

Enhanced Diffusion Weighting Generated by Selective Composite Adiabatic Pulses

Z. Sun^{1,2} and R. Bartha^{1,3}

¹Imaging Research Laboratories, Robarts Research Institute, London, Ontario, Canada, ²Davis Heart & Lung Research Institute, The Ohio State University, Columbus, Ohio, United States, ³Department of Diagnostic Radiology, University of Western Ontario, London, Ontario, Canada

Introduction: Frequency selective adiabatic full passage (AFP) pulses have been used in imaging and localized spectroscopy with high tolerance to B_1 inhomogeneity [1-3]. Previous research demonstrated that selective chirp AFP pulses generate spin phase that is quadratically proportional to the local spin resonance frequency [4]. This nonlinear phase variation across the selected slice was compensated using two identical AFP pulses [1, 2, 4] to generate a robust spin-echo. Conversely, the nonlinear phase dispersion can also be increased using alternate frequency sweep (AFS) composite AFP pulse trains [5] incorporated into the localization by adiabatic selective refocusing (LASER) sequence [2] to increase diffusion weighting. However, a preliminary study using the LASER sequence demonstrated that the apparent diffusion coefficient (D) could not be measured accurately in nickel(II) doped water phantoms with short T_2 , due to the small b -value produced by the selective AFP pulses alone. In the current study, a pair of pulsed gradients was incorporated into the LASER pulse sequence to increase the associated b -value. The purpose of this study was to determine whether selective adiabatic composite pulse trains could produce enhanced diffusion weighting and be used in combination with pulsed gradients to measure $^1\text{H}_2\text{O}$ diffusion in phantoms spanning of a wide range of T_2 values. An expression for the b -value generated by the LASER sequence was derived using the Bloch-Torrey equation following the method previously described [6]. A conventional pulsed gradient spin-echo (PGSE) sequence was employed as the gold standard for diffusion measurements in all phantoms.

Methods: Frequency selective adiabatic composite pulse trains consisting of 1, 3, 5, or 7 hyperbolic secant (HS1, R10, [2]) AFP-AFS pulses were used in the LASER sequence (Fig. 1). Pulsed gradients were added symmetrically on both sides of the refocusing selective adiabatic composite pulse train separated by a diffusion time (Δ) (Fig. 1). Six phantoms (Ph-1 to Ph-6) were studied on a 4T Varian whole body MRI with a Siemens Sonata gradient coil using a hybrid birdcage transmit/receive radio frequency coil (7.7 cm ID). Phantoms consisted of 2.8 cm diameter (50 ml) plastic tubes containing a mixture of 10 μm ORGASOL polymer beads and 2 mM Gd-DTPA dissolved in 5% agar (Ph-1), and nickel(II) ammonium sulphate hexahydrate doped (0.8 - 56.3 mM) water solutions (Ph-2 to Ph-6). The transverse relaxation time constant T_2 was measured from a single 5 mm transverse slice in each phantom using a spin-echo (SE) sequence (TE = 40 - 60 ms in steps of 5 ms). The repetition time (TR) was varied (2 - 4 s) for each phantom to minimize T_1 saturation. The diffusion coefficient (D) of each phantom was measured with both PGSE and LASER by varying G_d while fixing Δ and TE for a particular phantom (FOV = 4 cm, matrix = 64 x 64, $G_d = 0 - 3.5$ G/cm in step of 0.5 G/cm, TE = 62 - 72 ms, TR = 2 - 4 s, $\delta = 10$ ms, $\Delta = 35 - 40$ ms, slice thickness = 5 mm). Ph-1 and Ph-4 were measured using 1, 3, 5, and 7 AFP pulses in the AFP-AFS pulse train, while other phantoms were measured with 1 and 5 AFP pulses only. T_2 time constants were calculated from the linear regression of the natural logarithm (ln) of the image signal intensity (SI) of the SE images with TE. The D -value for each phantom was calculated by the linear regression of ln(SI) (PGSE or LASER image signal intensity) with the b -value. The b -values for the PGSE and LASER sequences were calculated using Stejskal-Tanner equation [6] and self-derived equations, respectively.

