

## Analysis of quantitative MT using principal component analysis

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### Introduction

Quantitative magnetization transfer (MT) is based on a two-pool model of MT [1], and yields several indices, reflecting properties of the macromolecular pool. The relationship between these indices, and the amount of overlapping information they contain, is not clear, and the interpretation of white matter changes measured by quantitative MT is not always straight-forward. Principal component analysis (PCA) can be used both to reduce the dimensionality of data by reorienting them so that the first few dimensions account for as much of the variance as possible, and to gain insight into patterns of association between variables. Here we use PCA to aid interpretation of quantitative MT parameters.

### Methods

This work is based on the model of quantitative MT developed by Ramani et al. [2]. In Ramani's model, the MT pulse is replaced by continuous wave irradiation with the same mean square amplitude (continuous wave power equivalent approximation). Seven quantities can be derived from the model:  $RM_0^A$ ,  $f$ ,  $T_2^A$ ,  $T_2^B$ ,  $T_1^A$ ,  $T_1^B$  and  $gM_0^A$ . Here A and B label the liquid and the macromolecular pools, respectively,  $R$  is the exchange rate,  $f$  represents the bound proton fraction,  $M_0^A$  is the longitudinal magnetization of pool A at the equilibrium, and  $gM_0^A$  is the signal measured in the absence of MT saturation [2]. The seven parameters can be uniquely determined only if  $T_1$  of the system is ascertained independently, thus expressing  $T_1^A$  as a function of the other parameters [1].  $T_1^B$  is generally fixed to be 1 s [1-3]. Eight subjects (M/F=2/6, median age = 52.5 yrs, range=18-62 yrs) were scanned on a 3.0 T system using a 3D MT-weighted fast SPGR sequence (A) (TR/TE=25.8/2.7 ms, flip angle=5°, Gaussian MT pulses, duration=14.6 ms), which collects 11 volumes with various combinations of amplitude ( $\omega$ ) and offset frequency ( $\Delta$ ) of the MT pulse, optimised according to [4]. In addition to the MT data, (B) two 3D fast recovery FSE (TR/TE=300/24.3 ms, ETL=12, flip angles=60° and 120°, matrix=64x64), (C) 2 fast 3D SPGRs (TR/TE1/TE2=25.8/2.7/5.4 ms, flip angle 5°) and (D) 3 fast 3D SPGRs (TR/TE=6.0/2.8 ms, flip angles 15°, 7°, and 3°, #slices=88, slice thickness = 2 mm) were obtained. Images were collected in the coronal plane. Acquisition matrix (256x96), FoV (24x18 cm<sup>2</sup>), number of slices (34) and slice thickness (5 mm) were the same for all sequences unless otherwise specified. After image co-registration,  $B_1$  maps were obtained from sequence B using the double angle method [5] and  $B_0$  maps were obtained from sequence C [6].  $T_1$  maps were calculated from sequence D as described in [3]. Ramani's model was then fitted to the data as in [3], correcting  $\omega$  and  $\Delta$  based on the field maps. The 15° flip angle SPGR from sequence D was segmented into white matter, grey matter and CSF using SPM99 ([www.fil.ion.ucl.ac.uk/spm/](http://www.fil.ion.ucl.ac.uk/spm/)). White and grey matter segments were then combined to produce a brain mask to remove CSF. The brain-extracted images were processed using a local-developed algorithm for voxel-based PCA implemented in Matlab (The Mathworks, MA, USA), producing maps of the principal components (PCs), eigenvalues (i.e. the variances associated with each principal component), and component loadings (i.e. the correlation matrix of the principal component scores with the original data) for the system. Five variables were entered into the PCA:  $f$ ,  $RM_0^A$ ,  $T_1^A$ ,  $T_2^A$ , and  $T_2^B$ .

### Results

Maps of the PCs averaged across subjects after normalisation are shown in Fig 1. The eigenvalues for the system were, in descending order: [2.26,1.01,0.86,0.51,0.36]. Using the scree plot [7] we retained the first 3 components, which explain 82.6% of the total variance. The loadings for the first three components are reported in Table 1.

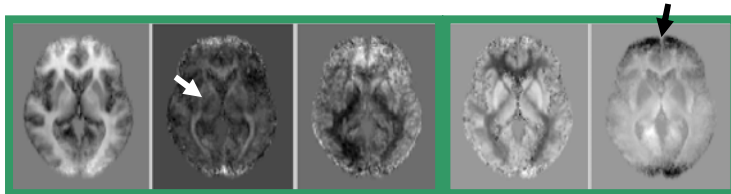


Fig 1. Average PC maps in descending order.

### Discussion

The principal component loadings suggest that  $T_1^A$  and  $T_2^A$  are highly correlated. This is not surprising as most of the variance is likely to come from grey-to-white matter contrast. Similarly,  $f$  appears to be correlated with the first PCA component.  $T_1$  is known to be affected by the presence of myelin in white matter [8], and  $f$  is believed to reflect the myelin content. Consequently, some (inverse) correlation between  $f$  and  $T_1^A$  is to be expected and has been previously observed.  $T_2^B$  appears to be independent of the other variables and almost entirely summarized by the second component, consistent with the results of Sled et al. [9], who interpreted it as a measure of the rigidity of the structure. The second component contains some interesting features such as contrast between white matter structures running within the axial plane and those running in the superior-inferior direction (white arrow). The third component is mainly represented by  $RM_0^A$ , with some weighting from  $f$  (thus suggesting that  $f$  conveys more information than  $T_1$  alone). A limitation of this type of analysis is the inability to separate variance components genuinely related to the underlying tissue characteristics from those due to image artefacts (see black arrow, pointing at artifact due to FoV edge, as the acquisition was coronal, and low frequency variation on PCs 1 and 2, which might reflect some spatial inhomogeneity of the input maps). A rotated solution might help in removing this ambiguity. It is also interesting to note that the fourth component is characterised by some asymmetry between left and right, which might be related to the  $B_1^{(+)}/B_1^{(-)}$  imbalance occurring at high field strength [10].

### References

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Table 1. PC loadings.

Absolute values >0.5 are highlighted.

	PC1	PC2	PC3
$f$	<b>0.77</b>	0.11	-0.43
$RM_0^A$	<b>0.54</b>	-0.27	<b>0.77</b>
$T_1^A$	<b>-0.87</b>	-0.07	0.02
$T_2^A$	<b>-0.76</b>	-0.29	0.00
$T_2^B$	0.23	<b>-0.91</b>	-0.28