

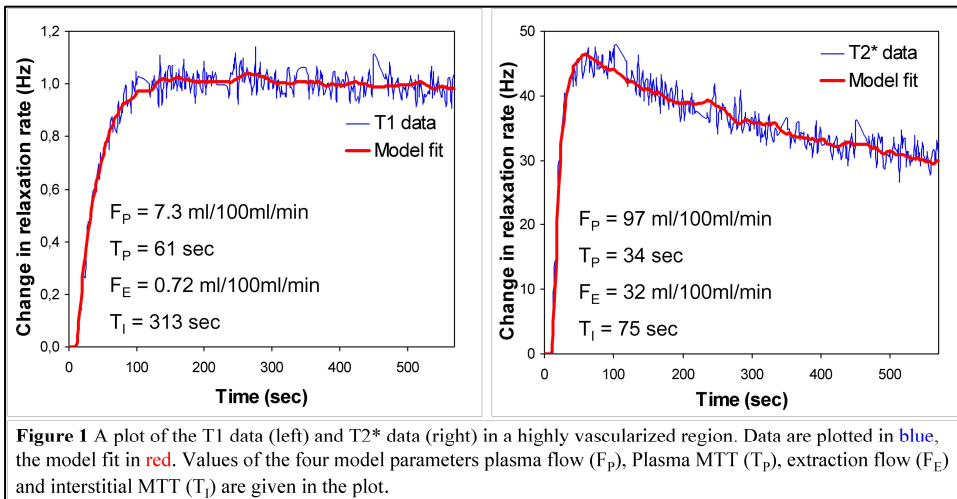
# Tracer kinetic analysis of a simultaneous T1- and T2\* measurement in a tumor model

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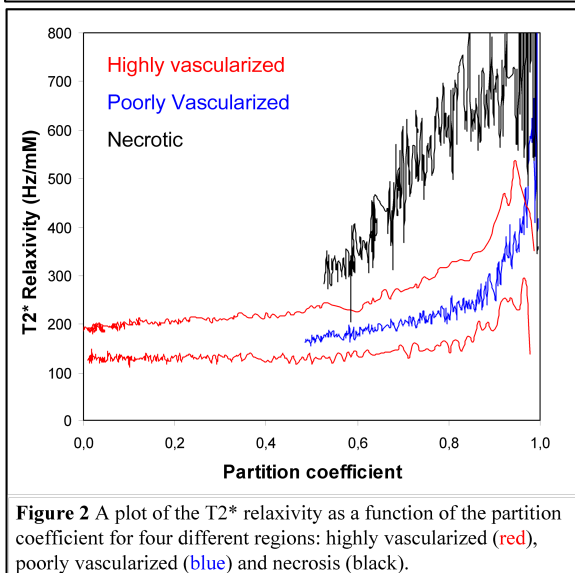
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**INTRODUCTION** Two measurement approaches exist for the quantification of tissue perfusion and permeability with a bolus-tracking experiment: T1-weighted dynamic contrast enhanced MRI (DCE-MRI), and T2\*-weighted dynamic susceptibility contrast MRI (DSC-MRI). The latter has the advantage of strong signal changes even when the tracer is compartmentalized in a small fraction of the tissue volume. On the other hand, DSC-MRI does not allow for absolute quantification since the relaxivity of the contrast agent differs strongly between tissue and arterial blood [1]. Moreover, when the tracer extravasates out of the vasculature the loss of microscopic gradients may lead to a gradual reduction in T2\* relaxivity during the acquisition. The aim of this study is to measure the magnitude of the error introduced by these effects using a sequence that allows for simultaneous measurement of T1 and T2\* at high temporal resolution [2].

