## Changes of Proton T<sub>1</sub> and T<sub>2</sub> Relaxation Times of Cerebral Metabolites Induced by Repeated Manganese Treatments in

Rat

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**Introduction** Chronic overexposure to manganese leads to selective deposition of the metal in the globus pallidus (GP) and clinical symptoms that resemble those found in Parkinson's disease<sup>[1]</sup>. Recently in vivo <sup>1</sup>H MRS was applied to study the effects of Mn neurotoxicity on cerebral metabolism in human and non-human primate<sup>[2-3]</sup> exposed to manganese chronically. The results showed that, compared to control, the patients had mildly reduced N-acetylaspartate (NAA)/creatine (Cr) ratio in the basal ganglia<sup>[2]</sup>, and the manganese-treated non-human primates had significantly decreased NAA/Cr ratio in the frontal white matter<sup>[3]</sup>. Yet Mn<sup>2+</sup> is a paramagnetic ion that can enter neurons via voltage-gated Ca<sup>2+</sup> channels, and it may affect the T<sub>1</sub> and T<sub>2</sub> relaxation times of both intracellular and extracellular metabolites, thus affecting their quantification. In fact, it has been demonstrated in phantom experiments that the presence of Mn<sup>2+</sup> reduces the T<sub>1</sub> of NAA and Cr methyl resonance<sup>[4]</sup>. In this study, Mn<sup>2+</sup>-induced changes in the T<sub>1</sub> and T<sub>2</sub> of water, NAA and Cr were measured in vivo at 4.7 T in the GP of rats subjected to repeated manganese exposures.

**Materials and Methods** Male SD rats (180-250 g) received daily i.p. injection of MnCl<sub>2</sub> (120 mmol/L, 3 ml/kg) for five consecutive days. T<sub>1</sub>-weighted imaging and in vivo <sup>1</sup>H MRS were performed on isoflurane (1.5-2.0%, in 70:30 N<sub>2</sub>O /O<sub>2</sub>) anesthetized rats before injection, on 2d before the 2nd injection and 1d after the last injection (i.e., on 6d) on a 4.7T/30cm Bruker Biospec scanner. A 12-cm diameter Helmholtz volume coil was used for excitation and a 2.5-cm diameter surface coil for reception. A PRESS sequence was used to acquire <sup>1</sup>H spectra from the GP with a voxel size of 2.5 mm×2.5 mm (Fig. 1). Spectral data obtained in saturation-recovery (SR) experiments were used to calculate T<sub>1</sub> of metabolites, and the changes of metabolite peak intensities with total TE was used to calculate T<sub>2</sub>. the T<sub>1</sub> of water and metabolites was measured with TE 136 ms and 9 TR values ranging from 0.5 to 10 s, and the T<sub>2</sub> with TR=2 s and 6 TE values ranging from 30 to 272 ms. Statistical analysis was performed with one-way ANOVA followed by post hoc Tukey's test.

**Results** A <sup>1</sup>H spectrum acquired from the GP of a  $Mn^{2+}$ -treated rat is shown in Fig. 1. Compared to control, the T<sub>1</sub> and T<sub>2</sub> of NAA and Cr methyl resonance at 2.02 ppm and 3.0 ppm had insignificant decreases after the 1<sup>st</sup> treatment, while the T<sub>1</sub> and T<sub>2</sub> of water reduced significantly by about 26±5% and 6±1%, respectively. After 5  $Mn^{2+}$ -treatments, the T<sub>1</sub> of NAA, Cr and water reduced significantly by 14±16%, 35±24% and 41±7%, and the T<sub>2</sub> by 26±15%, 12±10% and 12±1%, respectively.

**Discussion** The  $T_1$  and  $T_2$  of water decreased significantly already after the first treatment and further reduced after 5 treatments, suggesting that the concentration of  $Mn^{2+}$  in the GP increased as the rats had more  $Mn^{2+}$  treatments, agreeing with the results of Hazell et al<sup>[5]</sup>. It was estimated that at the injection dose used in this study, the  $Mn^{2+}$  concentrations in the GP were about 1.6 µg/g and 7.3 µg/g, respectively, after one and five treatments. The  $T_1$  and  $T_2$  of NAA and Cr had a trend to decrease after the first treatment and such reductions became statistically significant after 5 treatments, suggesting that the  $T_1$  and  $T_2$  of metabolite also depend on the  $Mn^{2+}$  concentration in the brain region where they are measured. Therefore, when in vivo <sup>1</sup>H MRS is used to assess  $Mn^{2+}$ -induced cerebral metabolic changes, the effects of relaxation time changes induced by the presence of  $Mn^{2+}$  should be taken into account in metabolite quantification, and this can be done by optimizing the spectroscopic acquisition parameters<sup>[4]</sup>. For example, using the  $T_1$  and  $T_2$  relaxation data obtained in this study and assuming the concentrations of metabolites do not changes, relaxation-induced signal intensity changes for NAA and Cr methyl resonance after 5  $Mn^{2+}$  treatments would be 1.1% and 0.9%, respectively, if TR of 6 s and TE of 20 ms are used, and the changes will increase to 4.4% and 11.7%, respectively, if TR of 1 s and TE of 136 ms are used.

Acknowledgements Supported by grants 10234070, 30370419 and 30400136 from Natural Science Foundation of China. **References** [1]Kim JW, et al, J Korean Med Sci, 1998 13(4):437-9. [2]Kim EA, et al, Neurotoxicology, 2006, In press. [3]Guilarte TR, et al, Toxicol Sci, 2006 94(2):351-8. [4]Madsen KS, Proc. Intl. Soc. Mag Reson Med, 2006:1489. [5]Hazell AS, et al, Neurosci Lett, 2006 396(3):167-71.



**Figure 2**:  $T_1$  (A) and  $T_2$  (B) relaxation times of water and metabolites in the globus pallidus of control rats (con) and rats treated with MnCl<sub>2</sub> for 1 day (1d) and 5 days (5d). \*p<0.05, compared to control, # p<0.05, compared to 1d.