Results: Typical diffusion weighted images are shown in Fig. 2. Increased image intensity was observed in the bead phantom (Ph-1) while decreased image intensity was observed in the nickel(II) doped water phantom (Ph-4) as the number of pulses in the AFP-AFS pulse train increased (while maintaining a constant TE (72 ms) and pulsed gradients (3.5 G/cm)). In the bead phantom (Ph-1), there is a clear increase in signal intensity associated with the use of more refocusing pulses in the AFP-AFS pulse train (i.e. 7 AFP line is above the 1 AFP line), while the converse is true for Ph-4. The T_2 time constants and D -coefficients measured for each phantom are summarized in Table 1.

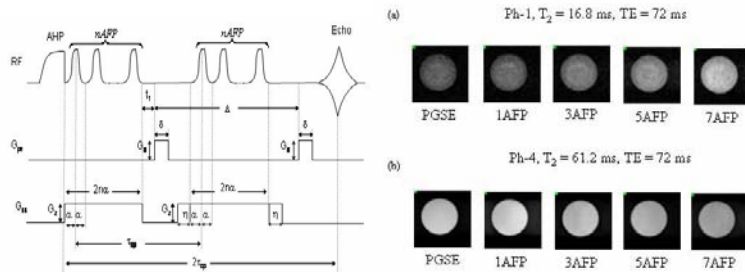


Figure 1

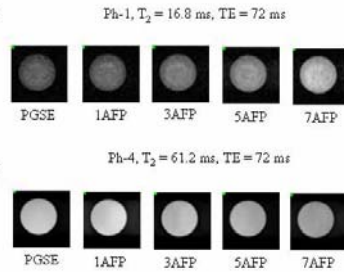


Figure 2

Discussion: A diffusion-weighted imaging sequence incorporating LASER was successfully implemented and used to measure the apparent diffusion coefficient in phantoms exhibiting a wide range of T_2 relaxation time constants. In comparison to the conventional PGSE sequence, the diffusion-weighted LASER sequence can generate unique diffusion contrast at the same pulsed gradient amplitude and TE due to the nonlinear phase dispersion generated and the spin-locking effect associated with the AFP-AFS pulse train. The combination of T_{2p} , T_2 , and diffusion weighting produced by the AFP-AFS-LASER sequence has demonstrated opposite effects on image signal intensity depending on the sample microscopic susceptibility characteristics [7, 8]. This important feature of the AFP-AFS-LASER sequence in comparison to the conventional PGSE sequence may be used to identify the presence of magnetic perturbors that have different magnetic susceptibility compared to surrounding material (i.e. tissue).

Acknowledgments and References: Funding provided by NIH (R01-EB001852), CIHR (MME 15594), and the Ivey-BMO Financial Group Award. The authors thank Dr. Michael Garwood for providing the LASER pulse sequence. [1] S. Conolly, G. Glover, D. Nishimura, A. Macovski, Magn. Reson. Med. 18 (1991) 28-38. [2] M. Garwood, L. DelaBarre, J. Magn. Reson. 153 (2001) 155-157. [3] R. Bartha, S. Michaeli, H. Merkle, et al, Magn. Reson. Med. 47 (2002) 742-750. [4] D. Kunz, Magn. Reson. Med. 3 (1986) 377-384. [5] Z. Sun, R., Bartha, In: Proceedings of the 14th Annual Meeting of ISMRM, Seattle, 2006. (abstract 2999). [6] E.O. Stejskal, J.E. Tanner, J. Chem. Phys. 42 (1965) 288-292. [7] S. Michaeli, H. Grohn, O. Grohn, et al, Magn. Reson. Med. 53 (2005) 823-829. [8] S. Nikolova, C.V. Bowen, R. Bartha, J. Magn. Reson. 181 (2006) 35-44.

Table 1. The measured D -coefficients and T_2 time constants

Phantom	T_2 (ms)	PGSE	1AFP	3AFP	5AFP	7AFP
Ph-1	16.7±0.2	0.75±0.04	0.74±0.02	0.30±0.02	0.26±0.02	0.94±0.02
Ph-2	21.2±0.1	2.34±0.08	2.19±0.08		2.31±0.04	
Ph-3	43.3±0.1	2.32±0.02	2.23±0.08		2.46±0.04	
Ph-4	61.2±0.4	2.30±0.02	2.24±0.02	2.33±0.02	2.33±0.02	2.26±0.04
Ph-5	131.1±0.7	2.40±0.04	2.30±0.02		2.40±0.04	
Ph-6	555.6±3.1	2.28±0.02	2.16±0.02		2.33±0.04	