

# Resolving Multiple T<sub>2</sub> Compartments in Cartilage with MRI

D. A. Reiter<sup>1</sup>, P.-C. Lin<sup>1</sup>, K. W. Fishbein<sup>1</sup>, and R. G. Spencer<sup>1</sup>

<sup>1</sup>National Institute on Aging, National Institutes of Health, Baltimore, MD, United States

**Introduction:** T<sub>2</sub> measurements have been used extensively to characterize properties of cartilage due to its sensitivity to macromolecular concentration and structure. T<sub>2</sub>'s are typically estimated by fitting NMR relaxation data to a monoexponential function. An increase in cartilage T<sub>2</sub> after enzymatic degradation has been consistently reported, although attempts to associate changes in average values of T<sub>2</sub> to loss of specific tissue components have been inconclusive [1]. Cartilage is composed of various macromolecular components that constitute compartments of water with differing fractions and mobility. Thus, transverse relaxation decay of cartilage is poorly described by a single exponential [2]. Multiexponential analysis of T<sub>2</sub> relaxation has typically been performed using spectroscopic data with good SNR and closely spaced echoes. In imaging experiments, the ability to achieve these conditions is much more constricted, limiting the ability to accurately resolve multiple T<sub>2</sub> components. It has been demonstrated in several biological tissues (i.e. brain, breast, and muscle), that multiple T<sub>2</sub> components can be resolved at SNR, TE, and number of echoes (N) attainable using imaging methods [3]. Therefore, we sought to identify the acquisition parameters (i.e. SNR, TE, and N) required for resolving multiple T<sub>2</sub> relaxation components in cartilage with typical component T<sub>2</sub> values and associated fractions. We applied this analysis to characterize multicomponent T<sub>2</sub> relaxation in intact and enzymatically degraded cartilage using two different enzymes acting primarily on collagen and proteoglycan (PG) matrix components.

**Materials and Methods: Cartilage Sample Preparation.** Bovine nasal cartilage (BNC) disks (diameter = 8 mm) were excised from the nasal septa of 5-6 month-old calves (Green Village Packing, Green Village, NJ) [4]. Samples were threaded onto a hollow tube and inserted into a four-well sample holder filled with DPBS buffer adjusted to pH 7.5 ± 0.1 for initial T<sub>2</sub> measurements. 1 mg/ml trypsin (Sigma-Aldrich, St. Louis, MO) or 30 units/ml collagenase type II (Worthington Biochemical Corp., Lakewood, NJ) was then added to the buffer, with T<sub>2</sub> measurements repeated after 24 hours of incubation at 37 °C in the degradative enzyme. **MRI T<sub>2</sub> Measurements.** MRI experiments were performed using a 9.4 T Bruker DMX spectrometer (Bruker Instruments, Billerica MA) using a CPMG imaging pulse sequence with: TE/TR = 12.8ms/5s, N = 64, NEX = 2, FOV = 4x1.5cm, 0.5mm slice thickness, and 256x128 matrix size. Spin-echo amplitude at each echo time in the relaxation decay was quantified by averaging the pixel intensities from a region of interest (ROI) in the sample. **T<sub>2</sub> Relaxation Simulation.** Relaxation data were simulated from the following expression:

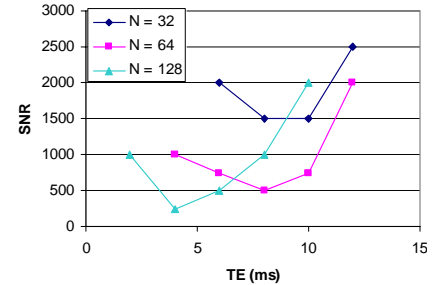
$$y(n \cdot TE) = B + y_0 \sum_{m=1}^M w_m e^{-(n \cdot TE)/T_{2,m}} + \epsilon(0, \sigma),$$

where  $y(n \cdot TE)$  represents the signal amplitude of the  $n^{\text{th}}$  echo,  $B$  represents a baseline offset,  $y_0$  represents the initial signal amplitude,  $w_m$  represents the fractional weight of the  $m^{\text{th}}$  T<sub>2</sub> component, and  $\epsilon(0, \sigma)$  represents the addition of Gaussian random noise with a mean 0 and standard deviation of  $\sigma$ . Simulation input values for T<sub>2</sub>'s and relative fractions ( $w$ ) were selected based on preliminary work from our lab which agree well with previously reported values [5]: T<sub>2,fast</sub> (7.7 ms), T<sub>2,slow</sub> (66 ms),  $w_{\text{fast}}$  (0.75), and  $w_{\text{slow}}$  (0.25). Simulations were performed with SNR ranging from 100 to 2000, TE ranging from 4 to 14 ms, and N ranging from 32 to 128. 100 trials were run for each condition. The multiexponential fit of simulation and experimental data was obtained using a nonnegative least squares (NNLS) method similar to that described in [6].

