## Optimal spoiling of the transverse magnetization in the Actual Flip-angle Imaging (AFI) sequence for fast B<sub>1</sub> field mapping

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**Introduction.** Fast methods for  $B_1$  measurements recently gained popularity due to the advent of high-field clinical and research scanners operating at 3T and higher field strengths. One of such methods, actual flip-angle imaging (AFI) (1) allows utilization of a short sequence repetition time and provides fast single-scan 3D acquisition of  $B_1$  field maps. The theoretical description of the AFI method (1) is based on the ideal spoiling approximation, and therefore does not take into account the possibility of errors caused by transverse interference. The purposes of this study were to optimize conditions required for sufficient spoiling of the transverse magnetization in the AFI sequence and to describe possible errors in  $B_1$  measurements caused by incomplete spoiling.

**Methods.** *Pulse sequence.* The AFI sequence (Fig. 1) acquires two signals,  $S_1$  and  $S_2$ , which allow calculation of the actual flip angle from their ratio  $r=S_2/S_1$ :  $\alpha=\arccos[(rn-1)/(n-r)]$ . Typically, AFI is used with TR<sub>1</sub>=10-30 ms, the factor  $n=TR_2/TR_1=4-6$ , and the nominal flip angle 50-70° (1). The phase of RF pulses is

progressively incremented with the increment value  $\varphi_0$ , similar to a spoiled gradient echo sequence (2). The spoiling gradients are applied during delays TR<sub>1</sub> and TR<sub>2</sub> with areas proportional to delay durations. The last condition makes uniform any phase evolution, independent of the cause (external gradient or  $B_0$  non-uniformity). In the considered design, spoilers are simultaneously applied along slice-select and readout directions with on-axis areas  $A_{G1}$  and  $A_{G2}$ .

*Experiments.* Measurements were conducted on a 3T Philips Achieva whole-body scanner using a phantom with  $T_1/T_2=784/662$  ms and diffusion coefficient  $D=2.2 \times 10^{-3}$  mm<sup>2</sup>/s (0.2 mM Gd-DTPA solution). Dependences of  $S_1$ ,  $S_2$ , and measured  $\alpha$  on the phase increment  $\varphi_0$  were recorded in a range 0-180° with the step 3° for various TR<sub>1</sub>/TR<sub>2</sub> and  $A_{G1}/A_{G2}$ . *Simulations.* Simulations were performed using a combined isochromat summation and diffusion propagator model (3,4). To describe magnetization evolution during delays TR<sub>1</sub> and TR<sub>2</sub>, a distribution of isochromats after each RF pulse is multiplied by relaxation terms and convolved with the propagator of the Bloch-Torrey equation, which can be written for a pair of isochromats with spatially dependent phases  $\psi_i$  and  $\psi_k$  as follows:

$$_{ik} = (4\pi Dt_D)^{-1/2} (\gamma A_G)^{-1} \exp[-(\psi_j - \psi_k)^2 / (4Dt_D \gamma^2 A_G^2) + i(1/2)(\psi_j + \psi_k) - (1/12)\gamma^2 A_G^2 t_G D)] \quad , \quad [1]$$



**Fig. 1.** Diagram of the AFI sequence with RF pulses and spoiler gradients shown.

where  $t_D$ =TR<sub>1</sub> or TR<sub>2</sub> and  $t_G$  is the time when a gradient is turned on. The propagator [1] describes the Gaussian probability of magnetization exchange between isochromats caused by diffusion assuming the free diffusion case (first term), average phase shift in the gradient field (second term), and irreversible signal loss due to diffusion in the presence of the gradient (third term). RF pulses are described by the standard rotation matrix dependent on  $\alpha$  and  $\phi_0$ . Similar to the traditional isochromat summation technique, the computational procedure is iterated until the total magnetization (vector sum of isochromats) achieves the steady state. The algorithm was implemented in a custom-written C program.

