haacke EM, et al. *NMR Biomed*, 7(1994):54. [6] Hlustík P, et al. *NeuroImage*, 7(1998):224. [7] Krings T, et al. *Am J Neuroradiol*, 20(1999):1907. [8] Menon RS, et al. *Magn Reson Med*, 30(1993):380. [9] Barth M, et al. *Proc ISMRM*, 14(2006):893. [10] Jochimsen TH, et al. *NeuroImage*, 40(2008):228. [11] Newbould RD, et al. *Proc ISMRM*, 15(2007):1451. [12] Ma J, et al. *J Magn Reson B*, 111(1996):61. [13] Bandettini PA, et al. *Magn Reson Med*, 30(1993):161. [14] Prinster A, et al. *NeuroImage*, 6(1997):191. [15] Dennie J, et al. *Magn Reson Med*, 40(1998):793. [16] Troprès I, et al. *Magn Reson Med*, 45(2001):397.

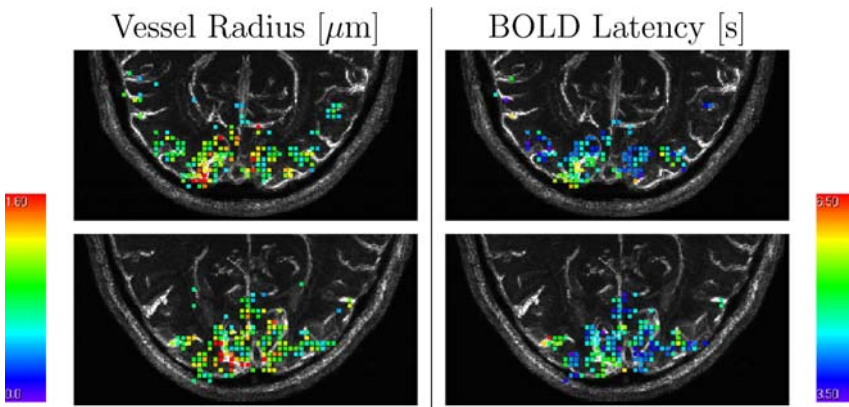


Figure 1: Representative maps of $\cdot r_v$ (left column) and $\cdot t_D$ (right column) in activated voxels overlaid onto venograms. Values of $\cdot r_v$ are color-coded by the decadic logarithm of $\cdot r_v$ in $\cdot \mu\text{m}$ and values of $\cdot t_D$ are color-coded in s. The activation patterns differ since only voxels whose value is in the color-coded range are included in the map.

	$\cdot r_v$ [$\cdot \mu\text{m}$]	$\cdot t_D$ [s]	$\cdot V_{SWI}$	$\cdot \Delta R_2$ [1/s]	$\cdot \Delta R_2^*$ [1/s]	$\cdot \Delta\omega$ [rad Hz]
micro(6602)	5. ± 2.3	4. ± 1.2	0.15. ± 0.046	-0.13. ± 0.29	-0.27. ± 0.56	0.012. ± 0.41
macro(3179)	14. ± 8	4.5. ± 1.3	0.24. ± 0.13	-0.12. ± 0.6	-0.47. ± 1.1	-0.0079. ± 0.35
all(9781)	7.8. ± 6.4	4.2. ± 1.3	0.18. ± 0.093	-0.13. ± 0.42	-0.21. ± 1.3	0.0057. ± 0.39
SE(2063)	6.2. ± 4.2	4.7. ± 2.4	0.2. ± 0.11	-0.24. ± 0.61	-0.55. ± 2.9	-0.018. ± 0.54

Table 1: Average parameters of clusters. The number of voxels are listed in parentheses. The clusters 'micro' and 'macro' contain voxels with mostly micro- or macrovascular BOLD effects, respectively. Moreover, the table also lists the entire ensemble of voxels (all). The last line contains voxels activated only in the $\cdot R_2$ data (SE). Values given are mean and standard deviation.