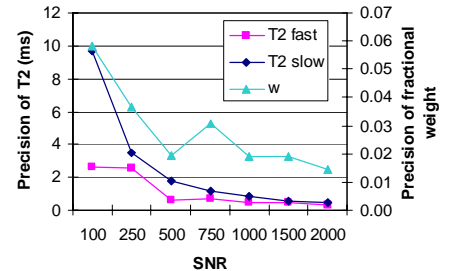
**Results and Discussion:** Figure 1 shows the minimum SNR required for each combination of TE and N to obtain admissible fits in 95% of the runs. Admissible fits were defined as having two components with T<sub>2</sub>'s and fractional weights resolved to within 10% of the input simulation value. These results indicate the optimum TE for a given number of echoes for reliable biexponential T<sub>2</sub> relaxation measurements in cartilage. Figure 2 shows precision of T<sub>2</sub> and  $w$  as it relates to SNR under similar imaging conditions used in the cartilage degradation experiment. Analysis of relaxation data from an imaging experiment is typically performed on an ROI as opposed to pixel by pixel; averaging over a large ROI results in increased SNR. In these experiments, ROI's were chosen such that SNR was ~1500, resulting in T<sub>2</sub> and fractional weight precision within 1 ms and 2% respectively. Both enzymatic treatments resulted in a significant shift in the fractional amounts and T<sub>2</sub> components (Fig. 3), while maintaining ( $w_{\text{fast}} > w_{\text{slow}}$ ). The importance of this work is to detect changes in cartilage matrix components as they relate to degradation. Cartilage matrix consists primarily of collagen and PG. Previous work has demonstrated that water compartments are associated with these macromolecules and that their T<sub>2</sub> decay is faster than that of bulk water [2, 7]. The T<sub>2</sub> components and associated weights of these compartments have been documented as changing with maturation of cartilage and have been associated with an increase in collagen density [5]. Our results revealed two compartments and demonstrated a shift in water fraction from T<sub>2,fast</sub> to T<sub>2,slow</sub> with enzymatic degradation which can be interpreted as a loss of macromolecular-associated water fraction to bulk water. This is the first work to demonstrate the diagnostic potential of multiexponential T<sub>2</sub> analysis of MR images in cartilage.

**Conclusions:** Simulations were performed to define the conditions under which multiple T<sub>2</sub> relaxation components could be reliably performed in cartilage. Experimental results were sensitive to the matrix changes presumed to result from enzymatic degradation. Further work will be directed at achieving measurements of more diagnostic potential by more extensive multiexponential analysis of relaxation time data.

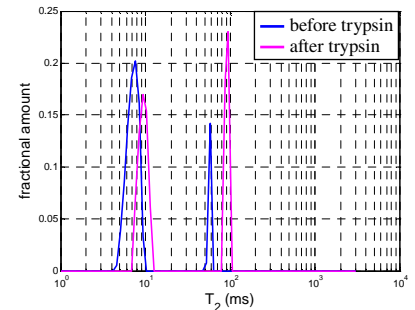
**References:** 1. Menezes, N.M., Magn. Reson. Med. 2004; 2. Lattanzio, P.J., Magn. Reson. Med. 2000; 3. Graham, S.J., Magn. Reson. Med. 1996; 4. Fishbein, K.W., Magn. Reson. Med. 2007; 5. Keinan-Adamsky, K., Magn. Reson. Med. 2006; 6. Whittall, K.P., J. Magn. Reson. 1989; 7. Ghiassi-Nejad, M., Biomaterials. 2000.



**Figure 1.** Minimum SNR required for a given TE and N. Admissible solutions are based on the following: 95% of the trials properly demonstrated two components, and the T<sub>2</sub> components and fractional weights  $w$  were resolved to within 10% of the input value.



**Figure 2.** Precision of T<sub>2</sub> and fractional weight  $w$  determination as a function of degraded cartilage parameters appropriate for degraded cartilage (TE = 12 and N = 64).



**Figure 3.** Representative T<sub>2</sub> histogram before and after trypsin treatment using NNLS.