

A New Bioactivated MRI Contrast Agent : Synthesis, Characterization and MR Imaging Studies

Y-M. Wang¹, Y-Z. Su¹, T-L. Cheng², Y-T. Kuo³, T-S. Jaw³, and G-C. Liu³

¹Faculty of Medicinal and Applied Chemistry, Kaohsiung Medical University, Kaohsiung, Taiwan, ²Faculty of Biomedical Science and Environmental Biology, Kaohsiung Medical University, Kaohsiung, Taiwan, ³Department of Medical Imaging, Kaohsiung Medical University Hospital, Kaohsiung, Taiwan

Abstract

A smart contrast agent, the β -glucopyronurate-containing gadolinium(III) complex GdL₁ (L₁ = 1-(2-difluoromethyl-4-(1-(4,7,10-triscarboxymethyl-(1,4,7,10-tetraazacyclodecyl))acetamido)phenyl)- β -D-glucopyronurate) was synthesized and characterized. Relaxometric studies show that the T₁ change percentage in the value of GdL₁ decreases dramatically (51%) in the presence of β -glucuronidase (β -G) and human serum albumin (HSA). A significant signal change percentage enhanced by MR images was observed for GdL₁ solution in the presence of β -G and HSA. The MR images also shows high intensity enhancement in CT26(+ β G) with β -G gene expression but not for the CT26(- β G) without β -G gene expression.

Introduction

MRI offers several advantages over other clinical diagnostic techniques for molecular imaging, including high spatial resolution, non-invasiveness, high anatomical contrast and lack of harmful radiation. However, sensitivity of MRI to depicting small molecule is constrained by the ubiquitous protons in the body, resulting in a high background and lower signal to noise ratio (SNR). Hence, the alternative amplification strategies using smart contrast agents are required to yield a higher sensitivity. Enzyme motif such as β -G, an often-used reporter enzyme in molecular biology, has been explored for smart MR contrast agent. In this study, we characterized an enzymatic contrast agent, which could be bound with HSA or β -G and activated by β -G *in vitro*.

Methods

The spin-lattice relaxation time T₁ measurements were made using a NMR relaxometer operating at 20 MHz and 37.0 ± 0.1 °C (NMR-120 Minispec, Bruker). The percentage changes of T₁ value in these solutions were plotted against the incubation time. CT26 colon carcinoma cells, CT26(+ β G) and CT26(- β G) were used in this study. Both CT26(+ β G) and CT26(- β G) cells were incubated with 1.0 mM GdL₁, washed by PBS buffer 3 times and scanned by 3.0 T MRI.

Results and Discussion

The purity of ligand and Gd(III) complex were performed by HPLC. In Fig. 1, the kinetics of enzyme-catalyzed hydrolysis of GdL₁ were measured, the T₁ value of GdL₁ decreasing about 17 and 51 % after incubation with different concentration β -G (5 μ g/ml and 0.1mg/ml), respectively in HSA. In high β -G concentration (0.1mg/ml), the T₁ decreasing of GdL₁ in the presence of β -G and HSA is significantly higher than that of GdL₁ in the presence of low β -G concentration (5.0 μ g/ml). These results indicated the GdL₁ is a smart contrast agent in the presence of high concentration β -G. The MR images for GdL₁ solution in the presence and absence of β -G in HSA solution were shown in Fig. 2. The MR image results of GdL₁ in the presence of β -G and HSA for 3.0 T MRI indicates that the enhancement (65%) is higher than that of GdL₁ in PBS buffer. In Fig. 3, the *in vitro* MR images of the CT26(+ β G) is significantly higher (16%) than that of CT26(- β G). This image clearly indicates that GdL₁ can be activated by CT26(+ β G) and worth to notice that β -G on the CT26(+ β G) was functionally active *in vitro*.

Conclusion

A new bioactivated MRI contrast agent GdL₁ was synthesized and characterized successfully. T₁ decreasing (51 %) of enzymatic cleavage of GdL₁ in HSA indicates that the HSA and β -G bind to Gd(III) chelate after that the glucopyronurate residue was removed. The MR images of enzymatic experiment shows the signal change percentage of GdL₁ in the presence of β -G and HSA is higher than GdL₁ in PBS buffer. The *in vitro* MR images show a higher intensity enhancement in CT26(+ β G) but not for the CT26(- β G). GdL₁ possesses enzymatic cleavage, as well as higher percentage change in relaxation time and higher signal change percentage of MR images in the presence of β -G and HSA. Therefore, we can conclude that GdL₁ is a potential candidate for a bioactivated MRI contrast agent in tracing gene expression.

