Histone deacetylase inhibition by MRS: Comparison of in vitro and in vivo studies

M. Sankaranarayanapillai¹, J. A. Bankson², Q. Yuan², H. Dafni³, D. D. Webb², A. Pal¹, E. F. Jackson², J. G. Gelovani¹, W. P. Tong¹, and S. M. Ronen³

¹Experimental Diagnostic Imaging, U.T. M.D. Anderson Cancer Center, Houston, Texas, United States, ²Imaging Physics, U.T. M.D. Anderson Cancer Center, Houston, Texas, United States, ³Department of Radiology, University of California at San Francisco, San Francisco, California, United States

Introduction.

Histone deacetylase (HDAC) inhibitors (HDACI) are a novel class of promising antineoplastic agents. Our goal is to develop a magnetic resonance spectroscopy (MRS) based method to directly and noninvasively detect HDAC inhibition. We have previously shown that intracellular levels of the ¹⁹F MRS-detectable Boc-Lys-TFA-OH (BLT) are correlated with HDAC activity and that this method can be used in both cells and *in vivo* tumors (1, 2) to monitor HDAC inhibition. We and others have determined that in cells HDAC inhibition also leads to an increase in phosphocholine (PC) and total choline (tCho) levels, due to heat shock protein 90 (HSP90) inhibition (1,3) by HDACIs. However, we show here that surprisingly, this observation does not translate to our xenograft model.

Materials & Methods.

In vivo ¹H and ¹⁹F MRS on PC3 human prostate cancer xenografts was performed and analyzed as described (2), on control (DMSO treated) (n=6) and 50mg/kg suberoylanilide hydroxamic acid (SAHA) treated (n=6) subcutaneous tumor bearing mice using a 4.7T Biospec (Bruker Biospin, Billerica, MA) prior to treatment (day 0) and on days 2 and 7 of treatment, using a 1.5 cm retunable ¹H/¹⁹F surface coil. ¹⁹F MRS (TR=1s, 45° flip angle, 300 averages) was performed before and after i.p. injection of 100mg/kg BLT (Advanced Chem-Tech, KY USA). The ¹H/¹⁹F surface coil was then replaced with a ³¹P coil and a ³¹P MR spectrum was recorded (TR=2s, 45° flip angle, 900 averages). Western blot analysis of acetylated histone-H3 (Ac-H3), Ac-H4 and HSP90 client proteins (c-Raf and cdk4) were performed on control and treated tumor tissue lysates.

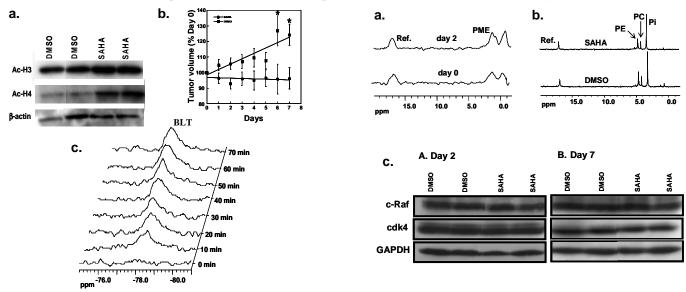


Figure 1. (a) Western blot analysis of acetylated histone H3 and H4 in tumors, (b) Relative change in tumor volume in SAHA (\bullet) and DMSO (\blacksquare) treated mice, (c) Time course of ¹⁹F MR spectra following BLT administration illustrating BLT uptake in the tumor

Figure 2. (a) *In vivo* ³¹P MRS of a SAHA treated tumor, (b) ³¹P MRS of tumor extracts, (c) Western blot analysis of HSP90 client proteins c-Raf and cdk4 in tumor tissue lysates

Results & Discussion.

In PC3 tumor xenografts, HDAC inhibition was confirmed by the accumulation of Ac-H3 and Ac-H4 as seen in the western blot analysis of tumors treated with SAHA for 2 days (Fig.1a). Inhibition of tumor growth was significant by day 6 of SAHA treatment (Fig.1b). The sequential ¹⁹F spectra following SAHA treatment (Fig.1c.) show an increase in tumor BLT levels further confirming HDAC inhibition. Specifically, in SAHA-treated mice, tumor BLT levels were higher by 77% and 132% on days 2 and 7 of treatment compared to pre-treatment levels (p<0.05) while BLT levels remained unchanged in DMSO controls and in normal tissue (data not shown). However, in contrast to cell findings (1) and observations in a different tumor model (3) no significant difference in the PME levels was observed in the ³¹P MR spectra of SAHA treated mice compared to controls (Fig.2a). This observation was also confirmed by *ex-vivo* high resolution MRS of tumor extracts indicating unchanged PE and PC levels (Fig.2b). Interestingly however, and consistent with the ³¹P MRS results, HSP90 client proteins were not depleted following SAHA treatment in this tumor (Fig.2c).

Conclusion.

In summary, our results suggest that ¹⁹F MRS of BLT is a potential non invasive method of monitoring HDAC activity *in vivo* whereas PC is not a reliable marker of HDAC inhibition. However the increase in PC remains an indicator of HSP90 inhibition. This inhibition occurs following treatment with some HDACIs, and was observed in PC3 cells in culture, but not in the PC3 tumor model investigated here. This study thus also highlights the need for validation of cell results *in vivo*.

References. 1. Sankaranarayanapillai, M. et al (2006) *Mol. Can. Ther.* 5: 1325-1334, 2. Sankaranarayanapillai, M. et al (2007) *Proc. ISMRM* 15, p.2990, 3. Y-L.Chung et al (2007) Proc. ISMRM 15, p.2823.