**MATERIALS AND METHODS** The measurement sequence is described in detail in [2]. It combines a saturation-recovery Turbo-Flash sequence with a multi-gradient-echo sequence with a temporal resolution of 1.8s. Data were obtained from three mice, measured during the passage of a bolus of 0.6 mmol/kg Gd-DOTA (Dotarem, Guerbet, France) injected in 10s. For each mouse, signal-time curves were measured in the heart to determine the arterial input function, and in three different regions within subcutaneously implanted fragments of a human colorectal cancer: in the necrosis, and in a highly- and poorly vascularized part of the tumor. Data were processed in IDL 6.4 (ITT Visual Information Solutions, Boulder, CO). Relaxation rates R1 and R2\* were quantified for each dynamic using the signal analysis presented in [2]. R1 and R2\* time curves were fitted independently to a full two-compartment model [3], which produces four independent parameters: the plasma flow  $F_p$ , the plasma mean transit time  $T_p$ , the extraction flow  $F_E$  across the capillary wall, and the mean transit time of the interstitium  $T_i$ . These parameters can be combined using the central volume theorem to determine the volumes  $V_p$  and  $V_i$  of plasma and interstitium, respectively. To verify the dependence of the T2\* relaxivity on the distribution of the tracer within the tissue, the fitted model parameters for the T1 data were used to determine the tracer concentrations  $C_p(t)$  and  $C_i(t)$  in the plasma and interstitium, respectively. From these time curves, a partition coefficient  $\lambda = (C_p - C_i)/(C_p + C_i)$  was determined as a function of time. With this definition,  $\lambda = 1$  if the tracer is purely intravascular,  $\lambda = -1$  if the tracer is purely extravascular, and  $\lambda = 0$  when the tracer is equally distributed over both compartments. The T2\* relaxivity was measured as a function of time by  $k2^* = \Delta R2^*/(V_p C_p + V_i C_i)$ , and plotted as a function of the partition coefficient  $\lambda$ .



**RESULTS** An example of the data and the model fits to R1 and R2\* data is given in figure 1. The R2\* curves typically decrease more rapidly after the initial vascular phase, but the model fits are equally accurate for both types of curves. The flow- and volume parameters calculated from the R2\* curves are significantly higher than those of the R1 curves: on the average,  $F_p$  values are higher by a factor 18,  $V_p$  by a factor 9.5,  $F_E$  by a factor 35 and  $V_i$  by a factor 13. The mean transit times are lower when measured from R2\* curves: a factor 0.5 for  $T_p$  and a factor 0.3 for  $T_i$ . With one exception, the total volume of distribution  $V_p + V_i$  calculated from R2\* was unphysical (mean 160 ml/100ml) but in all cases it was within acceptable ranges for R1 (mean 15 ml/100ml). Plasma mean transit time values are overestimated with both methods, presumably due to large bolus dispersion between the arterial site and the tissue. The T2\* relaxivity is plotted in figure 2 for four examples. The partition coefficient never



reached negative values, and has a minimum near  $\lambda = 0.5$  for tissue types with slower extravasation. In all cases the function  $k2^*(\lambda)$  was increasing on the interval  $[0,1]$ . The equilibrium relaxivity  $k2^*(0)$  ranged between 100-200 Hz/mM, and the relaxivity  $k2^*(1)$  of the intravascular tracer ranged from 300-700 Hz/mM. Figure 2 also shows that the regions differ in the rate and the manner in which the relaxivity decreases to its equilibrium value.

**DISCUSSION/CONCLUSION** The results provide experimental confirmation of two different quantification errors in DSC-MRI. First, the effect of the difference in T2\* relaxivity between pure blood and tissue [1] produces a serious overestimation in all flow- and volume parameters. Second, the T2\* relaxivity changes when the contrast agent redistributes within the tissue due to extravasation. The loss of T2\* effects leads to an apparent wash-out of tracer which cannot be distinguished from actual wash-out by tracer kinetic analysis, leading to underestimated values for the mean transit times. This is independent on T1-related leakage effects [4], which also lead to T2\* signal increase during extravasation, but which are eliminated here by the fact that T2\* values are measured dynamically. As such, these results indicate that tissue permeability cannot be measured from T2\* data alone. On the other hand, they show that the combination of T1- and T2\* data produces additional information in the form of the  $k2^*(\lambda)$  curves. The T2\* relaxivities measured in this tumor model are significantly higher than the value of 40 Hz/mM predicted for normal brain tissue [1], which suggests a strong sensitivity to tissue type. If a theoretical model is developed that provides a deeper understanding in the parameters affecting the behaviour of  $k2^*(\lambda)$ , additional parameters may be derived that allow a more complete characterization of the tissue geometry.

**REFERENCES** [1] Kiselev (2005) *JMRI* 22: 693-696, [2] Heilmann et al. (2007) *MAGMA* DOI 10.1007/s10334-007-0082-2, [3] Brix et al (2004) *MRM* 52: 420-429, [4] Quarles et al (2005) *MRM* 53: 1307-1316