Results. Spoiling behavior of the AFI sequence. The spoiling behavior of AFI is different from that of a regular gradient echo sequence (2). Particularly, the spoiling dependence on the phase increment is symmetric relative to 90°, whereas for the gradient echo sequence such symmetry occurs relative to the 180° point (2). Depending on the gradient areas and TRs, the spoiling patterns of the AFI sequence can be classified into two regimes: weak spoiling and strong spoiling. The examples are shown in Fig. 2. In the weak spoiling regime (Fig. 2a), signals  $S_1$  and  $S_2$  have multi-peak dependence on the phase increment, and flip angle measurements also have a complex dependence on  $\phi_0$ . Although optimal spoiling (i.e. correct  $B_1$  measurement) is possible at multiple  $\varphi_0$ , the dependences are steep and their shape is subjected to rapid changes when the flip angle, gradient areas, or the diffusion coefficient are varied. If sufficiently large spoiling gradients are used, the strong spoiling regime (Fig. 2b) occurs, where the dependence of the measured flip angle on  $\varphi_0$  has a smooth shape with the single minimum at 90°. There are two optimal  $\phi_0$  values producing correct measurements. They are positioned in ranges 20-40° and 140-160° and can be precisely determined from plots similar to that shown in Fig. 2b. Excellent agreement between experiments and simulations (Fig.2) allows using the described theoretical model for sequence optimization.

**Optimization of B\_1 measurement accuracy.** Theoretically, for any gradient

configuration, the exact optimal  $\varphi_0$  depends on the actual flip angle. However, such dependencies are smooth in the strong spoiling regime, and an appropriate  $\varphi_0$  value can be determined for a particular gradient configuration and timing to maximize the accuracy of  $B_1$  measurements within a specified range of  $B_1$  nonuniformities. Fig. 3 shows simulated relative  $B_1$  measurement errors for the three parameter settings of the AFI sequence assuming that the relaxation and diffusion properties of the object are similar to the brain white matter. Note that  $T_1$  and  $T_2$  do not affect the accuracy of measurements in the strong spoiling regime, but the D value is important. For slower diffusion (low D), the spoiling effect is weaker. For in vivo applications, the tissue with the lowest D should be used for sequence optimization (e.g. white matter in the brain). The combinations of gradient areas and TRs shown in Fig. 3 are easy to achieve on most clinical MR scanners permitting 20-25 mT/m gradient strength. With  $A_{G1}/A_{G2}>200/1000 \text{ mT*ms/m}}$  and  $\varphi_0=33-36^\circ$ , highly non-uniform  $B_1$  distributions (~50%  $B_1$  variations) can be mapped with relative errors <2%.

**Discussion and Conclusions.** The strong spoiling regime is preferable for  $B_1$  measurements using the AFI method. Accurate  $B_1$  measurements require the use of sufficiently high spoiling gradients and an optimal value of the RF phase increment. A reduced accuracy may limit AFI applications with ultra-short TR<sub>1</sub><10 ms, since the available hardware may not allow sufficient gradient strength to reach the strong spoiling regime. For TR<sub>1</sub>=15-25 ms, optimal spoiling conditions are easily achievable on most commercial MR scanners.

References: (1) Yarnykh VL. MRM 2007;57:192. (2) Zur Y, et al. MRM 1991; 21:251. (3) Gudbjartsson H, Patz S. MRM 1995;34:567. (4) Kiselev VG. JMR 2003; 164:205.



**Fig. 2.** Spoiling behavior of the AFI sequence in the weak (**a**:  $TR_1/TR_2=10/50$  ms,  $A_{G1}/A_{G2}=16.7/82.6$  mT\*ms/m) and strong (**b**:  $TR_1/TR_2=15/75$  ms,  $A_{G1}/A_{G2}=163/827$  mT\*ms/m) spoiling regimes. The independently measured actual  $\alpha=58.8^{\circ}$ . **Top:** Experimental signal measurements (points) are superimposed with simulations (lines). **Bottom:** Experimentally determined flip angles (black lines) are superimposed with simulations (red lines).