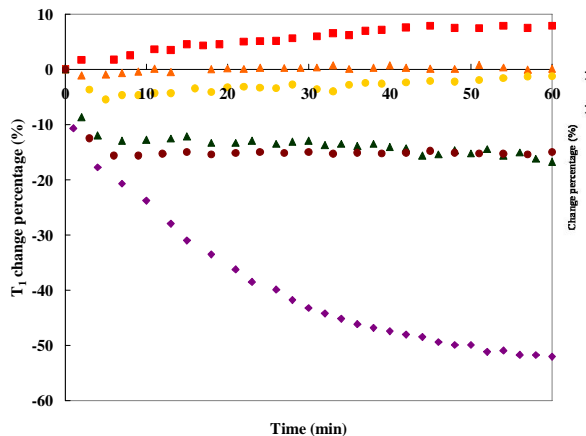


Fig. 1 Change in T₁(%) of enzyme-catalyzed hydrolysis of GdL₁ measured at 20 MHz, 37.0 ± 0.1 °C. (■) 0.5 mM GdL₁ and 0.1mg/ml β -G in 100 mM PBS buffer (pH=7.4 ± 0.1); (▲) 0.5 mM GdL₁ in 100 mM PBS buffer; (●) 0.5 mM GdL₁ and 5.0 μ g/ml β -G in PBS buffer; (●) 0.5 mM GdL₁ and 0.5 mM HSA in 100mM PBS buffer; (▲) 0.5 mM GdL₁, 0.5 mM HSA

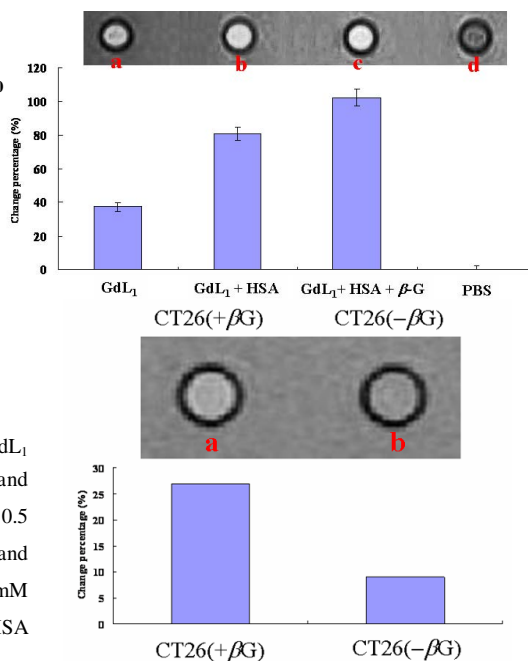


Fig. 2 Representative T₁-weighted (TR/TE 100/2.4 ms) MR images of solutions and the signal change percentages in test tubes at 3.0 T MR scanner. (a) 0.5mM GdL₁ in PBS buffer solution; (b) 0.5mM GdL₁ and 0.5mM HSA in PBS buffer solution; (c) 0.5mM GdL₁, 0.1mg/ml β -G and 0.5mM HSA in PBS buffer solution; (d) PBS buffer solution.

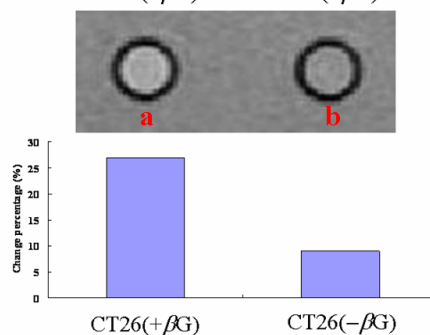


Fig. 3 *In vitro* MR images (TR/TE 150/5.8 ms) of CT26(+ β G), CT26(- β G) cells and the signal change percentages in the test tubes at 3.0 T MR scanner. (a) CT26(+ β G) cells incubate with GdL₁; (b) CT26(- β G) cells incubate with GdL₁.

References

- [1]. Moats, R. A.; Fraser, S. E.; Meade, T. J. *Angew. Chem. Int. Ed. Engl.* **1997**, *36*, 725-728.
- [2]. Duimstra, J. A., Femia, F. J., Meade, T. J. *J. Am. Chem. Soc.* **2005**, *127*, 12847-12855.
- [3]. Ou, M. H., Tu, C. H., Tsai, S. C., Lu, W. K., Lee, W. T., Liu, G. C., and Wang, Y. M. *Inorg. Chem.* **2006**, *45*, 244-254.
- [4]. Wang, Y. M., Li, C. R., Huang, Y. C., Ou, M. H., and Liu, G. C. *Inorg. Chem.* **2005**, *44*, 382-392.
- [5]. Chang, Y.-T.; Cheng, C.-M.; Su, Y.-Z.; Lee, W.-T.; Hsu, J.-S.; Liu, G.-C.; Cheng, T.-L.; Wang, Y.-M. *Bioconjugate Chem.* (2007)(in press).