

Assessing Manganese Efflux Using SEA0400 and Cardiac T₁-Mapping Manganese-Enhanced MRI in a Murine Model

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Introduction

The Na⁺/Ca²⁺ exchanger (NCX) is involved in the regulation of intracellular cardiac Ca²⁺ concentration via either the forward (Ca²⁺ extrusion) or reverse (Ca²⁺ influx) mode. Up to 81% of Ca²⁺ efflux in mice occurs via the NCX (1). During the early phase of reperfusion following myocardial ischemia, the reverse mode of the NCX is activated and intracellular Ca²⁺ overloading takes place. The pathological increase in intracellular Ca²⁺ concentration leads to various cell injuries, which have successfully been reduced using a NCX inhibitor (2). SEA0400 (3), 2-[4-[(2,5-difluorophenyl) methoxy] phenoxy]-5-ethoxyaniline, is such a potent and selective NCX inhibitor, having been shown to provide protection from cardiac ischemia/reperfusion injury, from digitalis induced arrhythmia and myocardial stunning.

Monitoring strategies to assess the changes in Ca²⁺ flux *in vivo* could therefore be diagnostically important but are currently limited. One potential technique to monitor relative myocardial Ca²⁺ efflux is with a surrogate biomarker of Mn²⁺. Producing cardiac T₁-maps at multiple time points post-MnCl₂ infusion allows *in vivo* washout curves to be produced and studied. The first part of the study was designed to examine the effect of varying the infusion dose of MnCl₂ on the washout rate. Secondly, NCX inhibitor SEA0400 was injected into the mice to examine the effect of inhibiting this efflux pathway on the Mn²⁺ efflux.

Methods

Manganese-enhanced MRI (MEMRI) experiments were performed in adult male C57Bl/6 mice (7-13 weeks old, 24.7±2.1g). The animals were anesthetized with a mixture of medical air, oxygen (1:1) and isoflurane and maintained at 1.0-2.7% isoflurane. MnCl₂ was infused into the tail vein at a rate of 0.6 ml/hr. For the dose dependent study MnCl₂ doses of 280, 190, 120 and 65 nmoles/g total body weight (BW) was infused into control mice. To study the effect of inhibiting the NCX a second study was performed. Each group was infused with 190 nmoles/g BW MnCl₂, with 50, 35, 20 or 10 mg/kg SEA0400 injected i.p. one hour post-MnCl₂ infusion. Images were acquired on a 7.0-T, 20-cm horizontal bore BioSpec MRI spectrometer (Bruker Instruments, Billerica, MA) equipped with a micro imaging gradient insert (950 mT/m). Animal setup procedures followed those previously described (4). A 35 mm inner diameter volume coil was used to transmit and receive at ¹H frequency. An ECG-gated, flow-compensated Look-Locker MRI pulse sequence was used to acquire both pre-MnCl₂ and post-MnCl₂ T₁-map short axis heart images, as previously described (5). The T₁-mapping parameters were as follows: matrix = 128 x 128; INTER/TR = 2.5 ms/10 sec; slice thickness = 1.0 mm; FOV = 3.0 x 3.0 cm; NA = 2; inversion time/interval = 9/138 ms; echo images = 50; imaging time ~ 43 minutes. The T₁ value of each pixel was calculated in two steps using a custom written C++ program (5). Regions of Interest (ROI) analysis for the LV Wall was performed using AMIDE (6). The average regional T₁ value and ΔR₁ values were calculated, where ΔR₁ = (post-MnCl₂ infusion R₁) - (pre-MnCl₂ infusion R₁).

Results

Figure 1 shows sample cardiac short axis T₁-maps before MnCl₂ infusion (Figure 1a), 0.2 hours post- (Figure 1b), and 5 hours post- (Figure 1c) MnCl₂ infusion for a control mouse. Temporal changes in ΔR₁ for the MnCl₂ and SEA0400 dose dependent studies are shown as an approximation to the Mn²⁺ efflux rate determination in Figures 2a and 3a along with first order exponential fits. The dose dependence on the exponential half-life from these fits is displayed in Figures 2b and 3b. The half-lives range from 2.0 hrs to 9.5 hrs for the 280 nmoles/g BW and 65 nmoles/g BW groups, respectively. The 35 and 50 mg/kg SEA0400 group efflux half-life values are significantly different (*p* < 0.05) from the control group. The half life for the 50 mg/kg group was extended to 5.6±1.1 hrs.

