

Lack of MRI evidence for increased iron in the substantia nigra of PD brains at 7T

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Background:

Parkinson's disease is a common neurodegenerative disorder leading to progressive motor and nonmotor symptoms related to subcortical neuronal cell loss. The underlying pathophysiology is not well understood and a role of iron and free oxygen radicals is debated.¹ There is strong post mortem evidence of increased total iron in the substantia nigra in PD brains with an indication of regional specificity for the pars compacta^{2,3}. There is a long standing interest in developing biomarkers of cerebral iron, and the iron sensitivity of the MRI signal makes it a suitable candidate. A number of studies reported increased iron in the substantia nigra of PD using different MRI measures (T₂, T₂*, T₂' , field dependent relaxivity index, FDRI)⁴⁻⁹. There are however unresolved controversies regarding the respective sensitivities and T₂ abnormalities were least consistent. Using ferritin sensitive FDRI, Bartzokis⁶ showed increased iron only in patients with early-onset PD, with reduced iron in patients with late-onset disease. Findings in the basal ganglia are even less consistent claiming both elevated and reduced iron storage. Ultrahigh-field MRI is expected to provide a more reliable and sensitive assessment of brain iron stores.

Aim: To assess T₂ and T₂* in the substantia nigra and other subcortical nuclei in patients with Parkinson's disease using a gradient echo-spin echo (GESE) sequence at 7T, assuming that T₂ and T₂' are good markers of paramagnetic iron.

Method: 5 patients with Parkinson disease (PD, mean age 57.4, range 34-81), 2 with multisystem atrophy (MSA, mean age 68) and 5 matched controls (mean age 52.2, range 33-66) were included. All subjects gave written informed consent; the study was approved by the local Ethics Committee. For scanning a gradient echo spin echo sequence was used on a Philips Achieva 7T MRI scanner. The sequence and fitting is described in more detail elsewhere¹². In brief, it consists of a series of gradient echoes superimposed on a spin echo (SE) repeated N times for each phase encoding step. The signal obtained at each gradient echo¹⁰ is fitted using the Powell algorithm¹¹ to give T₂ and T₂'. The sequence is insensitive to RF pulse errors since errors in either of the pulse flip angles will cause equal attenuation of all the echoes, leading to a simple reduction in SNR. It is also insensitive to T₁ saturation (weighting) since the time between the refocusing pulse and next 90° pulse is kept constant. The sequence parameters were 1x1x3 mm voxel size, TR=2s, TE=1.43ms, M=25, total time=3mins; 3T: TE=1.43ms, M=31, total time=3mins. Regions of interest were manually drawn using ANALZE software for the substantia nigra with one ROI including the total SN including the pars reticulata (SNT), and a more refined ROI in the laterodorsal SN centered on the pars compacta (ldSN), the dorsolateral putamen (dPUT), medial globus pallidus (Pall), red nuclei (RN) and reference WM (WM).

Results: Patients with PD did not show reduced T₂, T₂* or T₂' values in any of the selected ROIs. This is in contradiction to previous reports at lower field suggesting enhanced transverse relaxivity due to increased paramagnetic iron. Of note, PD patients showed a trend for higher putamenal T₂ values in PD (p<0.1, table).

Individual comparison of relaxation times in the two MSA patients showed variability between the two for the putamen and apparently lower T₂* in the red nuclei.

All values in ms	SNT	ldSN	dPUT	Pall	RN	WM
T ₂ : Controls	24.0 +/-2.6	34.0 +/- 2.8	30.0 +/- 5.0	32.7 +/-10.6	29.3 +/-3.8	61.2 +/-41.2
T ₂ : PD	27.6 +/-5.4	33.7 +/- 2.9	37.1 +/- 6.7	36.5 +/-11.6	28.6 +/-2.6	62.2 +/-45.9
T ₂ : MSA	28.0 +/-5.7	37.7 +/-4.7	33.5 +/- 7.6	26.9	29.8 +/-7.4	56.0
T ₂ *: Controls	18.7 +/-3.4	23.3 +/-4.4	23.3 +/- 5.7	20.9 +/-8.0	19.2 +/-1.8	30.0 +/-4.1
T ₂ *: PD	17.1 +/-1.6	20.6 +/-1.5	25.5 +/-5.0	27.2 +/-10.0	19.1 +/-2.1	26.2 +/-5.0
T ₂ *: MSA	22.8 +/- 3.9	23.5 +/-0.5	59.7 +/-31.9	20.8	15.3 +/-0.4	28.0
T ₂ ': Controls	34.9 +/-16.3	68.3 +/-27.4	68.3 +/-26.5	49.5 +/-14.4	46.4 +/-15.4	109.3 +/-89.0
T ₂ ': PD	36.1 +/-12.9	54.6 +/-7.7	73.9 +/-26.7	50.7 +/-22.2	37.8 +/-12.9	71.4 +/-28.1
T ₂ ': MSA	49.0 +/-7.1	59.4 +/-14.8	90.2 +/-41.7	38.0	50.8 +/-15.9	35.0

Discussion:

Contrary to our hypothesis we were unable to detect alterations in T₂, T₂* or T₂' in the substantia nigra of patients with PD at 7T. Given the high sensitivity of ultrahigh-field for iron, the findings were unexpected. They accord however with a lack of altered FDRI in patients with late-onset disease⁶. Increased iron as seen post mortem and in transcranial sonography¹³ in PD brains may not translate proportionately into enhanced relaxivity as previously assumed. It has been recognised that inferences about the effect of tissue iron on relaxation times is problematic because of two critical assumptions (i) that tissue iron is mainly stored as ferritin and (ii) that iron concentration is the dominant factor governing relaxation behaviour. Both assumptions may be wrong in PD brains. Reduced nigral ferritin levels were reported in PD³. The second assumption is unlikely to be true in brain diseases with altered tissue water. In fact, a MRI study at 11.7T of a genetic rodent model of iron overload concurs well with our findings: no T₂ reduction was observed in the substantia nigra despite histologic evidence of increased ferric iron and ferritin. Histogram analysis in that study suggested a bimodal T₂ distribution with some pixels showing T₂ elevation offsetting the effects from iron increase. T₂ increase was felt to be associated with increased free fluid.¹⁴ Similar vacuolisations may be present in PD brains and in fact are the most likely cause of the observed T₂ increase in the putamen that was also previously noted.⁸

In conclusion, despite increased iron sensitivity of ultra high-field MRI, no increase of iron was detected in the substantia nigra of PD patients. If we do not challenge the compelling post mortem evidence of increased iron, the data support the notion that iron concentrations cannot be directly inferred from relaxation time measurements of diseased tissue even at ultrahigh-field. Direct validation studies on post mortem material is warranted.

References

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