Conclusions

The MEMRI T₁-mapping protocol provides a quantitative means of measuring temporal changes in the LV Wall relaxation rate, allowing for the inferred study of Mn²⁺ efflux mechanisms non-invasively. There is a Mn²⁺ dose dependence on the ΔR₁ washout rate, with higher doses providing higher ΔR₁ at steady state initially along with a relatively rapid efflux rate. SEA0400 treatment has been shown to attenuate the Mn²⁺ washout rate. Above 20 mg/kg SEA0400 a significant increase in ΔR₁ washout half-life can be observed. Thus far MEMRI T₁-mapping has proved sensitive enough to detect NCX_i modulation fluctuations. Combining information from both studies could allow for individual efflux mechanisms and their relative rates to be determined. The rate of cardiac Mn²⁺ washout could suggest potential Ca²⁺ handling mechanisms, with alterations to the washout rate following inhibition of the NCX with SEA0400 allowing for estimates of the contribution of the different efflux components. Studying Mn²⁺ efflux using this MEMRI T₁-mapping protocol therefore would provide a pre-clinical murine model for examining alterations in Ca²⁺ efflux and to potentially monitor disease progression.

References

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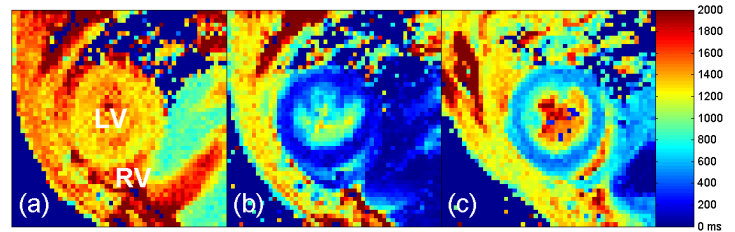
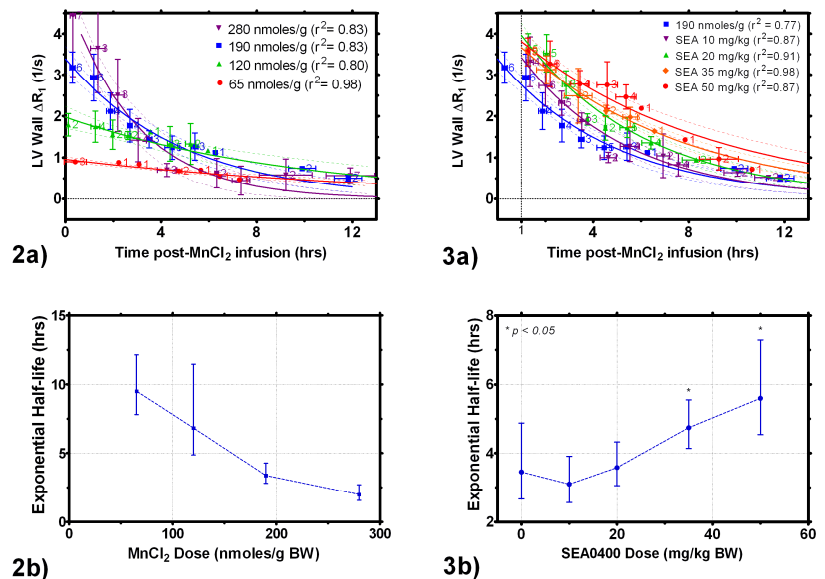


Figure 1. Example short-axis mouse heart T₁-maps (a) pre-MnCl₂ infusion (b) 0.2 hours and (c) 5 hours post-MnCl₂ infusion.



Figures 2 – 3. Temporal ΔR₁ changes post-MnCl₂ infusion for (2a) dose dependent study and (3a) SEA0400 study. First order exponential fits and 95% CIs are shown. 2b) and 3b) show the dose dependence of the exponential half life.